

**Inferring reciprocal evolutionary histories in associated species
of plants and insects in two European pollination systems**



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par

María Anahí Espíndola

Acceptée sur proposition du jury:



Dr. Nadir Alvarez, co-directeur de thèse
Prof. Martine Rahier, co-directeur de thèse
Prof. Bryan C. Carstens (Louisiana State University, USA), rapporteur
Dr. Laurence Després (Université de Grenoble, France), rapporteur
Dr. Marc Gibernau (CNRS Toulouse, France), rapporteur
Dr. Felix Gugerli (WSL, Birmensdorf, Zürich, Suisse), rapporteur
Prof. Philippe Küpfer (Université de Neuchâtel, Suisse), rapporteur

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Inferring reciprocal evolutionary histories in associated species of plants and insects in two European pollination systems

María Anahí Espíndola

UNIVERSITE DE NEUCHATEL

FACULTE DES SCIENCES

La Faculté des sciences de l'Université de Neuchâtel,
sur le rapport des membres du jury

Dr Nadir Alvarez (co-directeur de thèse, Université de Neuchâtel),
Prof. Martine Rahier (co-directrice de thèse, Université de Neuchâtel),
Prof. Bryan C. Carstens (Louisiana State University USA),
Prof. Laurence Després (Université de Grenoble F),
Dr Marc Gibernau (CNRS Toulouse F),
Dr Felix Gugerli (WSL, Birmensdorf CH),
Prof. Philippe Küpfer (Université de Neuchâtel)

autorise l'impression de la présente thèse.

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Le doyen :
P. Kropf

« Toy with different perspectives. Look for the unusual. Try consciously to innovate. Train yourself to imagine new schemes and innovative ways to fit the pieces together. Seek the joy of discovery. Always test your new thoughts against the facts, of course, in rigorous, cold-blooded, unemotional scientific manner. But play the great game of the visionary and the innovator as well.»

J. E. Oliver

« Gracias a la vida, que me ha dado tanto... »

V. Parra

A Cristina y Ricardo, mis padres.

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Keywords: specific interactions, coevolution, *Arum maculatum*, *Trollius europaeus*, *Chiastocheta*, Psychodidae, antagonism, mutualism, ecological niche model, comparative phylogeography, biogeography, diversification, hindcasting methods, arctic-alpine species, Size-Advantage Model, taxonomy, statistical phylogeography

Summary

Coevolution is defined as reciprocal evolutionary changes that might arise at any spatiotemporal scale. Despite every organism on Earth undergoes coevolutionary interactions, cases of one-to-one specific relationships are generally rare. However, because of the reduced number of interacting species they concern, these species-specific associations are interesting to evolutionary biologists because they allow testing hypotheses in simple frameworks.

Despite the history and evolution of coevolutionary interactions have been studied in several cases in the last decade, this topic remains difficult to fully circumscribe because of the multiplicity of factors that affect one or the other species concerned. Moreover, a lot is known about coevolution at a small scale, but little has been done at larger and more integrative scales spanning wider spatiotemporal ranges.

Phylogeography is a young area of biology that allows understanding the distribution of lineages in space and time. Despite that the idea of parallelly studying the history of species involved in specific interactions appears simple, this has rarely been done until now probably because of the technical efforts this would represent.

From a theoretical point of view, we could propose that in specific interactions, because of the dependence between the partners involved, we should observe some phylogeographic pattern associated to the type of interaction studied. In this way, while partners of mutualistic interactions should present similar postglacial histories, this should not be true for those associated by antagonistic relationships.

In this thesis, we exploit different techniques and approaches to test this general hypothesis. The final aim of this study is thus to understand if it is possible to identify a pattern of comparative phylogeography in relation to the type of interaction, using as case-studies two specific and obligate European interactions: the antagonistic relationship established between *Arum maculatum* L. (Araceae) and its Psychodid (Diptera) pollinating flies, and the nursery pollination mutualism involving *Trollius europaeus* L. (Ranunculaceae) and the *Chiastocheta* (Diptera: Anthomyiidae) species complex.

Before testing our comparative phylogeographic hypotheses and because studying the phylogeography of interactions requires a wide knowledge of the environmental, taxonomic and historical frameworks in which these ecological relationships arose, it was first needed to clearly delimitate the identity of species, their distribution and the environmental factors

influencing their survival to finally understand their comparative history. We thus took advantage of the potentialities that interdisciplinary approaches provide, applying molecular taxonomy, biological and evolutionary methods, biogeographic inferences, ecological niche models and hindcasting techniques, as well as classical and recently-developed phylogeographic analyses.

Our results indicate that the phylogeographic patterns of these specific and obligate antagonistic and mutualistic relationships appear to be related to the type of interaction. Antagonistic partners presented incongruent phylogeographic patterns, what can be notably explained by differences in their life-history traits. Species involved in mutualistic interactions partly showed congruent phylogeographic patterns (particularly in the cases of *T. europaeus* and *C. dentifera*). Flies interacting with *T. europaeus* appear moreover to present different histories, regardless of their important ecological similarities.

These results demonstrate that the systems studied appear to be far more complex than initially supposed, with crossed effects of environmental and historical features on the dynamics of the interaction. Because of the high complexity and interdependency of factors affecting one or the other partner, performing investigations in an interdisciplinary framework appears indispensable to disentangle the dynamics of interactions.

Mots-clés: interactions spécifiques, coévolution, *Arum maculatum*, *Trollius europaeus*, *Chiastocheta*, Psychodidae, antagonisme, mutualisme, modèle de niche écologique, phylogéographie comparée, biogéographie, diversification, méthodes de prédictions de distributions passées, espèce arctique-alpine, modèle d'avantage de taille, taxonomie, phylogéographie statistique

Résumé

La coévolution est définie comme la série de changements réciproques apparaissant entre des organismes. Malgré le fait que tous les organismes suivent des processus de coévolution, les cas des relations spécifiques un-un sont relativement rares. Ces types d'interactions spécifiques sont par contre très intéressants pour les évolutionnistes, notamment à cause du nombre réduit d'organismes qu'elles comprennent, ce qui permet de tester des hypothèses évolutives dans un cadre simple.

Malgré le fait que l'histoire et l'évolution des interactions coévolutives sont des processus assez explorés, le thème reste difficile à étudier spécialement à cause du grand nombre de facteurs influençant les espèces concernées. D'un autre côté, beaucoup est à ce jour connu à des petites échelles spatio-temporelles, alors que très peu a été fait à des dimensions plus larges et plus intégrées.

Le jeune domaine de la phylogéographie s'intéresse à la compréhension de la distribution des lignées génétiques dans l'espace et dans le temps. Malgré que l'idée de l'étude parallèle des histoires phylogéographiques de plusieurs espèces est un concept simple, il n'a été exploré que pour quelques cas isolés.

D'un point de vue théorique, nous pouvons proposer que dans des interactions spécifiques, nous devrions attendre à voir des patrons phylogéographiques liés au type d'interaction étudiée, spécialement à cause de l'interdépendance des espèces concernées. Alors que des partenaires mutualistes devraient présenter des histoires démographiques similaires, ceci ne devrait pas être le cas des espèces liées par des interactions de type antagoniste.

Le but ultime de ce travail est alors de comprendre s'il est possible d'identifier des patrons phylogéographiques spécifiques aux types d'interactions. Afin de répondre à cette question, nous testons nos hypothèses sur deux interactions européennes spécifiques et obligées: l'antagonisme formé par *Arum maculatum* L. (Aracée) et ses pollinisateurs de la famille Psychodidae (Diptères) ; le mutualisme *de crèche* présent entre *Trollius europaeus* L. (Ranunculacée) et le complexe d'espèces du genre *Chiastocheta* (Diptère: Anthomyiidae).

Comme avant de tester nos hypothèses phylogéographiques, il est nécessaire de posséder une connaissance approfondie des cadres environnemental, taxonomique et historique dans lesquels les interactions sont apparues et se développent, nous avons

d'abord défini les limites des espèces en relation, circonscrit leur distribution et compris les facteurs environnementaux influençant la survie des entités, pour ensuite inférer leurs histoires évolutives. Nous avons profité des potentialités que des approches interdisciplinaires fournissent, appliquant des méthodes de taxonomie moléculaire, des approches biologiques et évolutives, des inférences biogéographiques, des méthodes de modélisation de niche écologique et des prédictions des distributions passées, ainsi que des analyses phylogéographiques classiques et modernes.

Nos résultats indiquent que les patrons phylogéographiques des espèces en interaction sont liés aux types de relations étudiées. Ceux des espèces antagonistes sont incongruents et peuvent être expliqués par des différences dans les traits d'histoire de vie des espèces. Les espèces mutualistes ont des patrons phylogéographiques partiellement congruents (comme le cas de *T. europaeus* et *C. dentifera*). Le complexe d'espèces de mouches interagissant avec *T. europaeus* présenterait alors des histoires différentes, malgré ses similarités écologiques importantes.

Nos résultats démontrent que ces systèmes sont beaucoup plus complexes qu'initialement supposés, probablement à cause des effets croisés entre environnement et histoire. A cause de cette complexité et de l'interdépendance des facteurs affectant l'un ou l'autre partenaire, des études interdisciplinaires apparaissent alors comme extrêmement appropriées lorsque notre but est de comprendre des interactions.

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General Introduction

1. Coevolution

Since the formal definition of the coevolutionary theory in 1964 (Ehrlich and Raven 1964) numerous studies have demonstrated that coevolutionary processes shape the Earth's biodiversity (Thompson 2009). From the evolution of cellular organelles (Margulis 1970) to the main role played by interacting species in the maintenance of complex environments (e.g., fruit production in the cases of fig and fig-wasps (Shanahan, So et al. 2001); ecosystem survival in Southern temperate forests (Aizen and Ezcurra 1998); the effect of corals in maintaining reef environments (Thompson 2009)), coevolutionary processes appear to have been fundamental for the development of diversity of life.

Coevolution is defined as the reciprocal evolutionary change in interacting species (Thompson 1994) and is generally detected by the presence of coevolved traits appearing between the interacting organisms (Lunau 2004). The theory was first described based on observations of closely related plants which present also closely related specific herbivores (Ehrlich and Raven 1964). It was then surprising that related plants presented specific secondary compounds which helped them to be protected against herbivory, and that herbivores feeding on those plants were also closely related and appeared specifically resistant to the plants' toxins. In order to present an explanation of these unexpected congruences it was proposed that a "chemical escalation" of plant chemical defenses existed, which would have created a way for the plant to escape herbivory, but which would also have liberated new "empty" ecological niches, which could be occupied by "new" resistant herbivores. If the process was repeated several times, every physiological change would have the potential to delimitate a new taxon in both the plant and the herbivore, what was described as the "escape and radiate" process (Thompson 1994).

In fact, it is not random that the theory was first described for plants and insects because in these two groups coevolutionary processes are extremely frequent. Interactions between plants and insects are not only abundant (i.e., at least 400.000 species of herbivorous insects have been described up to now, Mitter, Farrell et al. 1991), but they appear to have

been happening for hundred million years (Farrell, Mitter et al. 1992). Parallel plant and insect fossil analyses have thus suggested that the increase in the diversification of angiosperms in the mid-Cretaceous was almost simultaneous to radiations happening in groups of insects specialized on pollination as Apoidea or Lepidoptera (Grimaldi 1999). As mentioned above, coevolution was initially “discovered” while investigating herbivory relationships, however it is not only restricted to it, and diversification in angiosperms is currently proposed to be especially a consequence of adaptations related to another type of interaction: zoophilous (animal-mediated) pollination (Crepet 1996). It has been demonstrated that mediated pollination directly influenced the rate of diversification of plants through its effect on floral innovations (Crepet 2000).

Considering the tightness of the process, different types of coevolutionary interactions have been proposed, with a separation along a continuum between one-to-one or specialized interactions (*i.e.*, occurring between a reduced number of species) and diffuse coevolution or generalism (*i.e.*, more than two interacting species) (*sensu* Lunau 2004). Another classification concerns the benefit they provide to the interacting species. While a mutualism is defined as an interaction in which each of the species obtains a net benefit through the “exchange of commodities in a *biological market*” (Bronstein, Dieckmann et al. 2004), in antagonistic interactions one species increases its own fitness by diminishing the other(s)’(s). Despite that these two types of interactions can be either specialized or generalist, it has been observed that most interactions involve far more than two species (Ollerton, Killick et al. 2007). However, in spite of being rare, specific interactions represent extraordinary natural laboratories, in which the forces of coevolution are the most evident and thus the easiest to observe.

Specialization is thought to have arisen from generalist interactions (Futuyma and Moreno 1988). It has been proposed that specialization increases fitness in terms of a raise of the benefit obtained from the interaction thanks to an optimization of the exploitation of the resource, or through the avoidance of interspecific competition (for a review see Pellmyr

2002). However, it has also been shown that not all cases of generalism are ancestral and that under certain situations specific interactions are not energetically positive, what appears to force transitions from specificity to generalism (Tripp and Manos 2008). The potential and capabilities to do this shift appear however to be species-dependent, and while some specific taxa have been able to rapidly change resources (e.g., *Vestiarina coccinea*, Smith, Freed et al. 1995), others experience fitness reductions after environmental changes induced the absence of their specific partners (e.g., long-term lack of seed production in absence of pollinators, Aizen and Feinsinger 1994).

Despite that these ideas have been around for a while, the number of studies focusing on this topic is limited (Tripp & Manos, 2008). One explanation for the small amount of information available is that most studies addressing questions of adaptive evolution concern changes happening (or having happened) at large temporal scales (e.g., million years). Much less has been done for understanding the stability and coevolutionary aspects of the history of species at shorter temporal scales (e.g., thousand years). At this scale, however, it could be possible to see the effect of differential demographic and distributional changes of the species involved, and their influence on the maintenance or loss of the interaction, especially in relation with major environmental changes.

2. The coevolutionary process in an ecological niche and spatial framework

Ecological niches have been initially defined by Hutchinson (1957) and include all conditions that allow a species to survive. These conditions include both abiotic (e.g., climatic, topographic, edaphic) and biotic (*i.e.*, inter-specific interactions) factors. The ecological niche of a species thus defines not only its “role” in the community in which it is included; it also has a direct effect on the capability of survival of the species at a given location. A species will thus survive only if conditions defined by its ecological niche are realized at specific locations.

Because environmental conditions are included in the definition of the ecological niche of a species, any variation in the abiotic factors has the potential to directly affect the distribution range of organisms. Accordingly, environmental variations have been demonstrated to influence the distributions of species. This fact has been shown to apply both for past (e.g., Cheddadi, Vendramin et al. 2006) as well as for current climatic changes (e.g., Walther, Post et al. 2002).

Similarly, the same effect is expected for the biotic component of the ecological niches, especially if the interaction clearly increases the fitness of the organism (Pellissier, Alvarez et al. in press). In a case where the presence of the associated species defines the survival of another species, the probability of survival and maintenance of the dependent species will be tightly related with the presence of its associated taxa. Consequently, distribution ranges of interacting species must overlap, at least partially, and if this is not the case, another species should fulfill the role of the one absent; otherwise the organism will not be able to survive at a given location.

The niches of all organisms are influenced both by biotic and abiotic factors. However, in species participating in obligate and specific interactions, the effect of the biotic component in the definition of the realized niche is primordial, since a reduced number of factors (*i.e.*, presence or absence of particular species) can drastically define the output of the equation allowing survival. In these species, because the biotic dependence relies on a restricted number of partners, their absence can drastically endanger the maintenance of the dependent species.

3. Phylogeography, or the effect of abiotic factors on the history of species

In comparison to other areas of biology, phylogeography is a relatively new domain in evolutionary biology interested in the study of the spatial variation of diversity at the intraspecific level (Avice 2009). The aim of this area of research is to identify the

geographical distribution of genetic lineages, considering that phylogenies are not only influenced by time but also by space.

During the last decades, numerous studies have investigated the phylogeography of species (e.g., Hewitt 2001; Schmitt 2007). Based on these studies, it could be demonstrated that climatic variations related with the Quaternary glaciations (2.6 Million years ago - present) had a non negligible effect on the spatial distribution of the genetic diversity of species (Hewitt 2000; Taberlet and Cheddadi 2002). In fact, it has been shown that the cyclic nature of these series of glacial and interglacial periods induced repeated organismal range shifts, which followed the distribution of their suitable habitat (e.g., Velichko, Novenko et al. 2007).

Unlike southern areas of the globe, the European subcontinent presents several characteristics that make it interesting when studying phylogeographic patterns. First, it is found at relatively high latitudes (higher than 45-40° latitude North) and presents an important exposed surface. Glacial events have thus an effect on the European area since polar ice-sheets easily reach the continent, and the impact of climatic variations is therefore extremely direct. Second, the region is bordered in the South by an internal sea, the Mediterranean, which acts as a climatic buffer, bringing heat and humidity, especially during cold periods, but blocking southward dispersal of organisms. Finally, the continent presents at its southern range a series of peninsulas (Iberia, Italy, Balkans), separated to the north of the rest of the continent by mountain ranges (Pyrenees, Alps, Carpathians-Dinaric mountains). During cold and dry periods as those identified during glacial events, these features create conditions in southern regions that are extremely favorable to temperate species, because they harbor warmer and moister conditions.

Geological studies concurred in demonstrating that during the last cold global events (Last Glacial Maximum, LGM, 21-18 thousand years ago –Kya), northern ice-sheets advanced in Europe as far south as northern Poland and Germany and covered the British Isles. At the same time, more southern mountain chains featured an extension of its glaciers

(Ehlers and Gibbard 2004). Moreover, because of the concentration of water in ice-sheets the sea level fell and several regions appeared interconnected by dry land due to the emergence of landbridges in the English Channel and the Adriatic. The presence of ice caps induced not only climatic variations (lower temperatures, drier climate), but also variations in the distribution of habitats on the remaining exposed land (Hewitt 1999). Considering this, it has been demonstrated that the tundra limit –today restricted to northern Europe and mountain zones– was shifted to central Europe, and that permafrost was present in regions as far south as southern France and northern Bulgaria (Hewitt 1999). These large environmental variations influenced organisms, which had to follow the cline of their suitable habitat by changing their distributions (*i.e.*, their realized ecological niche was spatially displaced). It has thus been proposed that, because conditions were milder in the southern peninsulas, temperate species found refugia in one or several of these regions (Stewart, Lister et al. 2010).

Taking these hypotheses into account, phylogeographic studies investigate the spatial distribution of the genetic diversity in different species distributed throughout the European area. These researches demonstrated that in temperate species there was a genetic signature indicating that southern regions harbored more important levels of genetic diversity than northern zones, less adapted for survival during glacial periods (Taberlet, Fumagalli et al. 1998). This pattern was explained by the fact that during glacial periods, different preglacial populations would have moved to these southern refugial areas, carrying with them their preglacial genetic diversity. From this point of view, each refugium would have thus potentially harbored different preglacial genotypes. Once climate warmed up, recolonization of ice-released lands would have happened from these refugia, with only some individuals invading successfully the deglaciated areas, and inducing an associated reduction in genetic diversity in those areas, a process called “leptokurtic” recolonization (Hewitt 1996). The consequence of this dynamics is that if we currently sampled the whole distribution ranges of temperate species, we would observe regions of high diversity, which

are generally placed in southern zones, with a drastic reduction in diversity when moving to the north.

Because the phylogeography of European organisms has been widely studied, it is today possible to identify in this continent regions of high and low genetic diversities (whatever species is considered), which are interpreted as refugia and postglacial recolonized areas, respectively (e.g., Taberlet, Fumagalli et al. 1998; Schmitt 2007). An integrated analysis of these studies allowed the identification and proposition of several phylogeographic patterns for the west-Palaearctic, which are currently considered as phylogeographic paradigms.

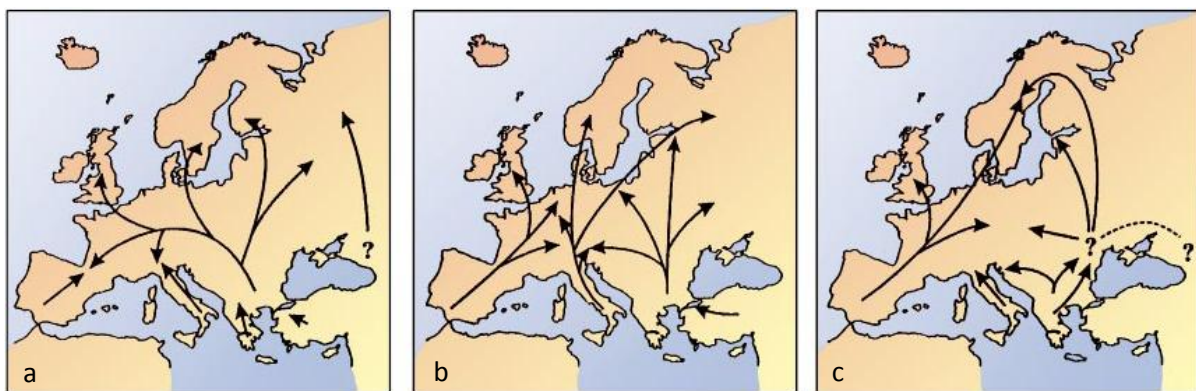


Figure 1 – Phylogeographic paradigms of post-glacial recolonization of Europe (Hewitt 2000). a – the grasshopper (*Chorthippus parallelus*); b – the hedgehog (*Erinaceus europeus/concolor*); c – the bear (*Ursus arctos*).

These paradigms are illustrated by the three species in which they were initially discovered (Figure 1): i) the grasshopper *Chorthippus parallelus*, which found shelter in three (or four) southern refugia, among which only the Balkanic lineage has recolonized the current distributional range; ii) the hedgehog *Erinaceus europeus/concolor* that found refugia in the three southern European peninsulas, from each of which recolonization of the current range happened; iii) the bear *Ursus arctos*, which was restricted to three (or four) refugia, with recolonization of the current area happening from the Iberian and the Balkanic/Carpathian refugia (Hewitt 2000).

Recently, palynological and genetic surveys have also informed that regions occupied by temperate species during glacial periods were not necessarily restricted to the southern peninsulas, but were also present in more northern latitudes, as in some regions south of the Carpathians or in north-western France (Stewart and Lister 2001; Provan and Bennett

2008). It is thus possible that, while main refugia were restricted to southern areas, other would have been present in more northern locations that harbored locally milder conditions than those observed in surrounding areas (regions recognized today as "cryptic refugia", Stewart, Lister et al. 2010).

Contrasting with the large number of studies centered on temperate species, far less is known about phylogeographic patterns of cold-related species (Brochmann, Gabrielsen et al. 2003). Probably the most important difficulty in the understanding of phylogeographic patterns for these species is that, unlike temperate species, they are currently experiencing populational contractions, and that regions they currently occupy should be considered as refugia (Stewart, Lister et al. 2010). Moreover, these species are generally difficult to sample, occupying mountainous areas or high latitudes regions.

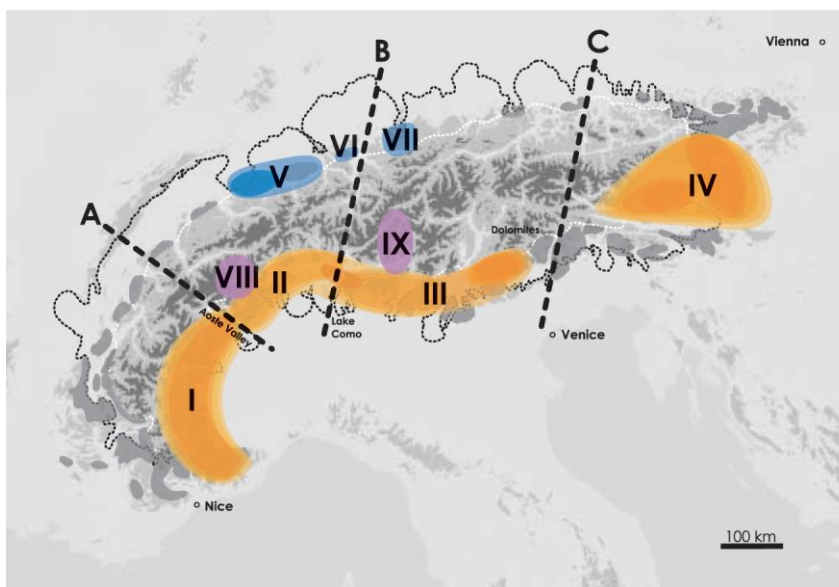


Figure 2 – Glacial refugial zones of mountain plant species in the Alps (Schönswetter, Stehlik et al. 2005). Colors indicate type of refugia: southern, central and northern. Broken lines indicate borders of biogeographic zones.

Despite that several arctic-alpine species have been already studied in Europe, their phylogenies either informed on the absence of clear genetic structure in the region (e.g., Alvarez, Thiel-Egenter et al. 2009) or on high admixture levels in southern mountain ranges (e.g., Schönswetter, Tribsch et al. 2004; Ehrich, Gaudeul et al. 2007). Moreover, the only area for which a phylogeographic paradigm similar to those of temperate species was proposed is the Alps (Figure 2; Schönswetter, Stehlik et al. 2005). In this region, the presence of genetic groups specific to the east, center, south and west of the mountain

range was identified in several plant species, with a clear differentiation in the distribution of refugial regions guided by soil chemistry (Alvarez, Thiel-Egenter et al. 2009).

Despite the fact that phylogeographic patterns of cold-related species are less investigated than those of temperate species, it is currently largely accepted that phylogeographic patterns in Europe are well known. However, little is known about the effect of ecological interactions on these phylogeographic patterns. Even though, as mentioned above, in the same way that climatic variations are able to force and guide the distribution of the suited habitat of a species, biotic interactions could also potentially influence the distribution of species through their effect on the definition of the realized niche.

4. Comparative phylogeography of interacting organisms, or the effect of biotic factors on the fate of species

Although the role played by biotic factors on the fitness of interacting species has been known for a long time (for a review, see Bronstein, Dieckmann et al. 2004), its effect on the intra-specific phylogeography of species has been rarely tested (e.g., Tsai and Manos 2010). In the current dissertation, I propose to test a simple hypothesis postulating that the type of interaction considered (mutualistic vs. antagonistic) influences the role that biotic factors have on the history of species. More precisely, because in mutualistic interactions partners depend one on the other, we could hypothesize that the survival and fate of each species should follow congruent phylogeographic patterns (*i.e.*, shared refugia and recolonization pathways). In contrast, in antagonistic interactions, where only one species depends on the presence of the other, we would expect a lack of congruence in phylogeographic patterns, since the independent species is able to disperse and move “freely”, while the dependent species systematically needs the presence of the other to assure its survival.

Trying to investigate these questions is the main topic of my PhD. During the time of my investigations, I have thus worked on two different biological models consisting of

emblematic European plant-insect pollination interactions, respectively mutualistic and antagonistic.

5. The models

Specific and obligate interspecific interactions are ideal models to test hypotheses related with the effect of the interaction on the level of congruence of phylogeographic histories of ecologically associated organisms. However, probably because of the important number of adaptations required for a specific interaction to evolve and maintain, and considering the risks associated with relying on only one partner for the survival of the species (especially in a variable abiotic environment), the number of these types of interactions is relatively low, particularly outside the tropics.

In the Palearctic, several of these interactions have been identified in systems involving plants and insects. Despite that they do not represent one-to-one interactions, they include a small number of interacting species, and in each case, the survival of a dependent species is threatened if the other species is not present.

5a. A case study of a mutualistic interaction – *Trollius europaeus* L. and *Chiastocheta* spp. Pokorny

Nursery pollination systems represent a special case of entomophilous mutualistic pollination: insects visit the flowers and actively or passively pollinate it, laying eggs on the developing flowers/fruits since larvae feed specifically on the plant reproductive organs (Dufay and Anstett 2003). Extremely specialized examples of this system are fig-fig wasps (Jousselin, van Noort et al. 2008), yucca-yucca moths (Pellmyr and Huth 1994), *Silene latifolia*-*Hadena bicruris* moths (Kephart, Reynolds et al. 2006) or *Trollius*-*Chiastocheta* flies (Pellmyr 1989). Among these, only the latter biological system has been shown to be widespread in the Palearctic region.

The genus *Trollius* (Ranunculaceae) is composed of between 18 and 31 geophyte species depending on the author (Doroszewska 1974; Pellmyr 1992; Després, Gielly et al.

2003). It presents a distribution associated to cold meadows and shows a range occupying the whole Palearctic kingdom. Because of its ecological requirements, it is distributed in lowlands at Northern latitudes higher than 50°, south of which it is present only in mountainous areas (alpine environments). Some species are also partially present in restricted regions of the Nearctic (*i.e.*, *T. riederianus*, *T. laxus* and *T. ledebouri*). A special feature that makes this genus interesting is that most species are visited and pollinated by different species of the Anthomyiid flies genus *Chiastocheta* Pokorny (Pellmyr 1992).

Flies of the genus *Chiastocheta* are exclusively distributed in the Palearctic, and it has been proposed that their centre of origin is eastern Asia (Pellmyr 1992). Posterior molecular studies suggested, however, that the distribution of the genus was first restricted to the European region and that only afterwards they invaded Eastern areas (Després, Pettex et al. 2002). Up to now, between 10 and 18 species have been described, based on different morphological characters (Hennig 1976; Michelsen 1985; Pellmyr 1992). All species are extremely specialized on the genus *Trollius*, whose seeds are eaten by the fly larvae (Pellmyr 1992), with no records of these species laying eggs on plants not belonging to the genus *Trollius*.

Among *Trollius* species, *T. europaeus* L. obtained the attention of biologists because of several special features. First, from a biogeographical point of view, this species is the only present at the Western edge of the Palearctic (Figure 3a), being isolated from all the remaining species by the Ural mountains of central Russia, where it comes partially into contact with other central Siberian taxa (Després, Gielly et al. 2003). Another of its special characteristics is that it displays an extremely distinctive flower morphology, composed of globose and closed yellow flowers, which contrast with the open-flower habit found in other *Trollius* species (Figure 3b, Pellmyr 1989). This latter trait is of particular interest for evolutionary biologists and ecologists who identified it as a character probably resulting of tight coevolutionary processes with associated pollinators (Pellmyr 1992). As a consequence of the closed nature of the flower cup, non-zoophilous (*e.g.*, wind) pollination is impossible

and the flower completely relies on insects for pollen transport, which is especially important considering its self-incompatibility (Pellmyr 1989). Moreover, because of the closed shape of the flower, the number of generalist visitors is strongly reduced, since it is difficult for them to reach the sources of nectar and pollen.

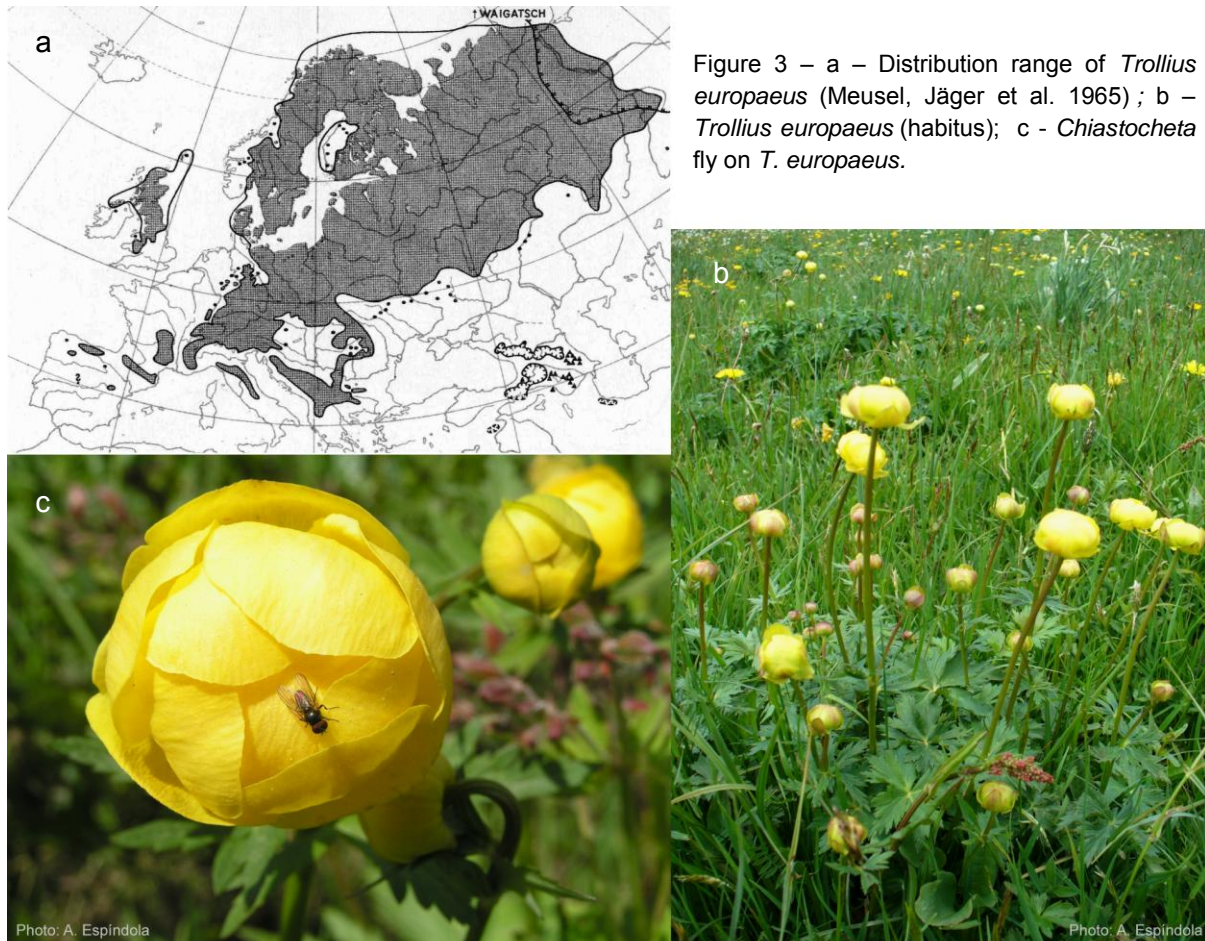


Figure 3 – a – Distribution range of *Trollius europaeus* (Meusel, Jäger et al. 1965); b – *Trollius europaeus* (habitus); c - *Chiastocheta* fly on *T. europaeus*.

Starting with Pellmyr (1989), investigations have gathered information that tended to confirm the coevolutionary nature of the forces that shaped this special floral display. For the last three decades, various researchers have demonstrated that the flower shape acts a “filter” for insects not belonging to the genus *Chiastocheta* (Figure 3c, Ibanez, Dujardin et al. 2009). Flies of this genus enter the closed flower to feed on nectar and pollen, mate and passively pollinate female flowers while getting covered with pollen from the enormous anthers. In exchange of the pollination service, flies lay eggs on the carpels into which larvae enter to feed on seeds (Pellmyr 1989). Up to now, eight species (*rotundiventris*,

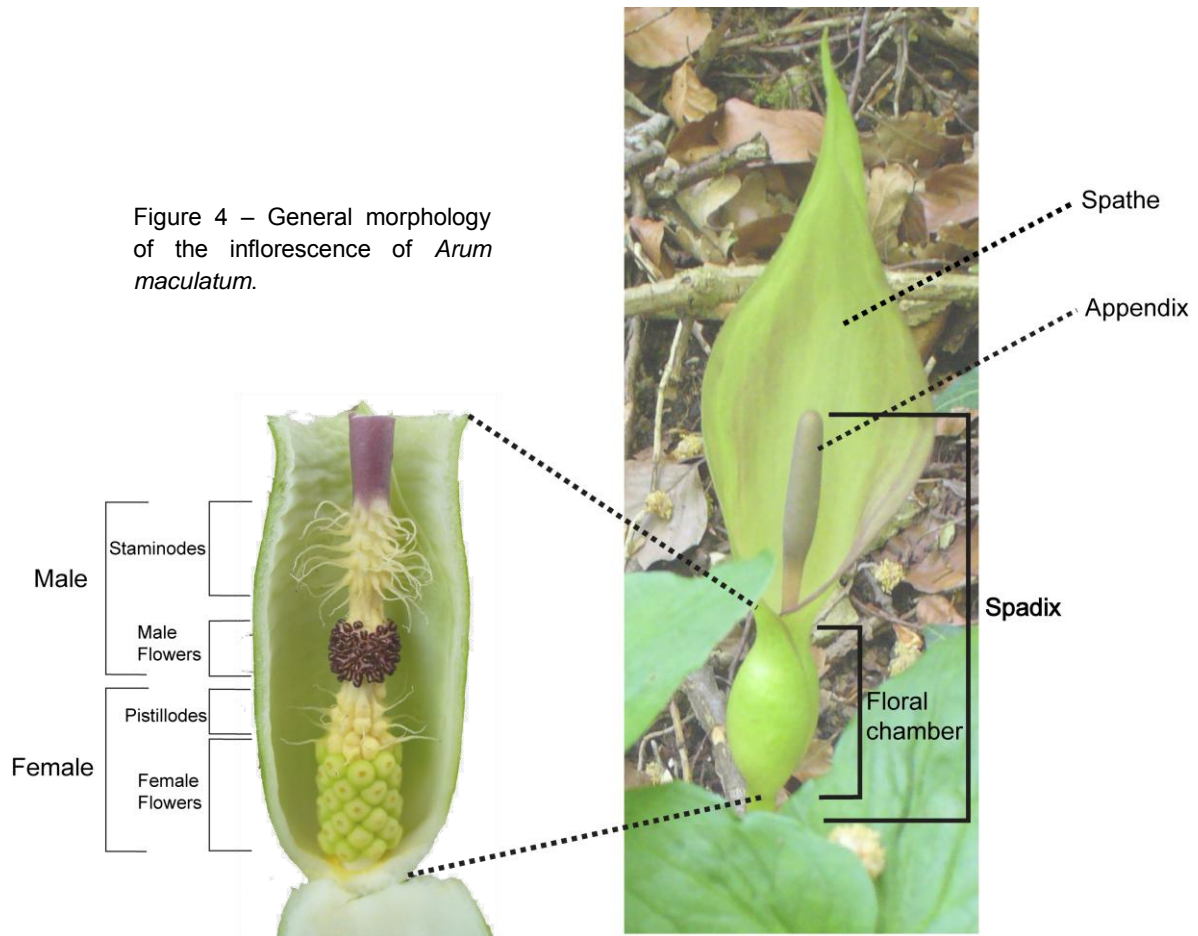
abruptiventris, *dentifera*, *lophota*, *inermella*, *setifera*, *trollii* and *macropyga*) have been demonstrated to exclusively visit *T. europaeus* (Pellmyr 1992; Jaeger and Després 1998).

Research on the biology of these species has also demonstrated that fly species present different and specific behaviors in terms of the position and timing of their laying strategy on the flower carpels. A temporal *suite* of fly visits is observed, with *C. rotundiventris* arriving at the very beginning of the flowering period, and laying generally one egg per flower head (Després and Jaeger 1999). Arriving at the very end of flowering, *C. dentifera* lays several eggs per flower head, ovipositing them between the central carpels (Pellmyr 1992; Després and Jaeger 1999). The remaining species arrive at different intermediate temporal points, laying also several eggs per inflorescence. Besides the fact that adults from different species behave differently during oviposition visits (Després and Jaeger 1999), larvae also differ in their exploitation of the seed resources (Pompanon, Pettex et al. 2006). It has been shown that probably because of an interaction between inter-larvae competition (Després and Jaeger 1999) and larvae-induced toxin accumulation in the plant (Ibanez, Gallet et al. 2009), each species presents a different seed consumption pattern. *C. rotundiventris* for example attacks several carpels and finally develops at the basis of the infructescence, and other species such as *C. dentifera* feeds on all seeds in one carpel (Pellmyr 1989). Once larvae finish feeding, they exit the fruit and fall onto the ground, where they pupate and overwinter (Ibanez, Gallet et al. 2009).

5b. A case study of antagonistic interaction – *Arum maculatum* L. and Psychodid flies

The most frequent antagonistic plant-insect interactions are composed of phytophagous insects that attack plants. However, rarer cases are those in which plants benefit from the interaction while insects are exploited. Among these rare cases are those involving lure-and-trap pollination. In these interactions, the plant attracts one or several insects to exploit them as pollen carriers. Examples of this type of exploitation are present in several species within

the genera *Aristolochia* L. (Berjano, Ortiz et al. 2009) and *Arum* L. (Gibernau, Macquart et al. 2004), both present in the West-Palearctic.



Unlike *Aristolochia*, the genus *Arum* (Araceae) is restricted to a peri-Mediterranean/Eurasian distribution (Boyce 1993; Boyce 2006). This genus is historically emblematic, because of its shape, biology and ecology. Its first mention goes back to ancient Greek records (Theophrastus 370 B.C.), being much later defined by Linnaeus, and currently providing the name of the family. All taxa included in this genus are geophytes and develop underground tubers (Boyce, 1993). As all Araceae, they harbor a special inflorescence morphology (Figure 4), formed by a spathe –a modified bract– that encloses a central spadix, harboring the sexual structures (Mayo, Bogner et al. 1997). In the genus *Arum*, the spadix is separated in two regions: a lower part, which displays female and male flowers, enclosed in the floral chamber –a closed space defined by a contraction of the lower part of the spathe– and an upper part, which is composed of the appendix, responsible of

heat and volatile production (see below) (Figure 4, Boyce 1993). Despite that the genus has been known for centuries, several systematic, historical and geographical issues blurred delimitation and definition of species (Boyce 1993; Bedalov and K pfer 2005; Boyce 2006).

Among all species within the genus *Arum*, the only one presenting a wide pan-European distribution is *A. maculatum* L. (Figure 5a; *A. italicum* and *A. cylindraceum* are also widely distributed but less northward). Besides being used for centuries as a source of starch, active principles in medicine and for magic grounds (Prime 1960), the species has interested evolutionary biologists because of its very special pollinator interaction (Lack and Diaz 1991). This species displays a specific and obligate lure-and-trap sapromyophilous pollination system that makes it dependent for cross-pollination on small Psychodid fly species (Figure 5b; Diaz and Kite 2002). Pollination is possible thanks to a series of adaptations displayed by the plant, which allow attracting and trapping ready-to-lay female flies (see below).

Figure 5 – a – Distribution range of *Arum maculatum*; b – *A. maculatum* (habitus); c – view from above of Psychodidae trapped in an *Arum* floral chamber.



Probably one of the most impressive adaptations is the one involving heat and volatile production. From one day prior to flowering to the second day of anthesis, *A. maculatum*

experiences several cycles of heat production, which can make the flower reach temperatures of up to 15-25°C higher than the surrounding air (Gibernau, Macquart et al. 2004). The first cycle happens the day before anthesis, and is supposed to be involved in female flower maturation. The second helps the spathe unfold and happens at the beginning of flowering. At this moment, volatiles are released from the top of the appendix in association with heat production. Volatile analyses have identified several majoritary compounds being largely present in animal faeces and urine (indole, *p*-cresol, 2-heptanone), and supposed to be key in the attraction of these coprophilous pollinators, whose females are seeking a laying site (Kite 1995).

Flies are thus lured and after landing on the oily spathe, slip into the floral chamber, where they stay trapped because of an array of male flowers modified into hairs (staminodes) that block the exit, and the presence of oil droplets on the chamber walls. Flies are kept prisoners for up to one day, a considerable amount of time if we consider the fly life-span (eight to twelve days; Vaillant 1971). By trying to flee, captured flies will –if they had been previously trapped by another *Arum*– pollinate the female flowers and, once pollen is ripe, they will be covered with it. After liberation of pollen, staminodes wither, walls lose their oily nature and flies are set free. Even if only a very small proportion of them will be trapped again by another *Arum*, this small part will be sufficient to pollinate all flowers and to allow cross-pollination in this species (Lack and Diaz 1991).

One might ask how this type of interaction is maintained through evolutionary times. Several factors could allow keeping up this interaction. On the one hand, fly species involved present an extremely short generation time (12-18 generations/year, Vaillant 1971), which means that only one or two generations are really selected by the plant, which massively flowers during a short period of time (about one month, Ollerton and Diaz 1999). This could indicate that even if selection is strong over one or two fly generations, its effects could be easily lost during the dozen additional generations occurring over the rest of the season. On the other hand, we could also ask why flies continue to be attracted by an odour that they

could avoid; a possible explanation invokes a scenario in which the chemical cue used by the plant to attract the insects is too selective to be counter-selected by them since most odors involved in the floral scents are present in the flies' laying sites. Finally, it is also possible that because insects rarely die in the floral chamber and because population sizes are huge (up to several thousands per square meter, Arshad and Moh Leng 1991), the strength of selection is not substantial compared to the effect of drift.

6. The structure of this study

The present work is divided in two parts, which investigate the different aspects presented above using, as case studies, the two models of specific and obligate plant-insect pollination interactions. The **first section** considers different topics related with *Arum maculatum* L. and its Psychodid pollinators. Here I first investigate, using phylogenetics, biogeography and character tracing, the systematics of the genus *Arum* (**chapter one**). Afterwards, I treat *Arum* phylogeny's counterparts, by identifying the phylogenetic relationships among members of the subfamily Psychodinae (**chapter two**). Further, I proceed to the analysis of the variation in the composition of *A. maculatum*'s visitors at different spatial scales (**chapter three**). Extending the analysis of the plant-insect interaction to a larger view, I evaluate the effects of the identity and abundances of pollinators on the reproductive strategy of two closely related *Arum* species, which strongly differ in their reproductive strategies (**chapter four**). Based on results provided by all previous chapters, I perform phylogeographic analyses of each member of the interaction to finally evaluate the level of congruence among them (**chapter five**).

In the **second section** I study topics related with those of the first section, but on a mutualistic biological model, *i.e.*, the nursery pollination system composed by *T. europaeus* and its *Chiastocheta* pollinators. Because when studying coevolutionary processes it is key to understand the identity and limits of the units under selection, I start this section by evaluating the current species circumscription in the genus *Chiastocheta* (**chapter six**). Then, I evaluate the phylogeography of *T. europaeus*, proposing a new technique to infer

the fate of genetic lineages after distributional contractions in cold-adapted species (**chapter seven**). Finally, I identify the flies' phylogeographies, which I further compare to the previously obtained spatial genetic structure of the plant (**chapter eight**).

Chapter one – Phylogeny, biogeography and systematics in the genus *Arum* L.

The circumscription of species in the genus *Arum* has been discussed for several years. One of the main difficulties in its systematics lays in the fact that traits in this genus are extremely plastic, which does not help in the identification of morphological synapomorphies. Moreover, because of historical reasons, several taxa have been simultaneously described in different regions under different names, which complicated even more the interpretation of the identity of species. In this chapter, using chloroplastic sequences and different analytical approaches, I aim at resolving the infra-generic phylogeny of the genus *Arum*. Considering the need to identify characters informative of the relationships among species, morphological characters are traced on the molecular phylogeny. Finally, I apply biogeographic analyses to identify the most probable center of origin of the genus and to infer the timing and spatialization of dispersal and radiation of the main lineages.

While some subgenera appear supported, most of them are challenged by our results. Phylogenetic inferences thus suggest that taxonomic revisions should be done in this genus. Probably one of the most interesting information provided by this study is related to the identity of *A. maculatum*, which appears separated into two different clades, strongly structured geographically. It thus seems that what was up to now identified as a member of this species to the east of Southern Bulgaria is another specific entity, for which further descriptions should be done in the future. The distribution range of *A. maculatum sensu stricto* is thus strictly European. By tracing morphological characters, we demonstrate that the level of ploidy and the tuber shape are informative of the relationships in the group. It is important to note that this result shows that characters used up to now to define subgeneric

systematic, which are only based on floral measures, are highly homoplastic: because flower characters are highly adapted to the pollination system in *Arum*, it is likely that they evolve extremely quickly with a high rate of homoplasy. Finally, biogeographic analyses demonstrate that the history of the genus is tightly related to the history of the Mediterranean Basin, with colonization and vicariance events happening synchronously with timing of geographical connections and disruptions that occurred in this area.

Chapter two – Phylogenetic relationships within the subfamily Psychodinae in a temporal framework (Diptera: Psychodidae)

Among-family taxonomic relationships in the order Diptera were continuously challenged in the last decades. Such a lack of clear systematics is also found at lower taxonomic levels, as for instance in the subfamily Psychodinae. Molecular phylogeny is a field that allows inferring species relationships without any prior consideration of morphological synapomorphies, which can be further identified once species groups have been settled. Here, I present a molecular phylogeny of this worldwide and very common subfamily, comprising notably species involved in a tight pollination interactions with European Araceae.

Besides showing that the two most abundant visitors of *A. maculatum* correspond to sister lineages distributed in the same sub-clade, we demonstrate that the current relationships in the group need to be strongly reconsidered, arguing for revised and new subdivisions of tribes, subtribes and genera. Moreover, our phylogenetic hypothesis shows that the timing of divergence in Psychodinae spanned the last 86 million years, an ancient temporal origin when investigating divergence time within subfamilies. Similarly, most species originated during the Paleogene (65.5-23 Mya), through a diversification process that followed the Cretaceous-Paleogene (K-T) massive extinction event.

Chapter three – Biogeography of *Arum maculatum*'s visitors

In the last century, *A. maculatum* has been described as a species specifically pollinated by one single fly taxon: *Psychoda phalaenoides* L. (Diptera: Psychodidae). However, studies were performed only in the western part of the plant's distribution range, while *A. maculatum* is widely distributed from southern Scandinavia to Bulgaria and from Spain to Poland. Even though these studies pioneered our knowledge on this antagonistic interaction, they provided only little insights in the consistency of specificity across the distribution range of the plant. By investigating the visitor composition throughout the whole distribution range of the plant, we test predictions from the theory of the Geographic Mosaic of Coevolution (GMC; Thompson 2005), which forecasts that some variation in the nature of interacting species occurs if the environment is heterogeneous.

In this study we sampled floral chambers from all over the distribution range of the plant and identified the variation in flower visitors. In order to quantify the environmental variation related to these sites, we extracted the climatic characteristics of each sampled location from climatic layers and performed ecological niche models analyses. We demonstrate that the composition of the flower chambers varied as a function of latitude, with the most abundant species (*P. phalaenoides*) dominating in central and north-western Europe, while it is almost absent in more Mediterranean environments, where the most abundant species is *Psycha grisescens* Tonnoir, a sister species of *P. phalaenoides*. We show that differences in the composition and relative abundances of one and another species are correlated with environmental factors. Our study thus shows that predictions of the GMC are confirmed in the case of *A. maculatum*.

Chapter four – The effect of insects in the reproductive strategies of three closely related *Arum* species

In coevolutionary interactions, it has been proposed that synergistic forces induce character coevolution in all species involved, which increases the fitness of each organism. The reproductive strategy of a plant can be considered as one of these selected characters. In Araceae, a remarkable reproductive strategy is the one defined by the application of the size-advantage model (SAM). SAM proposes that the energy allocated to different sexes is expected to vary in hermaphrodite organisms with the accumulation of energy in the organism, favoring an investment on the most costly sex when the organism has accumulated enough energy. The application of the SAM has already been investigated in some Araceae, and it has been demonstrated that, while it is applied in some species, this is not necessarily the rule for all species. In the genus *Arum*, more precisely, its application varies depending of the species considered. Former studies showed its application in *A. italicum*, but not in *A. maculatum*.

In this chapter, we propose that the application or not of the SAM could be correlated with the pollination strategy of the plant. In order to test this theory, we compared the pollinator composition of *A. maculatum* (a specialist) with those of *A. cylindraceum* and *A. italicum* (generalists). Based on the idea stated above, we make predictions of application or not of the SAM based on a simple model representing the putative relationship with pollinating insects (specialists vs. generalists) and perform allometric measures in the widespread, but neglected, *A. cylindraceum* to confirm or reject predictions.

Our results indicate that while *A. maculatum* displays pollination features that do not favor the application of the SAM as an adaptive trait, this is probably not the case in other *Arum* presenting less specialized reproductive biologies such as *A. cylindraceum*. Our simple predictive model seems to allow understanding the relationship between pollination strategy and application of the SAM.

Chapter five – Comparative phylogeography of *Arum maculatum* and its specific and obligate pollinators

One of the general objectives of my thesis was to investigate if the phylogeographic patterns of interacting species demonstrated remarkable characteristics. I aimed at testing whether or not the type of interaction could have an effect on the congruence in phylogeographies of associated species, both in mutualistic and in antagonistic specific and obligate interactions.

In this chapter we proceed to the comparison of the phylogeographies of species involved in the antagonistic interaction featuring *A. maculatum* and its pollinating Psychodids. Our working hypothesis is that since in this antagonistic relationship only one of the partners is ecologically dependent of the other, we expect that phylogeographic patterns of the plant and the insects should not be congruent.

In order to test this, we respectively sequenced and applied genomic screenings to insects and plants that we collected at the same locations throughout the whole distribution range of the plant. We afterwards inferred the genetic structure of both actors, identified their spatial distributions, and finally compared the resulting phylogeographic patterns.

Our study reveals that while the plant is clearly geographically structured, this is not the case of the flies. This lack of congruence between the two organisms confirms our initial expectation. It appears that when both species do not reciprocally need each other to survive, phylogeographic patterns are not congruent. Another explanation is however related to the generation times and population sizes present in each species: while the plant has a generation time of 10-15 years, flies undergo 12-18 generations per season, leading to extreme differences in the absolute rate of molecular evolution in both groups. Moreover, insects appear to be able to disperse over large distances, which might blur any phylogeographic signal.

Despite these results, it is important to point out that the genetic structure we obtained for the plant is almost identical to that previously obtained by the analysis of the variation in the composition of the visitors collected in floral chambers over a large spatial scale (chapter three). It is thus possible that some genetic structuring of the plant is driven by the insect species composition, or the other way around.

Chapter six – Phylogeny of *Chiastocheta*

It is not possible to work on coevolutionary systems if the limits of the units of selection are not well defined. In this chapter, we investigated the species concept and limits in *Chiastocheta* species associated with *T. europaeus*. Using different molecular markers and applying biogeographic approaches, we elucidate the phylogenetic relationships among *Chiastocheta* species. The main result of this study is that not all the morphologically defined species are supported by molecular data. While three species show an agreement between morphological and molecular synapomorphies, the others do not present such congruence. We further demonstrate that most species are currently experiencing hybridization, which blurs among-taxon boundaries. The fact that some of these species are hybridizing changes our view of this system, since if hybridization is possible, any coevolutionary force acting on these entities would be much more diffuse than previously thought. From a spatio-temporal point of view, we show that the genus is relatively ancient and that the whole infra-generic diversification occurred during the Quaternary. We also demonstrate that the most recent common ancestor of the genus was actually distributed in Europe, and that dispersal towards Central Asia independently occurred two or three times. Further dispersal towards East Asia and Japan occurred once each from Central Asia.

Chapter seven – Phylogeography of *Trollius europaeus* at the European scale

Little is known about phylogeographic patterns of cold-related organisms in Europe. Moreover, because these organisms present a currently contracted distribution range, the only processes that can be identified when studying their spatial genetic structure are those related to the dynamics of contraction and to range shifts. However, if our aim is to identify such processes, we need to have a view on past genetic diversities to compare past and current patterns. Generally this sight into past diversity is done by using ancient DNA found in fossils or by the inference of past distributions based on macroremains. Unfortunately, ancient material is not systematically available for all species, which restricts our capability to infer the genetic consequences of past demographic contractions related to the termination of glacial periods in cold-related species.

In this chapter we propose a solution to this problem by exploiting the potentials of hindcasting techniques to estimate the past distribution range of *Trollius europaeus* at the LGM. This indirect approach provides a probabilistic view into the past genetic diversity, which is especially useful when considering species for which no ancient material is known. Combining this modeling approach with the analysis of the current genetic structure of the plant, we demonstrate here that the termination of the LGM probably induced a relatively limited loss of genetic diversity in *T. europaeus*, which contrasts with other arctic-alpine species, for which the loss was much more important.

Chapter eight – Phylogeography of three *Chiastocheta* species and the influence of the plant in shaping their genetic structure

The aims of this chapter are to test if closely ecologically related species present similar phylogeographic patterns, and to understand what is the level of congruence between the phylogeographic patterns of the pollinators and the plant in the *Trollius-Chiastocheta* mutualistic interaction.

Based on results of chapter six, we investigate the phylogeographic patterns of the three species that appeared to be well-defined both morphologically and genetically in our previous molecular analysis (*i.e.*, *C. rotundiventris*, *C. lophota* and *C. dentifera*). To do so, we used sequence data to infer phylogenetic topologies. On the one hand we applied “traditional” phylogeographic methods, in which we visually estimated the geographic distribution of the genetic clades and clusters in space. Second, we applied statistical phylogeography techniques, which allowed us to directly test the probability that the genetic structure obtained in the phylogenetic inferences has been produced under a given coalescent scenario defined by the plant genetic structure (chapter seven).

Our results indicate that, despite having similar ecologies, each fly species presents a different phylogeographic pattern, with one species showing a clearly geographically defined genetic structure (*C. lophota*), another harboring very high levels of admixture (*C. rotundiventris*), and the third presenting a phylogeographic structure which does not statistically differ of what would have been obtained in the case that the plant’s spatial genetic structure had driven its phylogeography (*C. dentifera*). We thus demonstrate that the phylogeographic histories of at least one fly species and that of the plant statistically overlap, which partially confirms our initial working hypothesis of congruence in phylogeographic patterns of species involved in mutualistic interactions.

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Chapter one

*New insights into the phylogenetics and
biogeography of Arum (Araceae): unravelling its
evolutionary history*

Anahí Espíndola, Sven Buerki, Marija Bedalov,
Philippe Küpfer and Nadir Alvarez

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New insights into the phylogenetics and biogeography of *Arum* (Araceae): unravelling its evolutionary history

ANAHI ESPÍNDOLA^{1*}, SVEN BUERKI², MARIJA BEDALOV^{3,4}, PHILIPPE KÜPFER⁴
and NADIR ALVAREZ⁵

¹Laboratory of Evolutionary Entomology, Institute of Biology, University of Neuchâtel, Rue Emile-Argand 11, CH-2009 Neuchâtel, Switzerland

²Department of Biodiversity and Conservation, Real Jardín Botánico, CSIC, Plaza de Murillo 2, 28014 Madrid, Spain

³Department of Botany, Faculty of Sciences, University of Zagreb, Marulićev Trg 20/II, HR-10000 Zagreb, Croatia

⁴Laboratory of Evolutionary Botany, Institute of Biology, University of Neuchâtel, Rue Emile-Argand 11, CH-2009 Neuchâtel, Switzerland

⁵Department of Ecology and Evolution, University of Lausanne, Biophore Building, 1015 Lausanne, Switzerland

The heat- and odour-producing genus *Arum* (Araceae) has interested scientists for centuries. This long-term interest has allowed a deep knowledge of some complex processes, such as the physiology and dynamics of its characteristic lure-and-trap pollination system, to be built up. However, mainly because of its large distributional range and high degree of morphological variation, species' limits and relationships are still under discussion. Today, the genus comprises 28 species subdivided into two subgenera, two sections and six subsections. In this study, the phylogeny of the genus is inferred on the basis of four plastid regions, and the evolution of several morphological characters is investigated. Our phylogenetic hypothesis is not in agreement with the current infrageneric classification of the genus and challenges the monophyly of several species. This demonstrates the need for a new infrageneric classification based on characters reflecting the evolution of this enigmatic genus. To investigate the biogeography of *Arum* deeply, further spatiotemporal analyses were performed, addressing the importance of the Mediterranean basin in the diversification of *Arum*. Our results suggest that its centre of origin was the European–Aegean region, and that major diversification happened during the last 10 Myr.

ADDITIONAL KEYWORDS: character tracing – infrageneric systematics – Mediterranean biogeography – phylogenetic inferences.

INTRODUCTION

With 109 genera and over 3700 species described (Mayo, Bogner & Boyce, 1997) Araceae have a worldwide distribution and are found in a wide range of environments, from Arctic–Alpine (e.g. *Calla palustris* L.) to xerophytic (e.g. *Anthurium nizandense* Matuda),

with most species occurring in the tropics. The family encompasses a large variety of life forms, from epiphytic to aquatic, attesting the numerous adaptive radiations that have occurred in this early Cretaceous family (Chase *et al.*, 2006; Anderson & Janssen, 2009). A remarkable feature in Araceae is the evolution of heat production in several genera (Minorsky, 2003), especially those displaying pollination-related associations with arthropods, in which thermogenesis is associated with the emission of volatile compounds and the attraction of pollinators (Moodie, 1975).

*Corresponding author. E-mail: maria.espindola@unine.ch

One of the few Palaeartic representatives of Araceae is the herbaceous genus *Arum* L., which comprises 28 described species (Lobin *et al.*, 2007; CATE project, 2010). Because of its characteristic flowers, showing adaptations for trapping pollinators (Gibernau, Macquart & Przetak, 2004), and its ability to produce heat and odours, this genus has fascinated not only scientists but also the wider public for centuries (Theophrastus, 370 BC; Hruby, 1910; Boyce, 1993). As shown by archaeological and historical records, several *Arum* spp. have been used by humans since ancient times for food (tubers), medicine (fruits, leaves, tubers), fashion (tuber starch) and even magic (leaf decoctions) (Prime, 1960).

The inflorescences of *Arum* consist of two parts: a spadix and a spathe (Boyce, 1993). The spadix displays the unisexual flowers and harbours adaptations involved in heat production, whereas the spathe is a modified bract surrounding the spadix. One of the distinctive synapomorphies of the genus is the partition of the spadix. The lower zone comprises both female (lower portion) and male (upper portion) flowers placed in a floral chamber, which is usually delimited by male-sterile flowers modified as hairs: the staminodes. Its apex is a smooth, subcylindrical, usually stipitate organ, known as the spadix appendix (Boyce, 1993). This structure is also recognized as an efficient thermogenetic organ with which the plant attracts pollinating arthropods with heat and production of volatile compounds. The combination of odour emission and hair presence at the top of the floral chamber (acting as a fence) is a key feature for the efficient trapping of arthropods during the female receptive period and until pollen release (Gibernau *et al.*, 2004).

Historically, the genus was defined by Fuchs (1542) and later established by Linnaeus (1753). The different species were first circumscribed on the basis of morphology (Schott, 1832), and chromosome counts (Bedalov, 1981) led to the identification of different ploidies in the genus (di-, tetra- and hexaploids, $x = 14$; for a review, see Boyce, 1989). In the most recent revisions of the genus (Boyce, 1993, 1994, 2006; Bedalov & Küpfer, 2005), several morphological characters (tuber shape, flower disposition, growth period, spadix shape and structure of sterile flowers) have been used to build a classification comprising two subgenera, two sections and six subsections. The subgenus *Gymnomesium* (Schott) Engl. is monospecific, including only the Hercynian endemic *Arum pictum* L.f. Subgenus *Arum* Engl. includes sections *Arum* and *Dioscoridea* Bronner, the latter being divided into six subsections (Table 1).

This classification may be controversial, notably because: (1) several taxa have been defined on the basis of herbarium specimens (this approach may not

be optimal in this group as important characters are observable only on fresh material; Boyce, 1989); (2) species having large distributions and studied locally were sometimes either simultaneously described under different names (e.g. *A. italicum* Mill.) or assigned to different taxa when they belonged to the same taxon (e.g. *A. cylindraceum* Gasp.) (Bedalov & Küpfer, 2005); (3) following this last point, as several species harbour a high level of intraspecific polymorphism, this may even trigger the splitting of widely distributed taxa (Boyce, 2006). Therefore, it is now an appropriate time to evaluate the systematics of *Arum* based on molecular evidence. Published molecular phylogenetic analyses including species of *Arum* have focused on the investigation of relationships at the family level and have lacked sampling and resolution at the infrageneric level (Cabrera *et al.*, 2008; Mansion *et al.*, 2008). In this article, we aim to produce a phylogenetic hypothesis for the genus *Arum* by sequencing four plastid regions suitable for addressing relationships at the infrageneric level based on 26 of the 28 described species. This will allow us to assess the validity of the current classification and to examine the evolution of several key characters. To decipher the evolutionary history of this early Miocene genus (Mansion *et al.*, 2008), we perform spatiotemporal analyses to determine the events that played a central role in the radiation, dispersion and isolation of the different species (Sanmartín, Enghoff & Ronquist, 2001; Médail & Diadema, 2009). Finally, on the basis of our results, we suggest guidelines for a new infrageneric classification of the genus *Arum*.

MATERIAL AND METHODS

SAMPLING

Analyses were based on 64 specimens, representing 26 of the 28 described species and spanning all subgenera, sections and subsections of *Arum* (Table 1). On the basis of Mansion *et al.* (2008), *Dracunculus canariensis* Kunth, *D. vulgaris* Schott, *Biarum davisii* Turrill and *B. dispar* (Schott) Talavera were used as outgroup taxa. Samples were either provided by the DNA Bank of the Royal Botanic Gardens, Kew (UK) or extracted directly from dried plant material from herbaria or field collections (Appendix 1).

DNA EXTRACTION, AMPLIFICATION AND SEQUENCING

DNA of freshly collected material and herbarium samples was extracted using the DNeasy Plant Kit (Qiagen, Basle, Switzerland). The plastid regions *3'rps16-5'trnK*, *ndhA* intron, *psbD-trnT* and *rpl32-trnL* were amplified with the primers described in Shaw *et al.* (2007). Amplifications were performed in a

Table 1. Current subgeneric taxonomy of genus *Arum* L. Taxa with an “*” were not included in the present study.

| Subgenus | Section | Subsection | Species |
|--------------------|--------------------|------------------------------|--|
| <i>Gymnomesium</i> | | | <i>A. pictum</i> L.f. |
| <i>Arum</i> | <i>Arum</i> | | <i>A. byzantinum</i> Schott |
| | | | <i>A. concinnatum</i> Schott |
| | | | <i>A. italicum</i> Mill. |
| | | | <i>A. maculatum</i> L. |
| | | | <i>A. megobrebi</i> Lobin, M.Neumann, Bogner & P.C.Boyce |
| | <i>Dioscoridea</i> | <i>Alpina</i> | <i>A. cylindraceum</i> Gasp. |
| | | | <i>A. lucanum</i> Cavara & Grande |
| | | <i>Discroochiton</i> | <i>A. apulum</i> (Carano) P.Boyce |
| | | | <i>A. balansanum</i> R.R.Mill |
| | | | <i>A. besserianum</i> Schott |
| | | | <i>A. cyrenaicum</i> Hruby |
| | | | <i>A. elongatum</i> Steven |
| | | | <i>A. gratum</i> Schott* |
| | | | <i>A. hainesii</i> Riedl* |
| | | | <i>A. nigrum</i> Vell. |
| | | | <i>A. orientale</i> M.Bieb. |
| | | | <i>A. purpureospathum</i> P.C.Boyce |
| | | | <i>A. sintenisii</i> P.C.Boyce |
| | | <i>Tenuifila</i> | <i>A. jacquemontii</i> Blume |
| | | | <i>A. korolkowii</i> Regel |
| | | | <i>A. rupicola</i> Boiss. |
| | | <i>Hygrophila</i> | <i>A. euxinum</i> R.R.Mill |
| | | | <i>A. hygrophilum</i> Boiss. |
| | | <i>Poeciloporphyrochiton</i> | <i>A. dioscoridis</i> Sibth. & Sm. |
| | | | <i>A. palaestinum</i> Boiss. |
| | | <i>Cretica</i> | <i>A. creticum</i> Boiss. & Heldr. |
| | | | <i>A. idaemum</i> Coust. & Gand. |

master mix containing 0.5 × buffer, 150 mM deoxy-nucleoside triphosphate (dNTP), 0.7 mM MgCl₂, 0.3 μg μL⁻¹ bovine serum albumin (BSA), 0.5 μM primers and 1 unit of *Taq* Polymerase (Promega, Dübendorf, Switzerland) made up to a final volume of 30 μL with purified MilliQ water. Reactions were run in a TGradient thermocycler (Biometra, Goettingen, Germany). Initial denaturation was programmed for 2 min 30 s at 95 °C, followed by 35 cycles at 95 °C for 35 s, 54–60 °C for 45 s, 72 °C for 1 min and a final extension of 8 min at 72 °C. The purification of PCR products and fluorescence sequencing were performed by Macrogen, Inc. (Seoul, South Korea) and FASTERIS Life Sciences (Geneva, Switzerland) with the same primers as used for PCR amplification.

SEQUENCE ALIGNMENT AND PHYLOGENETIC RECONSTRUCTIONS

Automatically generated base-calls for all sequences were checked and edited using ChromasPro 1.41 (Technelysium Pty Ltd, Tewantin, Australia). For each

plastid region, alignment was performed using the ClustalW algorithm implemented in Bioedit 7.0 (Hall, 1999), followed by minor manual corrections. After concatenation of the four regions, a matrix of 3723 bp was obtained. Gaps were further coded following the simple method of Simmons & Ochoterena (2000), as implemented in FastGap 1.2 (Borchsenius, 2009).

The numbers of constant (C), variable (V) and potentially parsimony-informative (PI) sites were calculated for each partition using PAUP* v4.0b10 (Swofford, 2002). Before computing total evidence trees, we tested for incongruence among the four regions by applying the partition homogeneity test as implemented in PAUP* v4.0b10 with 100 replicates (this test is equivalent to the incongruence length difference test of Farris *et al.*, 1994; for convenience, it is referred to as the ILD test). Total evidence trees (*sensu* Kluge, 1989) were determined using both Bayesian inference and maximum parsimony (MP) approaches.

Bayesian Markov chain Monte Carlo (MCMC) methods were used to approximate the posterior prob-

ability distribution of the phylogenetic trees on the basis of the combined plastid dataset with four distinct partitions plus one partition for the gap information, by running MrBayes v.3.1.2 (Ronquist & Huelsenbeck, 2003). Model selection for the plastid DNA partitions was tested using MrAIC (Nylander, 2004) based on the Akaike information criterion (Akaike, 1973), and a restriction model was applied to the partition containing the coded gaps. Three independent runs with one cold and five heated chains were run for 5×10^7 generations each. Frequencies were sampled every 1000 generations and temperature was fixed to 0.5. The convergence of MCMC was tested by computing the potential scale reduction factor (PSRF; Gelman & Rubin, 1992) as implemented in MrBayes, and by determining the effective sample size for each parameter using Tracer v.1.4 (Rambaut & Drummond, 2004). Accordingly, the burn-in period was set to 10^7 generations until stationarity of the likelihood value was established among the runs, and 10 000 sample points were discarded (20% of the total number of trees). The remaining 40 001 trees from each run were pooled (120 003 trees in total) to estimate the posterior probability distribution of the phylogenetic inference. To yield a single phylogenetic hypothesis, the posterior distribution was summarized in the 50% majority-rule consensus tree (referred to as the half-compatible tree in MrBayes), with a Bayesian posterior probability (BPP) at each node indicating statistical support.

The combined dataset was further analysed under the MP criterion using the parsimony ratchet (Nixon, 1999) as implemented in PAUPrat (Sikes & Lewis, 2001). Based on recommendations by Nixon (1999), ten independent searches were performed with 200 iterations, and 15% of the parsimony-informative characters were perturbed using PAUP* version 4.0b10. The shortest equally most parsimonious trees were combined to produce a strict consensus tree. Node support was determined by computing decay indices (DIs) (Bremer, 1988) as implemented in TreeRot 3.0 (Sorenson & Franzosa, 2007). DI measures the number of extra steps in tree length required before a node collapses (Bremer, 1988; Baker & DeSalle, 1997).

Finally, the level of congruence between Bayesian and MP analyses was determined by computing the quartet distance (Estabrook, 1992) between the two topologies. Considering that the distances between the different topologies were small (see Phylogenetic inferences section in Results), the remaining analyses were only based on the Bayesian inference analysis.

CHARACTER EVOLUTION

Character tracing was performed on traits generally used in taxonomic studies of *Arum*. On the basis of the

topology of the 50% majority-rule Bayesian analysis, the following categorical characters were mapped using Mesquite 2.6 (Maddison & Maddison, 2009) with accelerated transformation optimization (ACCTRAN) and unordered parsimony: tuber form (rhizomatous/discoid), flower type (flag/cryptic), spadix/spathe ratio (0–0.5; 0.5–1; > 1) and ploidy (di-, tetra-, hexaploid). Characters were obtained from the latest systematic studies performed on the species (Boyce, 1993, 2006; Bedalov & Küpfer, 2005; Lobin *et al.*, 2007).

DATING AND BIOGEOGRAPHICAL ANALYSES

Because the molecular clock hypothesis was rejected (data not shown), the 50% majority-rule Bayesian inference tree was rendered ultrametric using the penalized-likelihood method (Sanderson, 2002; hereafter PL), as implemented in the program r8s v.1.71 (Sanderson, 2004) by applying a smoothing value of 1000 and the truncated Newton algorithm. The most external outgroup, *B. davisii*, was pruned for the estimation of the divergence time as required by the program (see Sanderson, 2004). The following calibration points were applied (according to Mansion *et al.*, 2008): (1) the root node (i.e. the most recent common ancestor of genera *Arum*, *Biarum* Schott and *Dracunculus* Mill.) was constrained to a maximum age of 30.2 Mya; (2) the most recent common ancestor of *Arum* and *Dracunculus* was constrained to a minimum age of 27.3 Mya; and (3) the stem group of *Arum* subgenus *Arum* was constrained to a minimum age of 16.1 Mya.

Areas were defined following different studies on the geological and biogeographical history of the Mediterranean basin and surrounding areas (Meulen-kamp & Sissingh, 2003; Mansion *et al.*, 2008; Ree & Sanmartín, 2009), and were set to a number of ten: East European, West European, Apennines, Aegean, Anatolian, Iranian, Arabian, North African, Macaronesian Islands and Caucasus (Fig. 3). The rules applied to define the area for each species were as follows: (1) if the origin of the sample was known, the sample was attributed to the area in which it was sampled; (2) if the origin was unknown, the sample was assigned to the area in which the plant is known to be distributed according to Boyce (1993, 2006) and the search engine of *Flora Europaea* (Flora Europaea, 2009) [in the case of *A. balansanum* R.R.Mill., *A. byzantinum* Schott, *A. sintenisii* (Engl.) P.C.Boyce, *D. vulgaris*, *D. canariensis* and *B. dispar*]; (3) if a sample belonged to a widely distributed and well-described species for which we did not possess samples from all the parts of the distributional area, it was assigned to its region of origin plus the remaining noncovered regions according to Boyce (1993) (only in the case of *A. italicum*).

Table 2. Sequenced regions, with corresponding total number and percentages of base pairs (bp), constant (C), variable (V) and parsimony informative (PI) sites.

| Region | Total (bp) | Constant (C) sites | Variable (V) sites | Parsimony-informative (PI) sites |
|-----------------------|-------------|--------------------|--------------------|----------------------------------|
| <i>3'rps16-5'trnK</i> | 845 (100%) | 791 (93.6%) | 54 (6.4%) | 17 (2.0%) |
| <i>ndhA</i> intron | 1077 (100%) | 1021 (94.8%) | 56 (5.2%) | 24 (2.2%) |
| <i>psbD-trnT</i> | 1024 (100%) | 993 (97.0%) | 31 (3.0%) | 13 (1.3%) |
| <i>rpl32-trnL</i> | 777 (100%) | 730 (94.0%) | 47 (6.0%) | 36 (4.6%) |

Dispersal-vicariance analysis (DIVA) is a method for inferring the most parsimonious reconstruction of ancestral ranges on a given phylogenetic tree by minimizing the number of dispersal and extinction events that are needed to explain the current terminal distributions (Ronquist, 1997). The program DIVA (Ronquist, 2001) uses a three-dimensional cost matrix to estimate the cost of moving from the ancestor to each of the descendants (Ronquist, 1997). It allows two different scenarios for range inheritance at speciation nodes: (1) duplication or within-area speciation, when the ancestor is distributed in a single area and each of the two descendants inherits the entire ancestral range (e.g. A to A); (2) vicariance, when the ancestor occurs in two or more areas and each descendant inherits a nonoverlapping subset of the ancestral range (e.g. AB to A and B). Only one dispersal event per branch (between two ancestral nodes) is allowed in the model, except for terminal branches leading to widespread taxa, for which DIVA postulates multiple dispersal events. To account for polytomies in the 50% majority-rule Bayesian inference tree, five more exceptions were required in our analysis (see below). DIVAs were run with the maximum number of areas allowed at ancestral nodes constrained to two. Uncertainty in phylogenetic relationships was accounted for in DIVA by using an approach proposed by Nylander *et al.* (2008), which integrates DIVA parsimony-based reconstructions over a Bayesian MCMC sample of trees representing the posterior probability of the tree topology (hereafter referred to as Bayes-DIVA). Specifically, we sampled one tree for every 16 trees (7501 in total) from the MCMC 'post-burnin' sample and used R scripts available from the second author to summarize/average ancestral area reconstructions over all sampled trees for each node in the 50% majority-rule Bayesian inference, which was used as the reference. Only those trees containing the node of interest were summarized in estimating the probabilities for that node. This approach allows an estimation of the marginal probabilities of ancestral ranges for a given node whilst integrating over the uncertainty in the rest of the tree topology (Nylander

et al., 2008). Ancestral areas and vicariance/dispersal events were recorded following Buerki (2009).

As several polytomies were found in the 50% majority-rule topology, the following rule was applied to solve incompatibilities between nodes and to estimate correct dispersal-vicariance events (that otherwise would violate DIVA assumptions): if the most probable area for a given node was incompatible (according to DIVA assumptions) with that of the next coming node or tip, it was combined with the following most probable area(s); this was performed until the ancestral areas of the node were congruent with the areas assigned to the following node or tip. In order to summarize the different dispersal events across the three geological epochs spanning the diversification of *Arum* (Miocene, Pliocene and Pleistocene), a pairwise matrix of dispersion was built for each epoch to address the links among the ten defined areas [this was performed using R (R Development Core Team, 2009), with scripts available on request from the second author]. When a branch spanned over more than one epoch, the proportion of the branch over each epoch was considered, and the fraction D of one single dispersal event in a given epoch was recorded ($0 < D < 1$). To summarize the results, arrows with variable widths (proportional to the number of dispersal events) were drawn on palaeogeographical maps corresponding to the three relevant epochs (Meulenkaamp & Sissingh, 2003).

RESULTS

PHYLOGENETIC INFERENCES

The combined dataset consisted of 250 sequences. Aligned lengths were 845 bp for *3'rps16-5'trnK*, 1077 bp for the *ndhA* intron, 1024 bp for *psbD-trnT*, 777 bp for *rpl32-trnL* and 104 binary positions corresponding to coded gaps. The final matrix thus contained a total of 3827 characters (3723 nucleotides and 104 gap presence/absence). Values for C, V and PI for each partition are given in Table 2. Partition *rpl32-trnL* provides slightly more informative sites (36) than the other partitions. The partition homoge-

neity test was passed ($P = 0.07$), indicating that the information provided by the four plastid regions was congruent. Topologies obtained with Bayesian inference and MP algorithms (Fig. 1) were highly congruent (quartet distance of 0.114, meaning that approximately 89% of the components were compatible between the two trees) and defined five major supported clades (see Fig. 1). Topologies depict *A. pictum* as sister to the rest of the genus, confirming the definition of subgenera *Gymnomesium* and *Arum* (supported by BPP = 1 and DI = 10; Fig. 1). In subgenus *Arum*, the first branching clade I includes *A. palaestinum* Boiss. and the different subspecies of *A. dioscoridis* Sibth. & Sm. (BPP = 1, DI = 3). Clade II contains *A. concinnatum* Schott and *A. italicum* (for which monophyly was not contradicted in the Bayesian analysis) (BPP = 0.95, DI = 3). Clade III includes the two most eastern taxa, *A. jacquemontii* Blume and *A. korolkowii* Regel, *A. megobrebi* Lobin, M. Neumann, Bogner & P.C. Boyce, *A. rupicola* Boiss. and the two easternmost samples of *A. maculatum* L. included in this study, and a new species, hereafter referred to as *A. sp. nov.* (BPP = 0.99, DI = 1). In the MP topology, *A. creticum* Boiss. & Heldr. is also included in clade III as the first branching lineage (Fig. 1). Clade IV includes two subclades containing *A. sintenisii* P.C. Boyce and *A. hygrophilum* Boiss., on the one hand, and *A. byzantinum* Schott, *A. nigrum* Vell. and some specimens of *A. elongatum* Steven, on the other (BPP = 0.84, DI = 1). In the Bayesian inference topology, *A. idaeum* Coust. & Gand. is also included in clade IV as the first branching lineage (Fig. 1). Finally, clade V is poorly resolved and includes the remaining taxa: *A. maculatum* (western samples), *A. cylindraceum* Gasp., *A. orientale* M. Bieb., *A. besserianum* Schott, *A. balansanum* R.R. Mill, *A. purpureospathum* P.C. Boyce, *A. euxinum* R.R. Mill, *A. apulum* (Carano) P. Boyce, *A. cyrenaicum* Hruby and one representative of *A. elongatum* (BPP = 1, DI = 2). The relative position of clade V swapped depending on the phylogenetic algorithm, as it was sister to clade IV in the Bayesian inference tree, but sister to clade III in the MP tree (Fig. 1). Incongruence between the two topologies concerned (1) the positions of clade V relative to clades III and IV, and (2) the branching of *A. idaeum* and *A. creticum*.

Sections as defined by Boyce (1989) were not supported by the phylogenetic hypotheses. Except for the cases of subsections *Poeciloporphyrochiton* (clade I, *A. dioscoridis* and *A. palaestinum*) and *Tenuifila* (subclade in clade III, *A. jacquemontii*, *A. korolkowii* and *A. rupicola*), our topologies did not support the current infrageneric delimitation (Fig. 1, Table 1). Finally, the monophyly of several widespread species (e.g. *A. elongatum* and *A. maculatum*) was not supported (see Fig. 1).

CHARACTER EVOLUTION

The reconstruction of ancestral states for the four studied characters is shown in Figure 2. The trait that appears to be most constrained from the phylogenetic reconstruction is ploidy (Fig. 2A), with an ancestral character state corresponding to diploidy ($2n = 2x = 28$) and one single evolution towards hexaploidy. Tetraploidy evolved several times. The remaining characters (Fig. 2B, tuber shape; Fig. 2C, flower shape; Fig. 2D, spathe/spadix ratio) show a pattern of multiple independent events and are much less informative at the infrageneric level. An exception could be the evolution of the rhizomatous tuber shape which, although largely symplesiomorphic, seems to be correlated with the level of ploidy.

BIOGEOGRAPHICAL ANALYSIS

Reconstructed ancestral areas for the different nodes corresponding to the 50% majority-rule Bayesian tree are shown in Figure 3. The two most probable ancestral areas from the crown nodes of the genus were the Aegean and West European regions. Later nodes show that the Aegean and Anatolian regions were the only areas to harbour ancestral lineages of the genus for a long time. Overall, a substantial proportion of the dispersion of *Arum* lineages towards their current distribution areas seems to have happened after the late Miocene (c. 10 Mya).

The rates and direction of dispersal events at three different time-slices corresponding to the Miocene, Pliocene and Pleistocene are shown in Figure 4. During the Miocene (23–5.3 Mya, Fig. 4A), dispersion mainly happened from the Aegean area to Anatolia. Exchanges were also possible between the newly emerging Caucasus region and the Aegean and Anatolian areas. Colonization of the Iranian area seems to have happened only during this period. Colonization of Macaronesia also occurred at this time, but other dispersals towards this area were probably also possible during the early Pliocene (5.3–2.6 Mya, Fig. 4B). During this epoch, the genus extended its distribution for the first time onto the Arabian plate, and important dispersion events seem to have happened from the Aegean (and, to a lesser extent, from the Anatolian region) to Eastern Europe. The North African region was colonized during the most recent geological epoch (Pleistocene, 2.6 Mya–present, Fig. 4C) probably via two pathways: (1) from the Apennines through the southern tip of the Italian Peninsula; and (2) from the Arabian region through the Gulf of Suez. Numerous dispersals also occurred from the Aegean to the Anatolian area in the Pleistocene and from the Apennine region to Western and Eastern Europe and the Aegean. During this last epoch, exchanges seemed to have halted between the

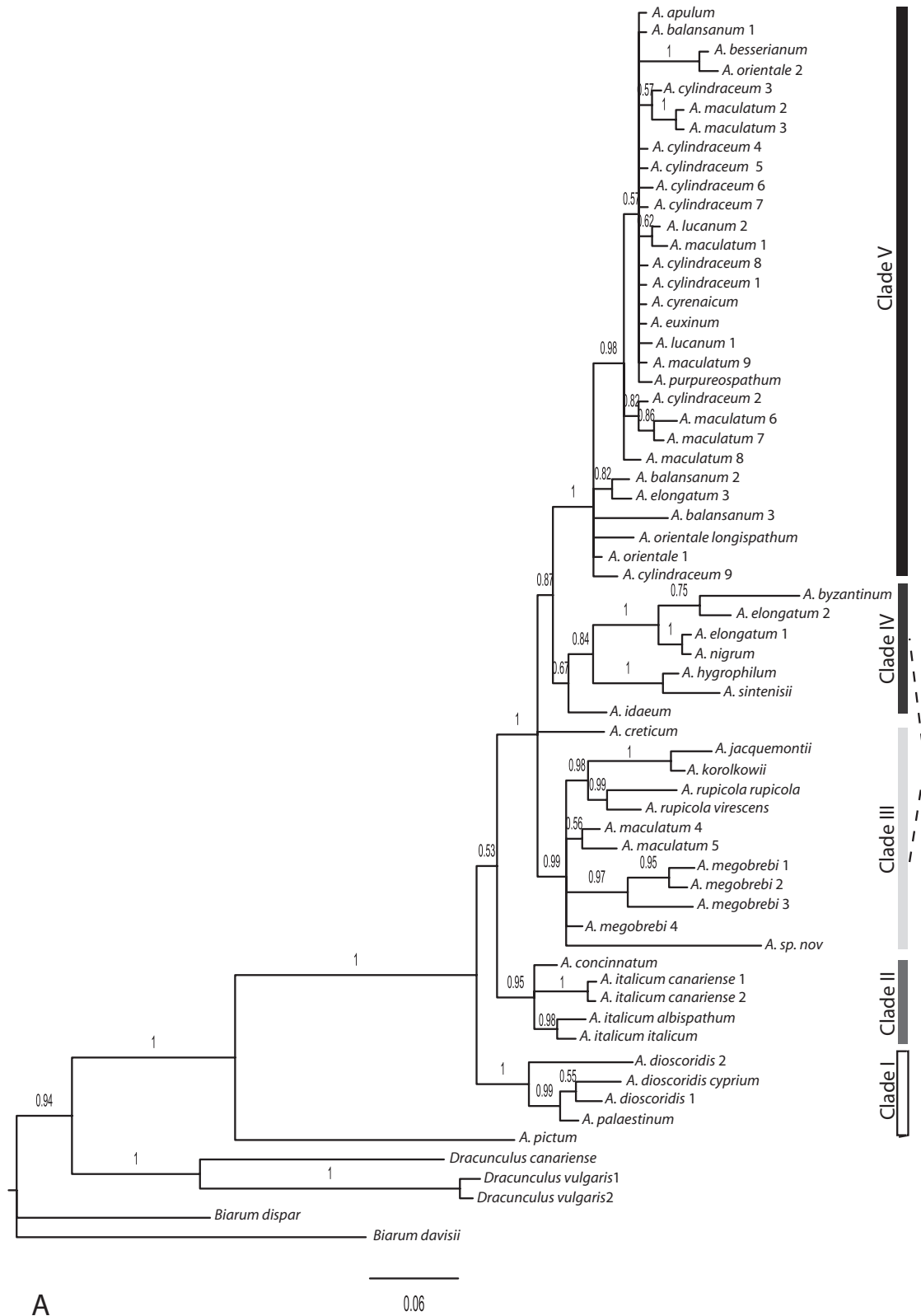


Figure 1. Inferred plastid phylogenies: A, Bayesian inference, half-compatible tree; B, maximum parsimony (MP), strict consensus tree. Values shown on the branches represent Bayesian posterior probability values (A) and decay indices (B). Vertical bars indicate major clades explained in the text.

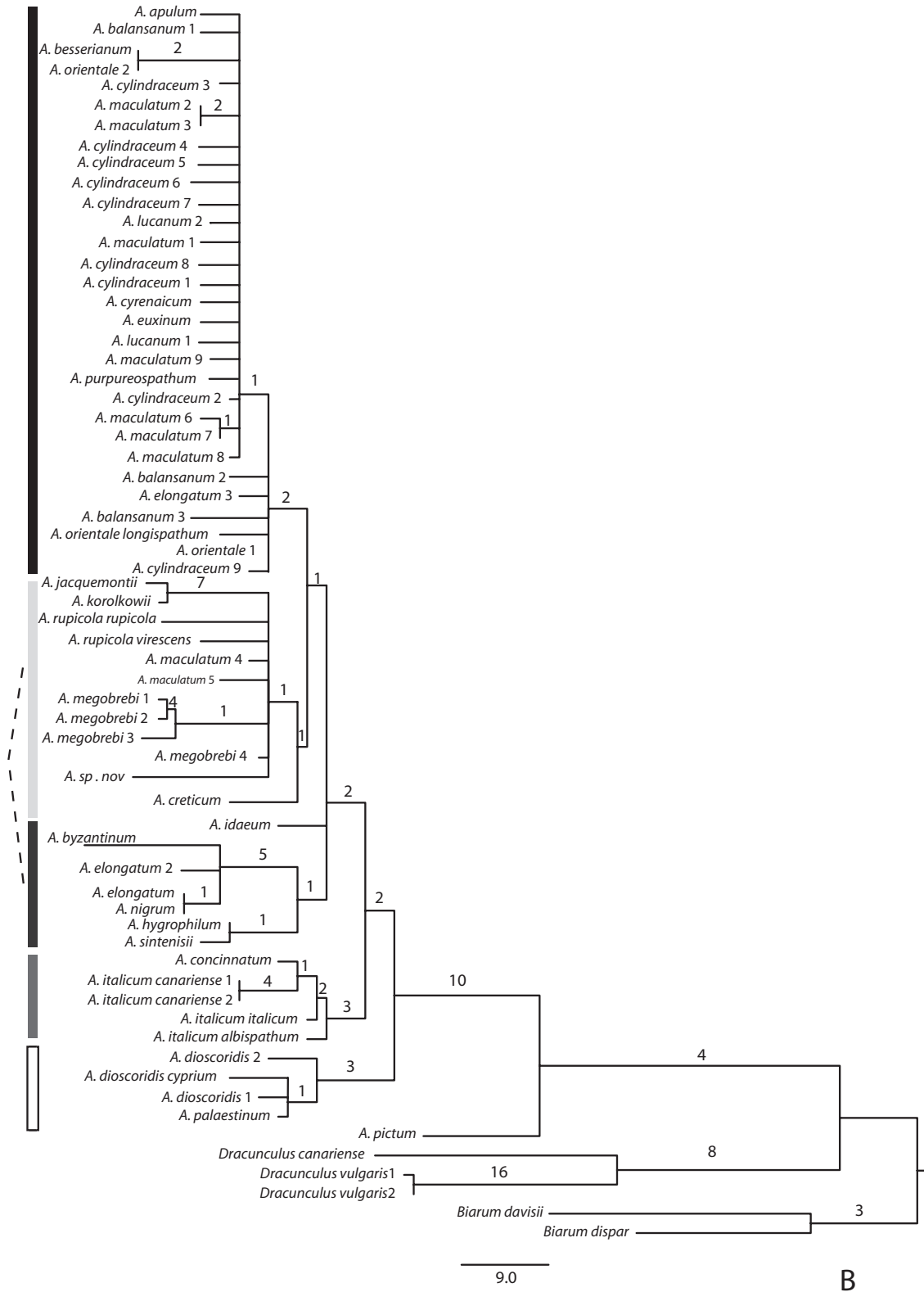


Figure 1. Continued

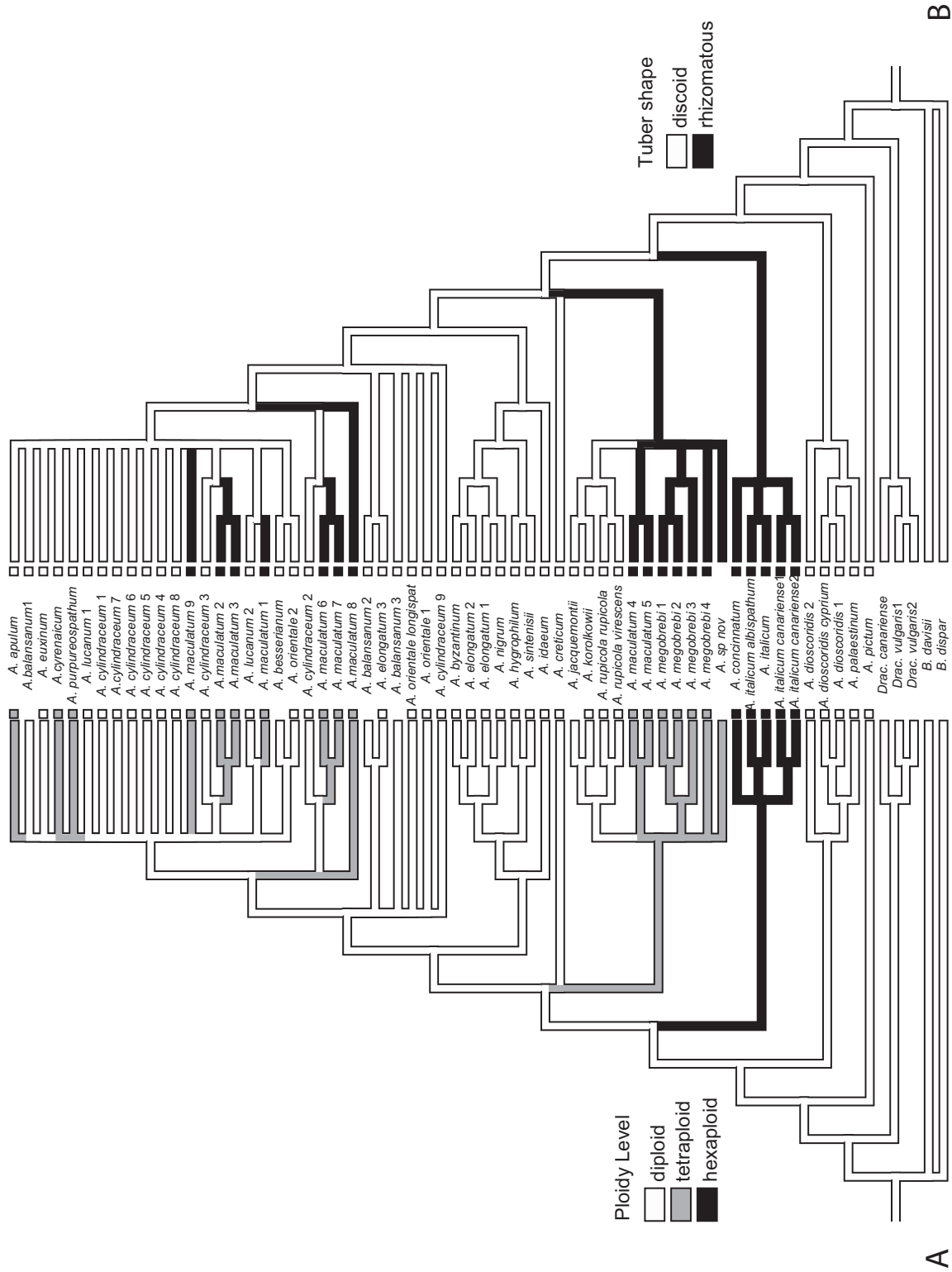


Figure 2. Evolution of categorical characters on the Bayesian topology: A, ploidy (2x, 4x, 6x); B, tuber shape (rhizomatous, discoid); C, flower type (flag, cryptic); D, spadix/spathe ratio (0–0.5; 0.5–1; > 1). Colours are explained in the legend of each figure. Missing squares at the level of terminal taxa indicate unknown characters.

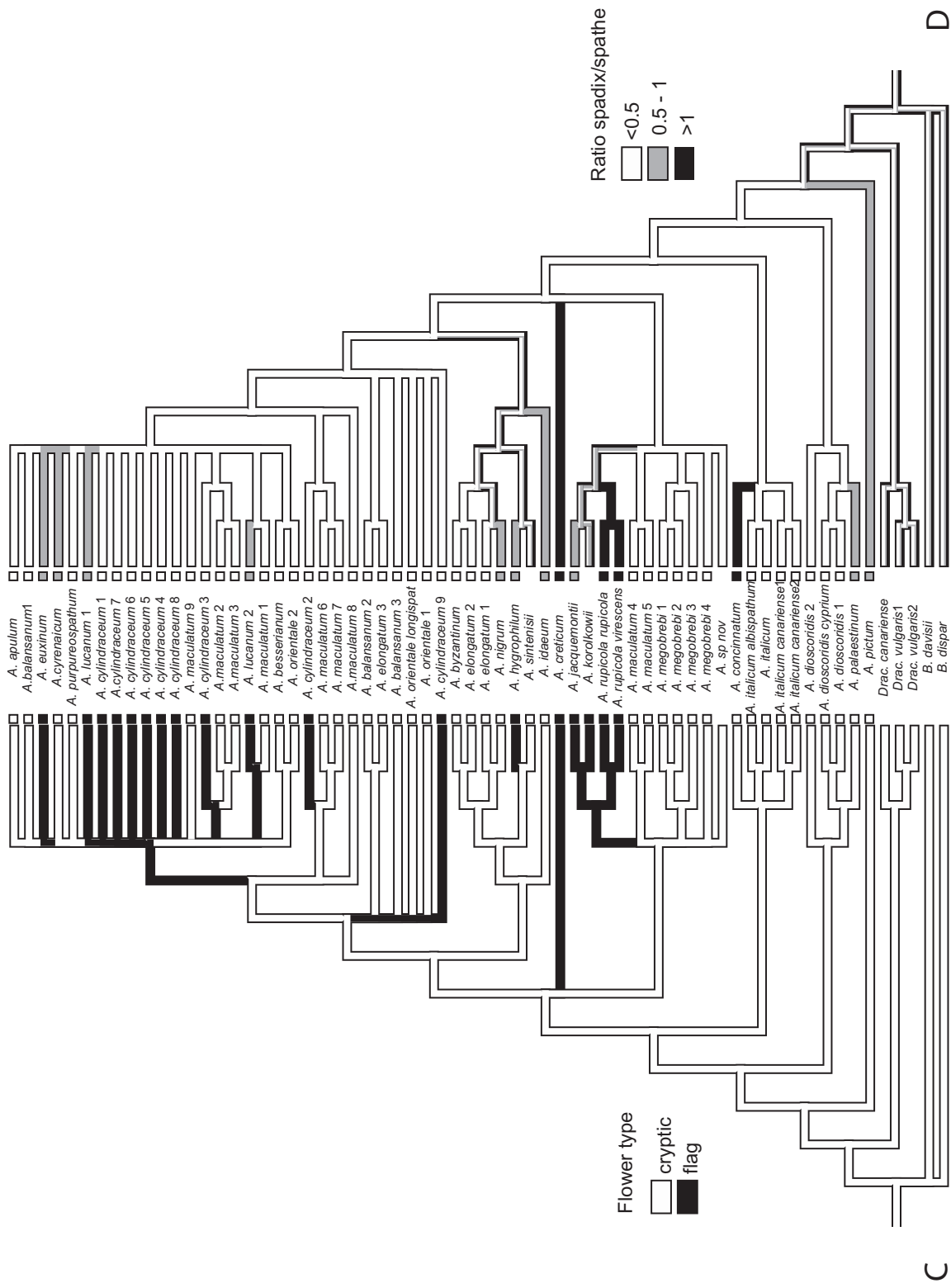


Figure 2. Continued

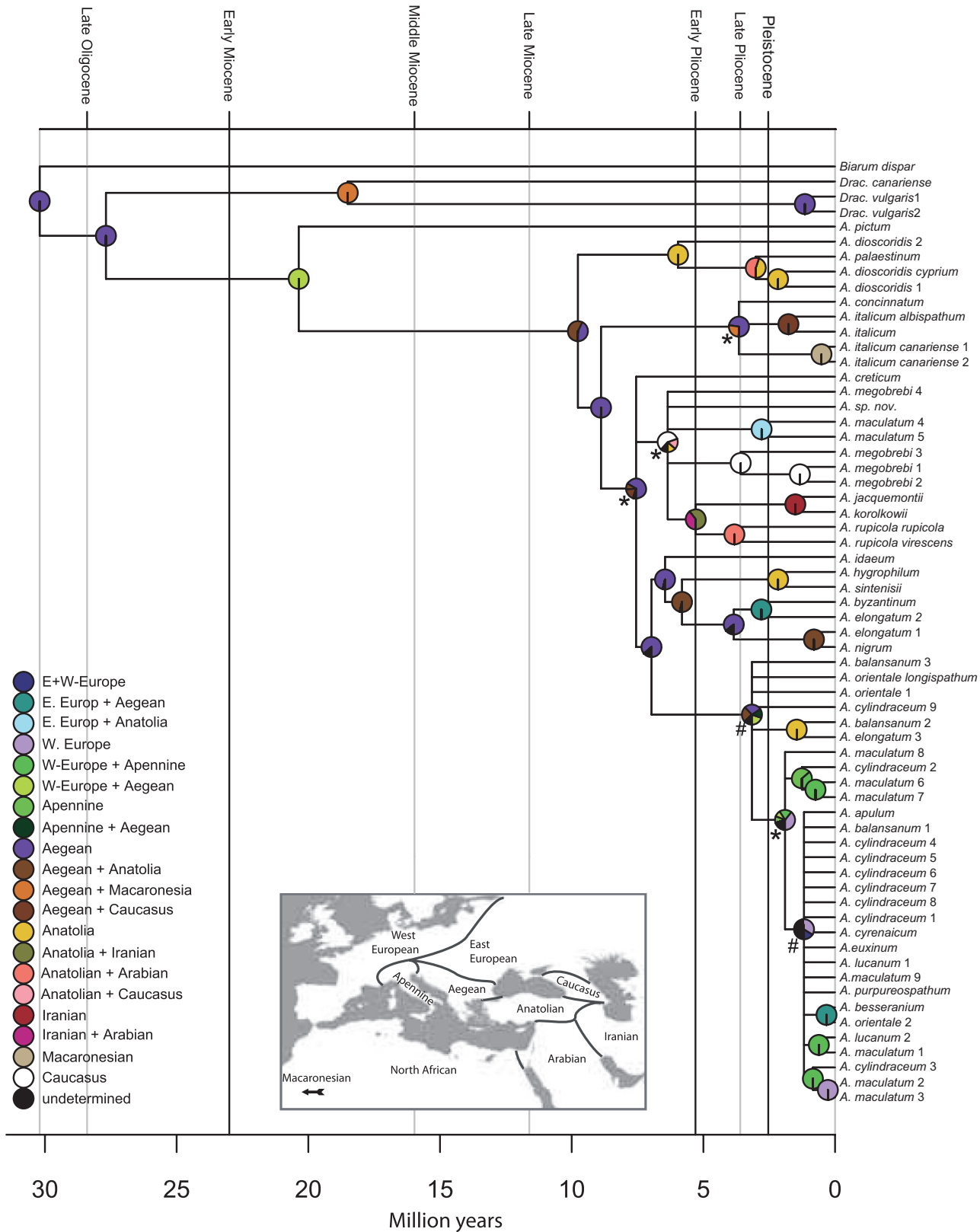


Figure 3. Ancestral areas assigned by dispersal-vice analysis (DIVA) to each node of the Bayesian topology. Colours represent ancestral areas (see legend). All areas with a probability < 0.1 were pooled and treated as a single undetermined area (in black). Nodes having been treated with special rules are indicated by “*” or “#”: “*” indicates that the second most probable area has been combined with the first in order to sketch compatible scenarios; “#” indicates that more than two ancestral areas have been combined to obtain the congruence of the nodes and tips. Scale corresponds to million years from present. Map shows areas defined for the biogeographical inference.

Aegean and Eastern Europe, despite the neighbouring position of these two regions.

DISCUSSION

INFRAGENERIC RELATIONSHIPS AND SPECIES’ IDENTITY

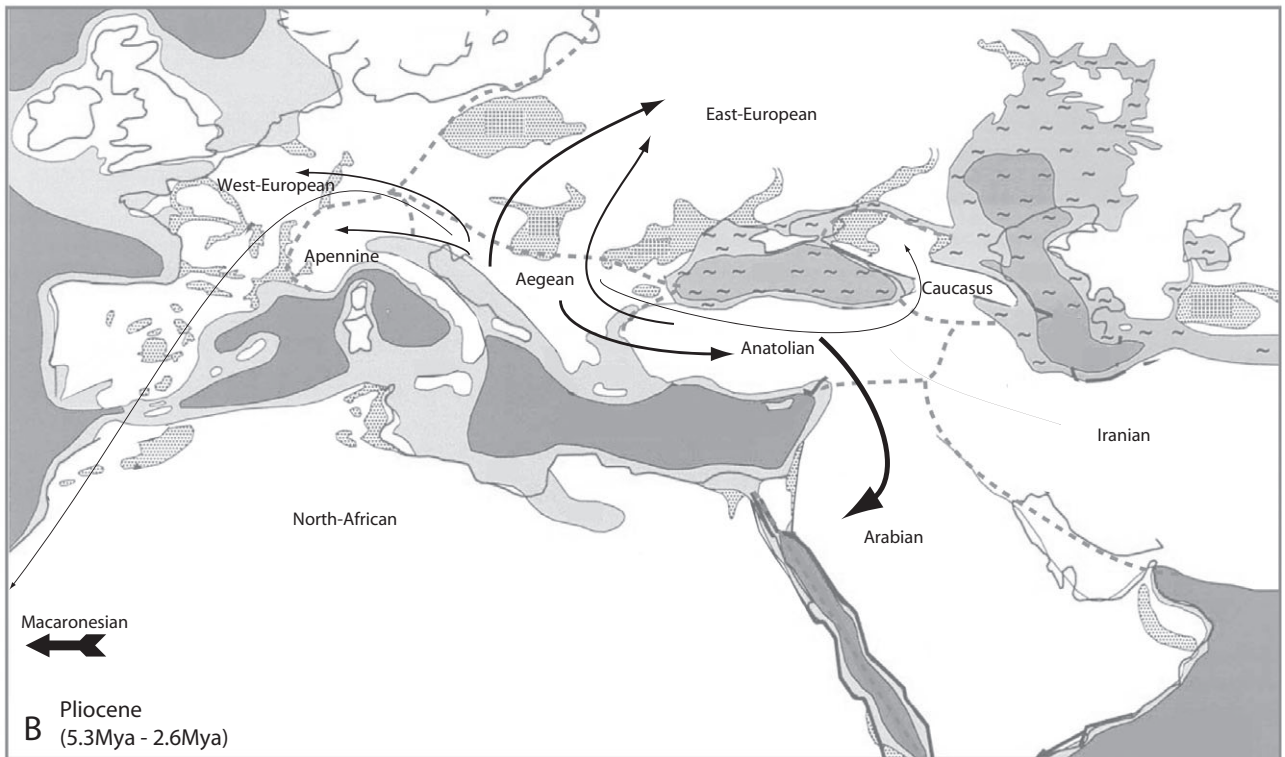
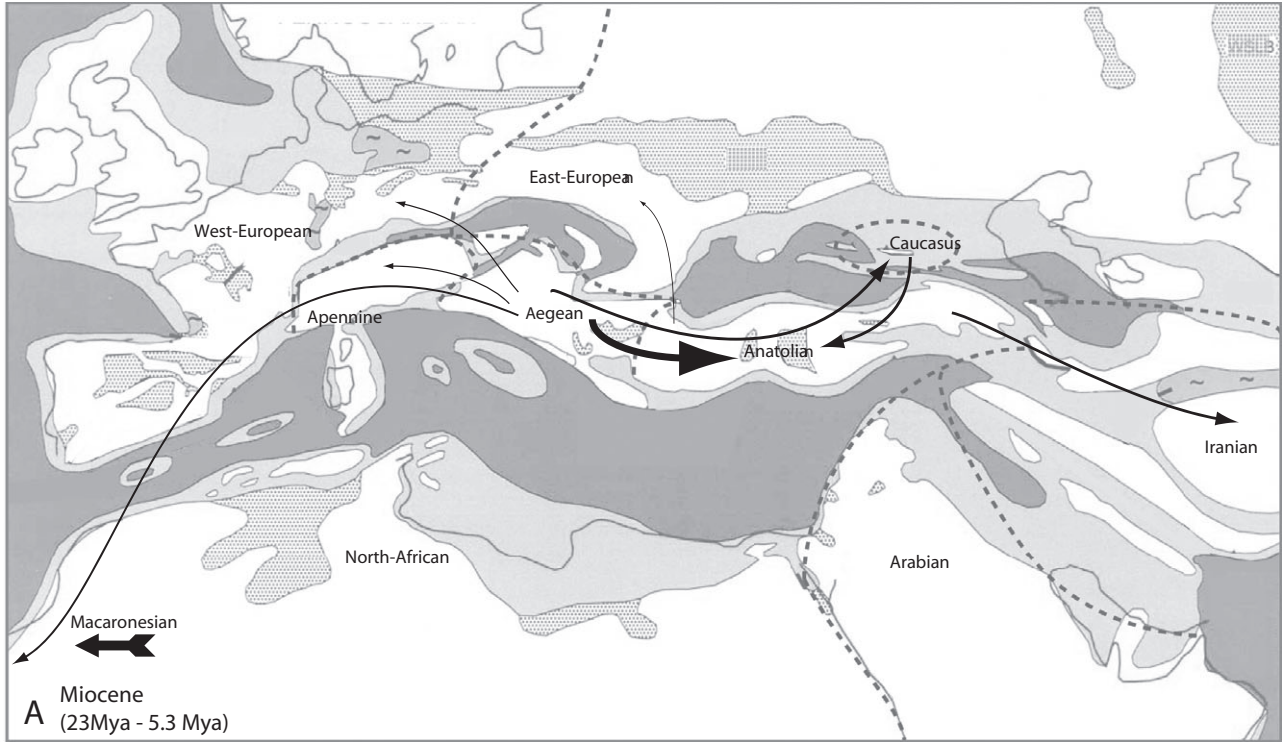
Classical taxon definition and circumscription in the genus *Arum* (Boyce, 1993, 2006) only partially match our phylogenetic hypothesis. As shown in Figure 1, the identity of sections, subsections (*sensu* Boyce, 1989, 1993) and species is strongly challenged, and it seems obvious that there is a ‘gap’ between the current classification and the genetic identities of the taxa. Our analyses, however, confirm the validity of the two subgenera, as *A. pictum* (subgenus *Gymnomesium*) is the first branching lineage of *Arum* (as shown previously by Mansion *et al.*, 2008). This result is supported by floral (staminodes present, but no pistillodes) and phenological (flowering in autumn and not in spring as in the rest of the species) characters restricted to *A. pictum*. For more than a century, the peculiarities of this Hercynian endemic have been recognized, and several authors have attempted to place it in a different genus (*Gymnomesium* Schott, 1855). The long branch separating this lineage from the other species (with a divergence estimated to be sometime between the early and middle Miocene; Fig. 3) favours the hypothesis of a palaeorelictual identity of *A. pictum* (as proposed by Mansion *et al.*, 2008). Within subgenus *Arum*, only subsection *Poeciloporphyrachiton* (Fig. 1, clade I) is retrieved by our phylogenetic hypothesis. It appears as the first branching lineage in the subgenus, confirming that *A. dioscoridis* and *A. palaestinum* are closely related and placed in an external position, as proposed formerly by several authors (Hruby, 1910; Boyce, 1989, 1993). Another exception could be subsection *Tenuifila*, which is nested in clade III in the Bayesian topology, and might still be considered as a valid entity (see below). No other subsection is compatible with our results.

Both Bayesian and MP topologies argue in favour of the monophyly of hexaploid taxa (Fig. 1, clade II; Fig. 2A), with *A. italicum* specimens clustering together with *A. concinatum*. The insular *A. italicum* ssp. *canariense* (Webb & Berthel.) P.Boyce is genetically differentiated from the ‘continental’ sub-

species from which it diverged during the Pliocene (Fig. 3). As the monophyly of *A. italicum* is not retrieved in the MP analysis, a more thorough analysis (e.g. using genomic screening markers) should be performed to confirm the status of *A. concinatum*.

Although the phylogenetic relationships among the three remaining clades (III, IV and V) are not yet resolved (i.e. the topology varies according to the phylogenetic algorithm), their respective monophyly is relatively well supported with $DI \geq 1$ and $BPP > 0.95$ (with the exception of clade IV, which shows a lower support of 0.84; see Fig. 1). Current molecular data do not allow the discussion of the phylogenetic relationships of *A. creticum* and *A. idaeum*, two species with overlapping distributions in Crete, which are either placed in a polytomy at the base of these three clades or as the first branching lineages of clade III (MP topology) and clade IV (Bayesian topology), respectively. These species are morphologically similar (open floral chambers, sweet or weak odour production vs. closed floral chamber and clear lure-oriented odour production in the other species) and were included in subsection *Cretica* (Boyce, 1989). Relationships among clades III, IV and V should also be examined carefully as our results do not allow conclusions to be drawn regarding the position of clade V, as it appears as either sister to clade III (MP topology) or to clade IV (Bayesian inference tree).

The strongly supported clade III (excluding *A. creticum*; $BPP = 0.99$; $DI = 1$) comprises all members of subsection *Tenuifila* (i.e. *A. rupicola*, *A. jacquemontii* and *A. korolkowii*, which form a well-supported monophyletic group in the Bayesian topology; $BPP = 0.98$) and all representatives of *A. megobrebi*, two specimens of *A. maculatum* from the easternmost edge of the distribution and one sample from the Caucasus area. The latter should be considered as a new species (referred to as *A. sp. nov.*). The placement of *A. maculatum* samples in clade III is unexpected as the two specimens found here are highly divergent phylogenetically with respect to the European representatives (clade V) of this widely distributed species (Fig. 1). As a consequence, they might merit treatment as a different species if further morphological studies confirm this status by identifying synapomorphies. Clade III has a biogeographical coherence as the taxa included are found in the eastern part of the



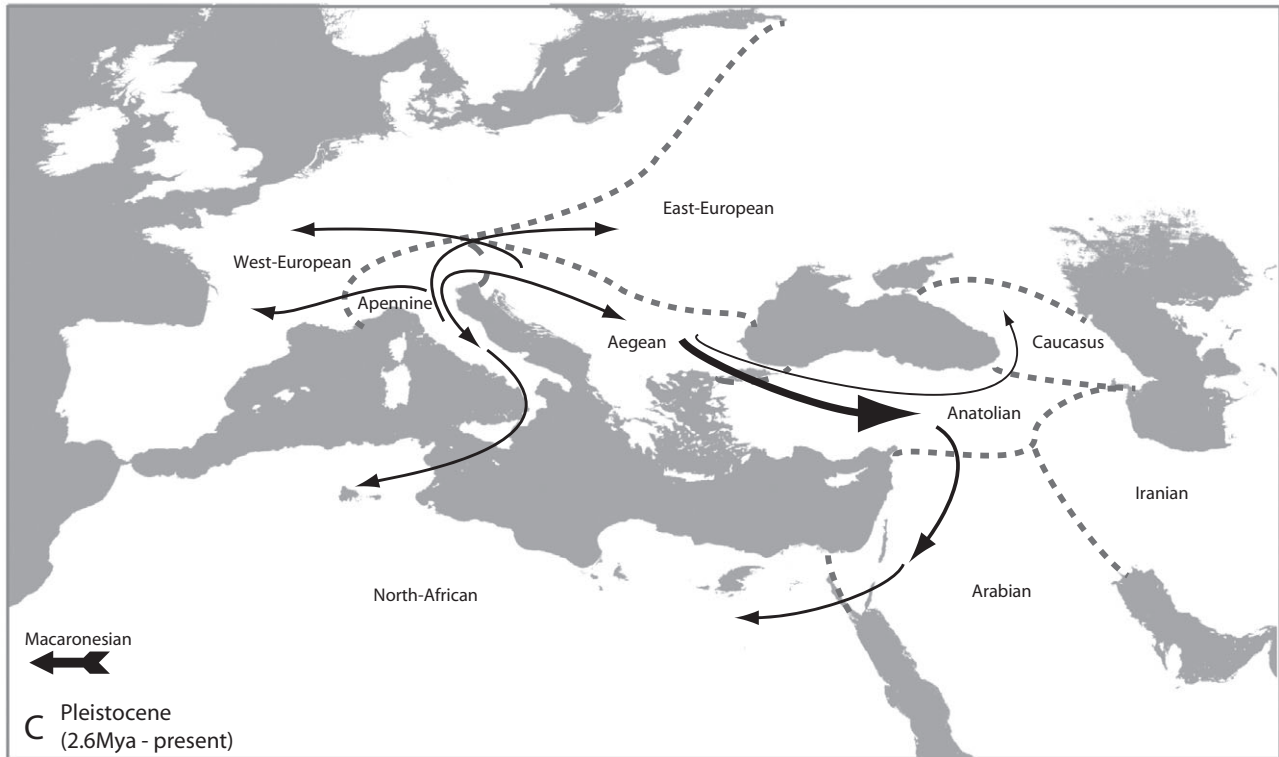


Figure 4. Dispersion events at three time slices: A, Miocene; B, Pliocene; C, Pleistocene (maps A and B, modified from Meulenkamp & Sissingh, 2003; with permission of the editors). The widths of arrows are proportional to dispersal rates. Broken lines indicate boundaries between biogeographical zones.

distribution area of the genus (from northern Turkey to the Iranian region), confirming the role played by this area in the diversification of *Arum* (as proposed previously by Bedalov & Küpfer, 2005).

Clade IV (excluding *A. idaeum*; BPP = 0.84; DI = 1) comprises two specimens of *A. elongatum*, which appears to be paraphyletic with respect to *A. nigrum*, and possibly *A. byzantinum* (in the Bayesian topology), although the position of the latter is not well supported (Fig. 1). More generally, the morphology typical for *A. elongatum* seems to be quite labile as one specimen is also found in clade V (see below). The other representatives of this clade are the closely related *A. sintenisii* (endemic to Cyprus) and the oriental *A. hygrophilum*, the former probably having diverged from the latter in the late Pliocene after a dispersal followed by an insular differentiation. This result was already predicted by Boyce (2006).

Finally, clade V is by far the least resolved, encompassing closely related taxa that diverged during the second half of the Pliocene and the Pleistocene (Fig. 3), most having colonized the Apennines and temperate habitats in Western and Eastern Europe (Fig. 4). In this clade, differentiation among specimens is weak and all species sampled more than once are paraphyletic (Fig. 1). Uncovering the relation-

ships among taxa within this clade would require further analyses based on, for example, genomic screening. This might help to address the status of widely distributed taxa, such as *A. cylindraceum* and the 'European' *A. maculatum*, with that of narrow endemics such as *A. apulum* and *A. purpureospathum*. Another case of interest is the well-supported group composed of *A. besserianum* and one specimen of *A. orientale* (BPP = 1; DI = 2; Fig. 1). *Arum orientale* is still poorly defined as attested by successive revisions during the last 15 years (Boyce, 1994, 2006; Bedalov & Küpfer, 2005). This taxon was first described as a species with several subspecies, present in Crimea and extending to the eastern part of the Balkans (Boyce, 1993). However, several morphological characters point to a close relationship with *A. besserianum* distributed in Ukraine and Poland (P. Küpfer, pers. observ.). Consequently, this taxon certainly encompasses different paraphyletic lineages, and both its status and that of other taxa (e.g. *A. balansanum*) should be investigated using novel genomic techniques coupled with taxonomy.

Therefore, we have observed that two different patterns arise when testing the monophyly of species for which more than one specimen was collected. On the one hand, some are well supported by our analyses:

this is the case for *A. dioscoridis* (clade I), *A. rupicola* (clade III) and, to a lesser extent, *A. megobrebi* (clade III) (Fig. 1). On the other hand, some species are clearly polyphyletic (comprising specimens from lineages that diverged as early as the Pliocene), such as, for instance, *A. maculatum* (clades III and V) and *A. elongatum* (clades IV and V) (Fig. 1). Finally, the case of *A. italicum* (clade II) is somewhat intermediate, as the monophyly is not contradicted by the Bayesian topology, whereas the species is paraphyletic (with the inclusion of *A. concinatum*) in the MP topology.

CHARACTER EVOLUTION: CHARACTERIZING THE IDENTITY OF THE MOST RECENT COMMON ANCESTOR

Among all the investigated characters, only ploidy seems to be related to the evolution of the genus (Fig. 2A). The remaining traits (spathe/spadix ratio, flower type, shape of the tuber) show patterns of multiple independent evolution and a looser correlation with the evolutionary history of *Arum*. Our results therefore support the hypothesis that the level of ploidy might constitute an informative character for the systematics of the genus, as first proposed by Bedalov & Küpfer (2005), and address the diploid status of the most recent common ancestor (a trait shared by both *A. pictum* and taxa within clade I). However, the abrupt transition from diploidy to hexaploidy (in clade II) seems to be unlikely and might require the existence of a transitional and yet extinct or undiscovered tetraploid form. The advantages of polyploids in terms of survival have been addressed recently in *Arum*, as artificial crossings between distinct species yielded polyploid hybrids that were 'robust and maintain themselves in cultivation without apparent difficulties' (Bedalov & Küpfer, 2005). Interestingly, the fact that clade V comprises both a substantial proportion of tetraploid lineages and an important number of recognized taxa could support the idea of an increased fitness in polyploids, facilitative for the radiation of this group (Fig. 3) (for a review of the ability of polyploids to colonize a wider range of habitats, see Prentis *et al.*, 2008). However, before arriving at any conclusion, it is important that the phylogenetic relationships among the specimens of this clade are clarified.

One local phylogenetic constraint on the flower type (flag vs. cryptic) was addressed in the subclade corresponding to the *Tenuifila* subsection, with all taxa sharing a flag flower, whereas the ancestral state for this trait within *Arum* was a cryptic flower. As there is a strong association between cryptic flowers and attract-and-lure pollination strategies (Boyce, 1989; Gibernau *et al.*, 2004), the latter should be considered as the ancestral pollination mode in the genus. It is important, however, to mention that the polyphyletic

status of this character is not surprising, as it is related to reproductive structures, which, in *Arum*, appear to be highly correlated with fast-evolving pollination syndromes (Chouteau, Gibernau & Barabé, 2008). The evolution of this character would thus reflect more strongly the ecological processes that species have independently undergone rather than the evolutionary history of the genus.

Finally, our results confirm that the ancestral state of the *Arum* tuber shape was discoid, as proposed by Bedalov & Küpfer (2005), and that the appearance of the first rhizomatous species happened at the same time as the transition from diploidy to hexaploidy (clade II, Fig. 1). Although the transition from a discoid to a rhizomatous tuber occurred several times, there seems to be a trend towards a correlation between ploidy and tuber shape: all rhizomatous species are polyploid. In contrast, several polyploid species (*A. apulum*, *A. cyrenaicum* and *A. purpleospathum*) have discoid tubers. As sections within *Arum* were classically defined on the basis of this homoplasious character, there is a strong need to consider morphological characters from other plant parts to build a new classification of *Arum* compatible with our molecular evidence.

ARUM THROUGH SPACE AND TIME

Bayes-DIVA provides strong support for an Aegean/Western European origin of the genus (Fig. 4) sometime in the early Miocene (*c.* 20 Mya). However, assuming that the earliest diverging lineage (now composed of only *A. pictum*) originated and survived in the Hercynian islands long before all other *Arum* spp. arose (according to the palaeorelictual hypothesis proposed by Mansion *et al.*, 2008), the ancestral area corresponding to the rest of the genus is the Aegean region (Fig. 3). This zone has acted as a natural laboratory allowing the diversification of lineages, sometime in the late Miocene (Figs. 3, 4). The Aegean also appears to be a main source of dispersal events throughout the evolutionary history of *Arum*. Its central position with respect to the other areas in which the genus is present today could have facilitated this. Most dispersal events recorded during the middle to late Miocene occurred from the Aegean to the Anatolian region (Fig. 4A). Later, the emergence of the Iranian plate allowed its colonization once a land-bridge was established with the Anatolian plate in the late Miocene (Meulenkaamp *et al.*, 2000). During this period, no dispersals were observed towards the Arabian plate that was still isolated from the northern lands by a marine transgression (Meulenkaamp & Sissingh, 2003). Once the Caucasian archipelago emerged (and possibly after its uplift and contact with the Northern Anatolian region), further dispersals to

and from this region occurred in the late Miocene. At this same period, long-distance dispersals are recorded from the Aegean to the Macaronesian regions.

The first dispersal to the Arabian zone occurred more recently, during the Pliocene, when a sea regression (Meulenkamp & Sissingh, 2003) allowed this land to come into contact with the Anatolian region (Fig. 4B). The regression of the western Para-Tethys (following the uplift of the European plates) could also have permitted the dispersal from the Aegean to Eastern Europe. Exchanges continued between the Aegean and Anatolia and through these two zones to the Caucasus and Eastern Europe.

During more recent times (Pleistocene), new dispersals from the Aegean to Anatolia were recorded (Fig. 4C), probably facilitated by the Mediterranean regressions characterizing Quaternary climatic oscillations (Peulvast *et al.*, 2000). At this time, North Africa was colonized twice through the Arabian plate and through the Apennines (Fig. 4C).

Although exchanges mainly occurred longitudinally (east–west) across land paths at the periphery of the seas during the early history of the genus, the pace of dispersion and diversification in *Arum* increased after the peri-Mediterranean region was unified (i.e. after the emergence of the Arabian and Iranian plates, the uplift of the Caucasus and the regressions of Tethys and Para-Tethys) (Meulenkamp *et al.*, 2000; Meulenkamp & Sissingh, 2003).

The colonization of Macaronesia requires special treatment. The taxon inhabiting this region (*A. italicum* ssp. *canariense*) appears to have arrived there during the late Miocene or early Pliocene, in agreement with the timing of colonization already observed in several other endemic taxa of these islands (Carine *et al.*, 2004). This ancient dispersal contrasts with the more recent colonization of North Africa. This discordance could be a result of either a first colonization of North Africa, having allowed the dispersal towards Macaronesia through mid-distance dispersal and further extinction of this lineage, or a long-distance dispersal directly from the Northern Peri-Tethys. Considering the morphology of the seeds (Mayo *et al.*, 1997), this latter hypothesis could be possible only in association with animals; birds have already been proposed as the main dispersers of *A. maculatum* (Snow & Snow, 1988), which could also be true for *A. italicum* (Méndez, 1997).

TOWARDS A NEW CLASSIFICATION OF *ARUM*?

Because our phylogenetic reconstruction strongly contradicts the current systematics of the genus, the need for a new classification is evident (i.e. a large number of homoplasies are suggested by the tracing of the characters currently used in the delimitation of

sections and species in our topology; Fig. 2). However, we recommend caution in formally proposing a new infrageneric classification until nonmolecular synapomorphies supporting the main clades are identified.

The two current subgenera, *Arum* and *Gymnomesium*, are supported by our analyses, although the status of the latter might be reconsidered, given the high level of phylogenetic differentiation of this monospecific subgenus. The characteristic morphology, development, distribution and, as shown in this study, phylogenetic position displayed by *A. pictum* could indicate that it would be more correct to place it in the monospecific genus *Gymnomesium* (as formerly proposed by Schott, 1855).

The two formerly defined sections within subgenus *Arum* are not supported by the phylogenetic analyses and, based on molecular evidence, we recommend a division of the subgenus into five sections (corresponding to clades I–V). Subsection *Poeciloporphyochiton* (corresponding to clade I) should be elevated to the rank of section, whereas new synapomorphies should be recovered for the other clades. The sectional classification of *A. creticum* and *A. idaeum* should also be investigated more thoroughly as our phylogenetic hypotheses only weakly associate them with clades III and IV, respectively. Although this study demonstrates the importance of ploidy as a putative synapomorphy in the case of clade II, a broad survey of morphological characters is strongly recommended.

Finally, the paraphyletic status of widespread species requires additional analyses to be performed with more variable markers in order to validate these findings. Nonetheless, our results already argue for a revision of species such as *A. maculatum* and *A. elongatum* in which major splits have been identified. Future taxonomic revisions should carefully consider characters not related to pollination, as lineages seem to be able to adapt quickly to changes in pollinator availability, leading to floral character convergence in distinct clades.

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NOTE ADDED IN PROOF

The authors of this study attest that at the time of the acceptance of our manuscript in *Botanical Journal of the Linnean Society* no phylogenetic studies of the genus *Arum* (Araceae) had been published. In the meantime, Linz *et al.* have published a phylogeny of *Arum* in the latest issue of *Taxon* (59: 405–415), i.e. two months before the release of our study. These studies were conducted independently and, although sharing the same group of interest, the sampling, approach and conclusions are consequently slightly different. For the sake of the understanding of the evolution of *Arum*, we encourage readers to compare the present study with that of Linz *et al.*

Linz J, Stökl J, Urru I, Krügel T, Stensmyr MC, Hansson BS. 2010. Molecular phylogeny of the genus *Arum* (Araceae) inferred from multi-locus sequence data and AFLPs. *Taxon* **59**: 405–415.

APPENDIX 1

Sample sources and GenBank accession numbers for all analysed samples. BG-Basle, Botanical Gardens, Basle, Switzerland; RBG-Kew, Royal Botanic Gardens, Kew, UK; pr. coll., private collection; NA, not available. Source information is provided in the following order: collector name, collection, voucher reference.

| Sample | Source | Country of origin | GenBank accessions | | | |
|--|---|-------------------|--------------------|-------------|------------------|-------------------|
| | | | <i>3rps16-trnK</i> | <i>ndhA</i> | <i>psbD-trnT</i> | <i>rpl32-trnL</i> |
| <i>Arum apulum</i> | Chase, RBG-Kew, 11022K | Italy | GU370965 | GU371038 | GU371101 | GU371165 |
| <i>Arum balansanum1</i> | Chase, RBG-Kew, 11009K | Turkey | GU370966 | GU371049 | GU371112 | GU371174 |
| <i>Arum balansanum2</i> | Küpfer, pr. coll., 09.06.01 4 | Turkey | GU370967 | GU371065 | GU371128 | GU371189 |
| <i>Arum balansanum3</i> | Koenen, Haller, pr. coll., NA | Turkey | GU370968 | GU371074 | GU371138 | GU371198 |
| <i>Arum besserianum</i> | Bedalov, pr. coll., 1507 | Ukraine | GU370969 | GU371061 | GU371124 | GU371186 |
| <i>Arum byzantinum</i> | Küpfer, pr. coll., 09.06.02 1 | Turkey | GU370970 | GU371063 | GU371126 | GU371188 |
| <i>Arum concinnatum</i> | Chase, RBG-Kew, 11014K | Crete | GU370973 | GU371060 | GU371123 | GU371185 |
| <i>Arum creticum</i> | Chase, RBG-Kew, 11037K | Crete | GU370974 | GU371068 | GU371132 | GU371193 |
| <i>Arum cylindraceum1</i> | Chase, RBG-Kew, 11013K | Romania | GU370982 | GU371028 | GU371091 | GU371155 |
| <i>Arum cylindraceum2</i> | Espindola & Revel, pr. coll., cylGIO1 | Italy | GU370975 | GU371080 | GU371144 | GU371204 |
| <i>Arum cylindraceum3</i> | Espindola & Revel, pr. coll., cylGYL3 | Denmark | GU370976 | GU371081 | GU371145 | GU371205 |
| <i>Arum cylindraceum4</i> | Espindola & Revel, pr. coll., cylMUS4 | Poland | GU370977 | GU371082 | GU371146 | GU371206 |
| <i>Arum cylindraceum5</i> | Espindola & Revel, pr. coll., cylPAD5 | Romania | GU370978 | GU371083 | GU371147 | GU371207 |
| <i>Arum cylindraceum6</i> | Espindola & Revel, pr. coll., cylSIS5 | France | GU370979 | GU371084 | GU371148 | GU371208 |
| <i>Arum cylindraceum7</i> | Espindola & Revel, pr. coll., cylTRA4 | Austria | GU370980 | GU371085 | GU371149 | GU371209 |
| <i>Arum cylindraceum8</i> | Espindola & Revel, pr. coll., cylTRP3 | Macedonia | GU370981 | GU371086 | GU371150 | GU371210 |
| <i>Arum cylindraceum9</i> | Neumann, pr. coll., I21/05 | Sicily | GU371026 | GU371073 | GU371137 | - |
| <i>Arum cyrenaicum</i> | Chase, RBG-Kew, 11030K | Libya | GU370983 | GU371072 | GU371136 | GU371197 |
| <i>Arum dioscoridis ssp. cyprium</i> | Chase, RBG-Kew, 11021K | Turkey | GU370985 | GU371077 | GU371141 | GU371201 |
| <i>Arum dioscoridis1</i> | Chase, RBG-Kew, 11015K | Turkey | GU370986 | GU371078 | GU371142 | GU371202 |
| <i>Arum dioscoridis2</i> | Küpfer, pr. coll., 09.05.30 4 | Turkey | GU370984 | GU371064 | GU371127 | - |
| <i>Arum elongatum1</i> | Boyce, RBG-Kew, 1990-2019 | Turkey | GU370989 | GU371079 | GU371143 | GU371203 |
| <i>Arum elongatum2</i> | Bedalov, pr. coll., 16.5.03 | Ukraine | GU370990 | - | GU371131 | GU371192 |
| <i>Arum elongatum3</i> | Küpfer, pr. coll., 09.05.09 1 | Turkey | GU370991 | GU371069 | GU371133 | GU371194 |
| <i>Arum euxinum</i> | Chase, RBG-Kew, 11019K | Turkey | GU370992 | GU371029 | GU371092 | GU371156 |
| <i>Arum hygrophilum</i> | Chase, RBG-Kew, 11027K | Israel | GU370993 | GU371030 | GU371093 | GU371157 |
| <i>Arum idaeum</i> | Boyce, RBG-Kew, 1993-1895 | Crete | GU370994 | GU371031 | GU371094 | GU371158 |
| <i>Arum italicum ssp. albispalum</i> | Chase, RBG-Kew, 11020K | Georgia | GU370995 | GU371035 | GU371098 | GU371162 |
| <i>Arum italicum ssp. canariense1</i> | Chase, RBG-Kew, 11032K | Madeira | GU370996 | GU371032 | GU371095 | GU371159 |
| <i>Arum italicum ssp. canariense2</i> | Chase, RBG-Kew, 11031K | Madeira | GU370997 | GU371034 | GU371097 | GU371161 |
| <i>Arum italicum ssp. italicum</i> | Boyce, RBG-Kew, 1978-4984 | Greece | GU370998 | GU371033 | GU371096 | GU371160 |
| <i>Arum jacquemontii</i> | Boyce, RBG-Kew, 1969-5385 | Afghanistan | GU370999 | GU371036 | GU371099 | GU371163 |
| <i>Arum korolkowii</i> | Boyce, RBG-Kew, 1994-3354 | Tadzhikistan | GU371000 | GU371037 | GU371100 | GU371164 |
| <i>Arum lucanum1</i> | Boyce, RBG-Kew, 1987-1133 | Italy | GU371001 | GU371039 | GU371102 | GU371166 |
| <i>Arum lucanum2</i> | Boyce, RBG-Kew, 1991-887 | Italy | GU371002 | GU371051 | GU371114 | GU371176 |
| <i>Arum maculatum1</i> | Chase, RBG-Kew, 11161K | England | GU371003 | GU371040 | GU371103 | GU371167 |
| <i>Arum maculatum2</i> | NA, RBG-Kew, MJC 0002 | England | GU371004 | GU371041 | GU371104 | GU371168 |
| <i>Arum maculatum3</i> | Boyce, RBG-Kew, 1990-475 | Wales | GU371005 | GU371042 | GU371105 | GU371169 |
| <i>Arum maculatum4</i> | Boyce, RBG-Kew, 1990-2018 | Turkey | GU371006 | GU371043 | GU371106 | GU371170 |
| <i>Arum maculatum5</i> | Bedalov & Küpfer, pr. coll., 1914b | Crimea | GU371007 | GU371066 | GU371129 | GU371190 |
| <i>Arum maculatum6</i> | Espindola & Zryd, pr. coll., macKIL3 | England | GU371008 | GU371087 | GU371151 | GU371211 |
| <i>Arum maculatum7</i> | Espindola & Zryd, pr. coll., macLAC2 | Italy | GU371009 | GU371088 | GU371152 | GU371212 |
| <i>Arum maculatum9</i> | Espindola & Zryd, pr. coll., mac SFG7 | France | GU371011 | GU371090 | GU371154 | GU371214 |
| <i>Arum megobrebi1</i> | Bedalov, pr. coll., 1432 | Georgia | GU371012 | GU371058 | GU371121 | GU371183 |
| <i>Arum megobrebi2</i> | Bedalov, pr. coll., 1436 | Georgia | GU371013 | GU371059 | GU371122 | GU371184 |
| <i>Arum megobrebi3</i> | Bedalov, pr. coll., 1442 | Georgia | GU371014 | GU371071 | GU371135 | GU371196 |
| <i>Arum megobrebi4</i> | Neumann, pr. coll., 24219 | Georgia | GU371015 | GU371075 | GU371139 | GU371199 |
| <i>Arum nigrum</i> | Boyce, RBG-Kew, 1992-2083 | Montenegro | GU371016 | GU371044 | GU371107 | GU371171 |
| <i>Arum orientale ssp. longispalum</i> | Bedalov & Küpfer, pr. coll., 13.05.02 98/15 | NA | GU371018 | GU371070 | GU371134 | GU371195 |
| <i>Arum orientale1</i> | Boyce, RBG-Kew, NA | NA | GU371019 | GU371045 | GU371108 | GU371172 |
| <i>Arum orientale2</i> | Bedalov & Küpfer, pr. coll., 284 99/61 | Macedonia | GU371017 | GU371067 | GU371130 | GU371191 |
| <i>Arum palaestinum</i> | Chase, RBG-Kew, 11016K | NA | GU371020 | GU371046 | GU371109 | GU371173 |
| <i>Arum pictum</i> | Chase, RBG-Kew, 11024K | Balearic | GU371021 | GU371047 | GU371110 | - |
| <i>Arum purpureospathum</i> | Chase, RBG-Kew, 11023K | Crete | GU371022 | GU371048 | GU371111 | - |
| <i>Arum rupicola ssp. rupicola</i> | Chase, RBG-Kew, 11036K | Jordan | GU371023 | GU371052 | GU371115 | GU371177 |
| <i>Arum rupicola ssp. virensensis</i> | Chase, RBG-Kew, 11034K | Turkey | GU371024 | GU371050 | GU371113 | GU371175 |
| <i>Arum sintenisii</i> | NA, BG-Basle, NA | NA | GU371025 | GU371062 | GU371125 | GU371187 |
| <i>Arum sp. nov.</i> | Neumann, pr. coll., 26940 | Russia | GU371027 | GU371076 | GU371140 | GU371200 |
| <i>Biarum davisii</i> | NA, RBG-Kew, 2002-2839 | NA | GU370971 | GU371057 | GU371120 | GU371182 |
| <i>Biarum dispar</i> | NA, RBG-Kew, 1991-123 | Italy | GU370972 | GU371056 | GU371119 | GU371181 |
| <i>Dracunculus canariensis</i> | NA, RBG-Kew, 1982-2008 | NA | GU370987 | GU371053 | GU371116 | GU371178 |
| <i>Dracunculus vulgaris1</i> | NA, RBG-Kew, 86 - 3894 | NA | - | GU371054 | GU371117 | GU371179 |
| <i>Dracunculus vulgaris2</i> | NA, RBG-Kew, NA | England | GU370988 | GU371055 | GU371118 | GU371180 |

Chapter two

Molecular relationships in the sub-family Psychodinae (Diptera: Psychodidae)

Anahí Espíndola, Sven Buerki, Anouchka Jacquier,
Jan Ježek and Nadir Alvarez

1 **Molecular relationships in the sub-family Psychodinae (Diptera:**
2 **Psychodidae)**

3

4 Anahí Espíndola¹, Sven Buerki², Anouchka Jacquier¹, Jan Ježek³ and Nadir Alvarez⁴

5

6 ¹Laboratory of Evolutionary Entomology, Institute of Biology, University of Neuchâtel.

7 Emile-Argand 11, 2000 Neuchâtel, Switzerland.

8 ²Molecular Systematics Section, Jodrell Laboratory, Royal Botanic Gardens, Kew,

9 Richmond, Surrey TW9 3DS, United Kingdom.

10 ³Department of Entomology, National Museum of Prague (Natural History), Praha 4,

11 Czech Republic.

12 ⁴ Department of Ecology and Evolution, Biophore Building, University of Lausanne, 1015

13 Lausanne, Switzerland.

14

15 **Abstract**

16 Taxonomic and phylogenetic relationships in the order Diptera are still unclear not only at
17 lower but also at higher taxonomic levels, as for instance in the family Psychodidae (lower
18 Diptera). This family is formed by six subfamilies, of which only one (Phlebotominae) has
19 been previously investigated on phylogenetic grounds. Among the remaining groups, the
20 subfamily Psychodinae has only recently interested entomologists, and despite its worldwide
21 distribution, little is known about their taxonomic and evolutionary relationships.

22 In the present study, we investigated the phylogeny of the subfamily using two molecular
23 markers, by focusing on European/Palearctic species. A phylogenetic hypothesis as well as
24 a temporal framework are inferred through the use of a dating approach that accounts for
25 phylogenetic uncertainty.

26 Our results demonstrate that only one currently defined tribe (Psychodini) is supported by
27 molecular data as soon as it is merged with subtribe Brunettiina. Though tribes are generally
28 not well supported, this is not the case of sub-tribes *sensu* Ježek, which were partly retrieved
29 by our analyses. New fossil evidence allowed estimating the origin of the subfamily to the
30 Late Cretaceous followed by the emergence of major clades and most current species
31 during the Paleogene. Unexpectedly, a long morphological stasis is observed since all
32 species diverged before the mid-Neogene, with the exception of two pairs of paraphyletic
33 species. The impact of the Cretaceous-Paleogene (K-T) massive extinction on the
34 diversification of the subfamily is also discussed.

35 Thanks to the application of this molecular approach to the investigation of the
36 relationships in the subfamily Psychodinae, we finally propose some future lines of research
37 to establish a new formal classification at the tribal and subtribal levels.

38

39 Introduction

40 The insect order Diptera (true flies) includes over 150'000 species, around 180 families
41 and is considered one of the largest and most diverse groups of organisms (Bertone and
42 Wiegmann 2009). In addition of being highly diversified, this order has also encountered an
43 incredible success by colonizing all ecosystems (Yeates and Wiegmann 1999). Such ability
44 was probably facilitated by the fact that species within this order are adapted to a wide
45 diversity of ecological niches and trophic levels (*i.e.*, predators, decomposers, parasitoids
46 and pollinators) (Bertone and Wiegmann 2009).

47 Despite their extreme abundance and species richness (several authors advocate that
48 the species diversity is twice the number presented above), the systematics of Diptera
49 remains problematic and therefore is only partially established. During the last century,
50 taxonomists (e.g., Hennig 1976; Hennig 1981) have focused their research on publishing
51 treatments for families harboring high species richness and/or showing some economic
52 interest. Indeed, a non negligible species number of Diptera are vectors of human and
53 animal pathogens (*e.g.*, Culicidae, Psychodidae) and many are pests of crops (*e.g.*
54 Anthomyiidae, Tephritidae) (Bertone and Wiegmann 2009). Traditionally, classifications
55 identified characters related to the antennae of adults and the head capsule of larvae and
56 used them to subdivide the order into two suborders: Nematocera (“thread-horn” flies) and
57 Brachycera (“short-horn” flies) (Yeates and Wiegmann 1999). More recently, the
58 popularization of molecular methods has triggered the inference of phylogenetic frameworks
59 for this group, which allowed testing previous classifications and using phylogenies as
60 guidelines to establish new systematics when necessary (e.g., Friedrich and Tautz 1997;
61 Bertone, Courtney et al. 2008). However, the fact that *ca.* 180 families were described within
62 Diptera and that most of the families are poorly known (in terms of number of species,
63 distributions, etc) have largely prevented the inference of accurate phylogenetic hypotheses.
64 Today, while the monophyly of Brachycera is supported by molecular and morphological
65 evidence, this suborder is considered to have originated from Nematocera, currently
66 regarded as a set of paraphyletic infraorders (e.g., Bertone, Courtney et al. 2008). In addition

67 of being paraphyletic, evolutionary relationships with the lower Diptera (that partially
68 corresponds to previous Nematocera) are not resolved and disagree with previous
69 classifications. Finally, as reviewed by Bertone and Wiegmann (2009), fossil and molecular
70 data support an origin of the order Diptera sometime during the Late Paleozoic [270-251
71 million years ago (Mya)] with divergence among families taking place in the Triassic and
72 Jurassic (251-146 Mya).

73 To provide a phylogenetic insight into the relationships of some of the families, we focus
74 our analyses on the evolutionary associations within the worldwide spread family
75 Psychodidae (lower Diptera), comprising currently *ca.* 3000 species (Ježek and Barták
76 2000). This family is divided into six subfamilies (Bruchomyiinae, Trichomyiinae,
77 Horaiellinae, Phlebotominae, Psychodinae and Sycoracinae), with only some of them
78 present in Europe. Current molecular clocks estimate the origin of the family to date back to
79 the Cretaceous (*ca.* 140 Mya, Bertone and Wiegmann 2009), whereas fossil evidence [not
80 taken into account for the dating of Diptera mentioned in Blagoderov *et al.* (2007)] suggests
81 an earlier origin during the Late Triassic-Early Jurassic (*ca.* 201 Ma). Within the family, a
82 phylogenetic hypothesis was only inferred for the subfamily Phlebotominae, a well-known
83 taxon because of the ability of several species to carry human diseases (Depaquit, Perrotey
84 *et al.* 1998; Beati, Caceres *et al.* 2004). In order to contribute to the knowledge of the
85 evolution of the family, we focus in this survey on another subfamily: the Psychodinae, and
86 most precisely on the study of samples inhabiting the European/Palearctic portion of its
87 distribution.

88 Subfamily Psychodinae, presents a broad morphological and ecological diversity that
89 allowed species from this group to disperse worldwide and to colonize and survive in remote
90 places such as oceanic islands and subantarctic regions (Withers 1988). Since the last
91 century, Vaillant (1971) and Ježek (1984) have provided a strong species-level taxonomic
92 framework. For instance, Vaillant (1971) attempted to describe and normalize characters for
93 classification of larvae and proposed identification keys to relate them to their respective
94 imagi, while Ježek (1984) reorganized nomenclatorial problems and described species for

95 regions little investigated. Despite these works, all supra-specific entities have been subject
96 to controversy and even challenged the first treatment done by Enderlein (1936) (several
97 lumpings and splittings of taxonomic entities have been proposed; see Table 1). First,
98 Vaillant (1971) recommended to subdivide the Palearctic Psychodinae into four tribes:
99 Telmatoscopini, Psychodini, Pericomini and Brunettiini. Unfortunately, most of Vaillant's work
100 lacked of nomenclatural (several types were wrongly cited) and cladistic rigor (relationships
101 between species and genera in trees were not clarified), and Ježek (1983; 1990) thus
102 proposed an alternative treatment based on a cladistic approach.

103 Although the circumscription of suprageneric levels of classification within the subfamily
104 remains a challenging task, the strong taxonomic background available in Palearctic species
105 (as well as the cladistic hypotheses provided by Ježek, 1983; 1990) allow confronting the
106 past view of the Psychodinae systematics with a molecular approach. Based on a sampling
107 covering all European tribes of Psychodinae, we use molecular sequencing of mitochondrial
108 regions to infer phylogenetic relationships within the subfamily. The aims of this study are i)
109 to evaluate the validity of Palaeartic tribes *sensu* Ježek or Vaillant; ii) when possible, to test
110 the monophyly of genera; iii) to estimate the temporal origin of the subfamily and of the main
111 tribes; iv) to estimate the age of current species in the context of a group that spanned the
112 last 250 Mya; v) to propose guidelines for future classification within Psychodinae.

113

114 **Material and Methods**

115 *Sampling*

116 The sampling comprising 52 species [distributed into 26 genera, representing all tribes of
117 the European Psychodinae (Ježek 2009) and including 87 samples] (see Supplementary
118 Material 1 for information on success of amplification of samples) was obtained from the
119 personal collection of Dr. J. Ježek. Samples were collected during several field trips, from
120 years 1995 to 2009 in Eastern Europe, and were preserved in 70% ethanol. Seven outgroup
121 species were added to the dataset based on recent phylogenetic studies on Diptera (see
122 e.g., Bertone, Courtney et al. 2008). The outgroup species are distributed as follows: six

123 species belonging to the infraorder Culicomorpha (*Anopheles funestus*, *A. gambiae*, *A.*
124 *quadrimaculatus*, *A. darling*, *Aedes albopictus* and *A. aegypti*) and one species belonging to
125 the subfamily Phlebotominae (*Phlebotomus riouxi*).

126

127 *DNA extraction, amplification and sequencing*

128 DNA from insects was extracted using the QIAGEN DNEasy Animal tissue extraction kit
129 (QIAGEN, Hombrechtikon, Switzerland). Two mitochondrial regions (hereafter mtDNA),
130 NADH1 and Cytochrome B (Cytb), were amplified with primers described in Simon *et al.*
131 (1994) (N1-J-12248 -AAG CTA ATC TAA CTT CAT AAG- and LR-N-12866 -ACA TGA TCT
132 GAG TTC AAA CCG G- for NADH1, and CB-J-11338 -CAC ATT CAA CCA GAA TGA TAT
133 TT- and N1-N-12051 -GAT TTT GCT GAA GGT GAA TCA GA- for Cytb). Amplifications
134 were performed in a 20µl master mix consisting of 0.5X buffer, 100mM dNTP, 0.5 µM of
135 each primer, 0.12mM MgCl₂, 1 unity of GoTaq Taq-polymerase (Promega, Switzerland) and
136 4 µl DNA. Reactions were run in a Biometra Thermocycler (Biometra, Goettingen,
137 Germany). An initial denaturation of 1:30 min at 95°C, followed by 35 cycles of 1 min of
138 denaturation at 95°C, 1 min of annealing at temperatures comprised between 42°C and
139 52°C, depending of samples and primers, and 30 s of elongation at 72°C. Cycles were
140 followed by 5 min of final elongation at 72°C. The purification of PCR products and
141 fluorescence sequencing were done by Macrogen Inc. (South Korea) and Fasteris SA
142 (Switzerland) with the same primers as used for the PCR amplification.

143

144 *Sequence alignment and phylogenetic reconstruction*

145 The program ChromasPro 1.41 (Technelysium Pty Ltd.) was used to assemble
146 complementary strands and verify software base-calling. The two mtDNA regions were
147 initially aligned applying the Clustal-Wallis algorithm (implemented in BioEdit 7.0.4.1; Hall
148 1999) and further visually adjusted. For each mtDNA region gaps were coded in FastGap
149 1.2 (Borchsenius 2009), applying the simple gap-coding algorithm of Simmons and
150 Ochoterena (2000). Finally, the number of constant, variable, parsimony informative and

151 total base pairs were calculated for each partition using MEGA 4.0.1 (Tamura, Dudley et al.
152 2007).

153 Before computing total evidence analyses, an incongruence length difference (ILD) test
154 (Farris, Källersjö et al. 1994) was performed using PAUP* 4b10 (Swofford 2003) with 100
155 replicates. If the probability value provided by the analysis is higher than 0.05, it is
156 statistically correct to assume that the information provided by both partitions is congruent
157 (Swofford 2003). Because the ILD test strongly rejected the incongruence between the
158 datasets (p -value=1), both regions were combined and total evidence analyses were
159 performed.

160 Phylogenetic relationships were inferred based on both probabilistic [*i.e.*, Bayesian
161 inference and Maximum Likelihood (ML)] and cladistic [*i.e.*, Maximum Parsimony (MP)]
162 approaches. A partitioned Bayesian analysis was performed using a Markov-Chain Monte-
163 Carlo (MCMC) approach, as implemented in MrBayes v3.1.2 (Ronquist and Huelsenbeck
164 2003) and following Nylander *et al.* (2004). Best-fit models for each region were selected
165 using MrModeltest v.3.0 (Nylander 2004) based on the Akaike information criterion (Akaike
166 1973). For both mtDNA partitions, the best-fit model was the general time reversible (GTR)
167 model with an alpha parameter for the shape of the gamma distribution to account for rate
168 heterogeneity among sites (Yang 1993). Since datasets were highly congruent (see above)
169 gaps from both mtDNA regions were merged into a single partition and the “restriction model”
170 was applied. Two Metropolis-coupled Markov chains with incremental heating temperature of
171 0.5 were run for 15 million generations and sampled every 1000th generation. The analysis was
172 repeated twice, starting from random trees. Convergence was accepted when standard
173 deviations attained values below 0.01 and when the Potential Scale Reduction Factor index
174 (Gelman and Rubin 1992) approached 1.0. We considered the MCMC sampling sufficient
175 when the Effective Sampling Size was higher than 200 –checked on Tracerv1.4 (Rambaut
176 and Drummond 2004). After a burn-in period corresponding to 5'000 trees a half-compatible
177 consensus tree (and its associated Bayesian posterior probabilities; BPP) was reconstructed

178 (based on the remaining 10'000 trees) using MrBayes v3.1.2 (Ronquist and Huelsenbeck
179 2003).

180 The ML analysis was done using RAxML 7.2.6 (Stamatakis 2006) with 10'000 rapid
181 bootstrap analyses followed by the search of the best-scoring ML tree in one single run. The
182 two mtDNA regions were considered as one single partition (by applying the GTR model)
183 and the gap partition was not considered. The analysis was done using the facilities offered
184 by the CIPRES portal (San Diego, CA, USA).

185 The combined dataset was analyzed using parsimony ratchet (Nixon, 1999) with PAUPrat
186 (Sikes and Lewis 2001). Based on recommendations of Nixon (1999), ten independent
187 searches (with 200 iterations and 15% of the parsimony informative characters perturbed)
188 were conducted. The most parsimonious trees were afterwards combined to create a
189 majority-rule consensus tree. Branch supports were estimated using the Decay Index
190 (Bremer 1988) as implemented in TreeRot v3 (Sorenson and Franzosa 2007). The Decay
191 Index indicates the number of steps necessary to collapse a node, informing on the stability
192 and consistency of each clade (Felsenstein 2004). The congruence of topological groupings
193 between the Bayesian, ML and MP algorithms was evaluated by visually comparing
194 topologies and node supports.

195

196 *Estimation of lineage divergence times*

197 Relative branching times were estimated on the halfcompat consensus tree of MrBayes
198 using Penalized Likelihood (PL; Sanderson 2002), as implemented in r8s v.1.71 (Sanderson
199 2004), and the Truncated Newton method algorithm. The smoothing value (10) was
200 established using the cross-validation routines implemented in r8s. One of the most external
201 outgroups, *Aedes aegypti*, was pruned for the estimation of the divergence time as required
202 by the program (see Sanderson 2004).

203 To account for phylogenetic uncertainty in the estimation of lineage divergence times, PL
204 analyses were performed on a random sample of trees (N = 4000) from the Bayesian MCMC
205 stationary distribution as done in Buerki et al. (in press). This was done as follows: i) PL

206 analyses were run independently on the set of trees using the same smoothing parameter
207 value as the consensus tree; ii) 95% confidence intervals of age estimates for each node
208 compatible with the halfcompat tree were calculated based on the sample of PL-dated trees;
209 iii) the intervals of confidence were finally represented on the consensus tree. These steps
210 were done using a set of R scripts (R Development Core team, 2009) available on request to
211 SB.

212 To estimate absolute ages for lineage divergences, three fossil calibration points (see
213 below) were used to set minimum age constraints for several nodes in the phylogeny (the
214 root was estimated using a maximum node age; see below). The calibration procedure was
215 done as in Buerki et al. (in press). For each calibration point, the oldest fossil record was
216 selected and the upper (younger) bound of the geological interval (Walker and Geissman
217 2009) in which the fossil was found was used to represent the minimum age constraint.

218 Calibration points were defined as follows: (a) the root node – *i.e.*, the most recent
219 common ancestor (MRCA) of Diptera – was constrained to a maximum age of 260 million
220 years (Mya). According to Bertone *et al.* (2008) fossil evidences supporting this calibration
221 point were estimated from the Capitanian to the Wuchiapingian (Mid- Late Permian). (b) The
222 stem group of the Psychodidae was set to a minimum age of 201.6 Ma. This calibration point
223 is based on fossil evidence (Blagoderov, Grimaldi et al. 2007) occurring from the Rhaetian to
224 the Hetangian (Late Triassic-Early Jurassic). (c) Finally, the stem group of Psychodinae was
225 set to a minimum age of 176 Ma. This point is based on fossils (Ansorge 1994) found for the
226 Toarcian to the Aalenian periods (Early-Mid Jurassic).

227 Finally, in order to evaluate the consistency and age of species, their monophyly was
228 retrieved from the Bayesian topology and the time between the species' MRCA and the
229 anterior split of the lineage (*i.e.*, the age of the node anterior to the MRCA, corresponding to
230 the split with the sister species) was calculated and recorded for each species using scripts
231 available to SB. Classes of five million years were considered to reconstruct histograms for
232 the visualization of the distribution of species divergence through time.

233

Results

Among 94 sequenced samples (87 in the ingroup and seven outgroups), 83 properly amplified region Cytb and 57 region NDHA1 (Supplementary Material 1). The combined aligned dataset was 1248bp length: 652bp for Cytb and 596bp for NADH1. A gap partition of 87 characters (coded as presence/absence) was added to the matrix. The number of variable, constant and potentially parsimony-informative sites are summarized in Table 2. The number of variable sites were highly similar between the two mtDNA regions and almost 50% of sites were parsimony-informative (Table 2).

The dating analysis (performed on 4000 trees randomly selected from the Bayesian posterior probability; Figure 1) estimated the origin of the MRCA of Psychodinae as having happened during the Late Cretaceous (*ca.* 85.6 Mya), and the stem of the subfamily going back to the Mid Jurassic (*ca.* 176 Mya) (Figure 1). Within Psychodinae, lineages diverged between the Late Cretaceous (*ca.* 75.01 Mya) and the Early Paleogene (59.97 Ma), with most of the differentiation taking place during the Paleogene. The ages of species' lineages were unexpectedly ancient, with most stems originating in the Paleogene, and no lineages more recent than 10 Ma (Figure 3).

The three phylogenetic approaches provided partially congruent topologies (Figure 2 and Supplementary Material 2). We defined main clades at higher taxonomic levels, by using a temporal cut-off at 75 Ma. Four clades were thus retrieved, namely I to IV (Figure 2). While clades I, II and III were also retrieved in the ML and MP approaches, this was not the case of clade IV, which was paraphyletic with regard to the other clades in the MP approach. Because clades II and IV comprised highly diverging lineages, we categorized taxa one step further and considered five subclades in each of these two clades (respectively IIa to IIe, and IVa to IVe) (Figure 2). These subclades were defined on a stem age ranging between 46 and 60 Ma. While half of them were monophyletic whatever approach used, the other half showed a looser level of congruence among phylogenetic approaches; node supports varied as well, ranging from low to high values (see Table 3 and Figure 2). Monophyly irrespective of the phylogenetic criterion was considered as strong evidence to validate a given clade. At

262 the generic level, seven of the twelve genera for which more than one species was sampled
263 were monophyletic in at least one criterion of phylogenetic reconstruction (Table 4). The
264 remaining five genera were para- or polyphyletic whatever criterion applied.

265

266 **Discussion**

267 *Intricate relationships between taxonomy and phylogeny*

268 Results presented in this study i) support the monophyly of subfamily Psychodinae, ii)
269 indicate the polyphyly of all tribes (with the only exception of Psychodini that is paraphyletic
270 with subtribe Brunettiina; see below) and iii) challenge the monophyly of several genera
271 (Figures 1 and 2).

272 To avoid misleading relationships caused by putative high saturations of the mtDNA
273 regions (see below), we discuss in this study only genera para/polyphyletic under the three
274 phylogenetic algorithms. From this perspective, five genera are either polyphyletic (*Logima*)
275 or paraphyletic (*Pericoma*, *Pneumia*, *Psychoda* and *Ulomyia*) (Table 4). The polyphyletic
276 genus *Logima* is retrieved in two clades (II d and II e), whereas the paraphyletic genera are
277 restricted to one clade (IV e). In the latter case, further investigations are required before
278 concluding on the status of the genera.

279 Although not in perfect agreement with the phylogenetic relationships, two of the four
280 subtribes defined by Ježek match our phylogenetic clades: Brunettiina (clade II b) and
281 Mormiina (clade IV c) (Table 3). The two remaining subtribes, Paramormiina (clade IV) and
282 Trichopsychodina (clades I and III) are paraphyletic (Figure 2 and Table 3). These new
283 findings suggest that part of the taxonomy/systematics of Ježek (based on cladistic
284 analyses) tends to reflect the evolutionary history of Psychodinae and should be considered
285 when proposing a new formal classification. In addition, even if Trichopsychodina is
286 paraphyletic, the two lineages including representatives of this tribe comprise only genera
287 *Threticus* (clade I) and *Trichopsychoda* (clade III) respectively, what might therefore also
288 constitute valuable information to circumscribe future subtribes.

289 Despite genera of tribe Psychodini were not clearly assigned to any subclade, they were
290 all restricted to clade II. Considering this, our result constitutes a step towards the
291 establishment of a new classification that incorporates both morphological and molecular
292 evidence. Tribe Psychodini is however paraphyletic due to the inclusion of Brunettiina
293 (*Atrichobrunettia*), a subtribe that previously challenged taxonomists (Figure 2, Table 3).
294 Even though this subtribe has been recognized as a separate entity, but at different
295 taxonomic ranks according to the researcher (Vaillant 1971; Ježek 1984), our analysis
296 supports its relationships with members of tribe Psychodini (Figures 1 and 2; Table 3).
297 Further investigations will be required to identify morphological synapomorphies supporting
298 the definition of these subclades. Based on our results, it appears that subtribe Brunettiina
299 presents an evolutionary history extremely related to that of tribe Psychodini. We thus
300 recommend taxonomists to adopt a broad concept of tribe Psychodini by also including
301 subtribe Brunettiina, since this would reflect historical and evolutionary relationships between
302 the two groups.

303 Finally, the remaining clade IV (Figures 1 and 2) is the most problematic when compared
304 with current classifications. With the only exception of subtribe Mormiina, none of the
305 published taxonomic entities corresponds to these phylogenetic subclades (Table 3). Tribe
306 Pericomini is retrieved in two non-sister clades (IVb and IVe) suggesting the need of
307 providing a new circumscription for this group. Moreover, a similar situation is observed with
308 subtribe Paramormiina (clades IVa, IVd). Although clades are usually moderately to strongly
309 supported, phylogenetic relationships within clade IV are not clear and would deserve
310 additional investigations before allowing further taxonomic recommendations.

311 To solve phylogenetic relationships (*e.g.*, basal polytomy of clades I, II, III, IV) and
312 recognize morphological synapomorphies supporting clades (at the basis of a new formal
313 classification) we suggest that researchers should i) increase their sampling to balance the
314 number of representatives at the different taxonomic levels, ii) sequence less polymorphic
315 markers (*e.g.*, 28S) and iii) compile an extensive morphological matrix. Although paradoxical
316 at first thought, the sequencing of less polymorphic regions will allow solving basal

317 polytomies that are most likely caused by high levels of saturation of the molecular regions
318 used in this study (data not shown). This putative high level of saturation might also be the
319 reason of the incongruence found between algorithms, especially between probabilistic
320 algorithms and MP inferences (e.g. Clade IV). Probabilistic algorithms -through the use of
321 evolutionary models- appear thus to be better suited to handle such situations and should be
322 preferred over the MP approach in the case of this group.

323

324 *The timing of diversification in Psychodinae*

325 Although our dating inference might be biased by taxon sampling, the new fossil evidence
326 (Blagoderov, Grimaldi et al. 2007) used here (compared to Bertone and Wiegmann 2009)
327 allows estimating the origin of subfamily Psychodinae to sometime during the Late
328 Cretaceous (ca. 85 Mya; Figure 1), with an emergence of all major clades during the
329 Paleogene (65.5-23 Mya; Figures 1 and 3). Our results also support a diversification of all
330 monophyletic genera during the Eocene (ca. 35 Mya) and, more strikingly, the emergence of
331 sampled species during the Miocene (before 10 Mya; Fig. 3). Once all lineages diversified, a
332 long period of “morphological” stasis is observed.

333 From a historical point of view, our results show that clades appear to have diverged
334 during the Paleogene, a period directly following the now well-known K-T mass-extinction
335 (around 65 Mya). This mass-extinction event was extraordinary and has been well identified
336 in the fossil register, where the extinction of high proportions (i.e. up to 95% in some taxa) of
337 species was observed in short geological times (Jablonski 2001). The period following this
338 massive extinction has been punctuated in several groups by a rapid diversification,
339 explained generally by the fact that massive extinctions left “free” environments with
340 changed ecological networks, in which species found ample niches to evolve in (Erwin
341 2001). It is thus interesting to note that the origin of tribe Psychodini *sensu lato* (including
342 Brunettiina) coincides with this post-K-T period (with a MRCA at 64 Mya). Moreover, almost
343 all main groups in the subfamily appear to have diversified during the period directly
344 following the *pulse* K-T extinction (sensu Erwin 1996), which would be in agreement with the

345 idea of post-massive extinction radiations with differential lag-times depending on the
346 organism studied (Erwin 2001).

347 The ancient origin of Psychodinae, together with the observed “morphological” stasis are
348 additional clues supporting the intricate evolutionary history of this group. These evidences
349 (probably combined with high extinction events as those proposed above) appear thus as
350 additional explanations for the multiple obstacles that have up to now prevented the
351 development of a convincing classification for the group. In this context, our study constitutes
352 a leap forward in the understanding of the evolution of Psychodinae and provides useful
353 clues for a new formal classification. We are convinced that the circumscription of high
354 taxonomic ranks within the subfamily should also be based on dating and evolutionary
355 grounds, especially when considering the difficulties experienced when trying to define
356 taxonomic borders using morphological characters only.

357

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362

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- 466
- 467
- 468

469 Table 1 – Systematic position of all sampled taxa. Names are given following the taxonomy
 470 proposed by Ježek (1984) and FAUNA EUROPAEA (Fauna Europaea Web Service, 2004). Supra-
 471 generic classification is given following Ježek (1984) and Vaillant (1971). Phylogenetic classification
 472 considering this work is given in the last column.

| Species name (Ježek) | Species name (FAUNA EUROPAEA) | Ježek (1984) | | | Vaillant (1971) | This study |
|-------------------------------------|---|--------------|--------------|-------------------|-----------------|------------|
| | | Sub-family | Tribe | Sub-tribe | Tribe | |
| <i>Threticus incurvus</i> | <i>Threticus incurvus</i> Krek 1972 | Psychodinae | Paramormiini | Trichopsychodina* | Telmatoscopini | I |
| <i>Threticus lucifugus</i> | <i>Threticus lucifugus</i> Walker 1856 | Psychodinae | Paramormiini | Trichopsychodina* | Telmatoscopini | I |
| <i>Threticus silvaticus</i> | <i>Threticus silvaticus</i> Ježek 1986 | Psychodinae | Paramormiini | Trichopsychodina* | Telmatoscopini | I |
| <i>Psychodocha cinerea</i> | <i>Psychoda cinerea</i> Banks 1894 | Psychodinae | Psychodini* | | Psychodini | IIa |
| <i>Psychodocha gemina</i> | <i>Psychoda gemina</i> Eaton 1904 | Psychodinae | Psychodini* | | Psychodini | IIa |
| <i>Atrichobrunettia graeca</i> | <i>Atrichobrunettia graeca</i> Ježek & Goutner 1993 | Psychodinae | Mormiini | Brunettiina* | Brunettiini | IIb |
| <i>Psychodula minuta</i> | <i>Psychoda minuta</i> Banks 1895 | Psychodinae | Psychodini* | | Psychodini | IIc |
| <i>Copropsychoda brevicornis</i> | <i>Psychoda brevicornis</i> Tonnoir 1940 | Psychodinae | Psychodini* | | Psychodini | IId |
| <i>Logima albipennis</i> | <i>Psychoda albipennis</i> Zetterstedt 1852 | Psychodinae | Psychodini* | | Psychodini | IId |
| <i>Logima satchelli</i> | <i>Psychoda satchelli</i> Quate 1955 | Psychodinae | Psychodini* | | Psychodini | IId |
| <i>Ypsydocha setigera</i> | <i>Psychoda setigera</i> Tonnoir 1923 | Psychodinae | Psychodini* | | Psychodini | IId |
| <i>Logima erminea</i> | <i>Psychoda erminea</i> Eaton 1898 | Psychodinae | Psychodini* | | Psychodini | Ile |
| <i>Psycha griseascens</i> | <i>Psychoda griseascens</i> Tonnoir 1923 | Psychodinae | Psychodini* | | Psychodini | Ile |
| <i>Psychoda crassipennis</i> | <i>Psychoda crassipennis</i> Tonnoir 1940 | Psychodinae | Psychodini* | | Psychodini | Ile |
| <i>Psychoda phalaenoides</i> | <i>Psychoda phalaenoides</i> Linnaeus 1758 | Psychodinae | Psychodini* | | Psychodini | Ile |
| <i>Psychoda uniformata</i> | <i>Psychoda uniformata</i> Haseman 1907 | Psychodinae | Psychodini* | | Psychodini | Ile |
| <i>Psychomora mycophila</i> | <i>Psychoda mycophila</i> Vaillant 1990 | Psychodinae | Psychodini* | | Psychodini | Ile |
| <i>Tinearia alternata</i> | <i>Tinearia alternata</i> Say 1824 | Psychodinae | Psychodini* | | Psychodini | Ile |
| <i>Tinearia lativentris</i> | <i>Tinearia lativentris</i> Berden 1952 | Psychodinae | Psychodini* | | Psychodini | Ile |
| <i>Trichopsychoda hirtella</i> | <i>Trichopsychoda hirtella</i> Tonnoir 1919 | Psychodinae | Paramormiini | Trichopsychodina* | Telmatoscopini | III |
| <i>Clogmia albipunctata</i> | <i>Clogmia albipunctatus</i> Williston 1893 | Psychodinae | Paramormiini | Paramormiina* | Telmatoscopini | IVa |
| <i>Paramormia polyscoidea</i> | <i>Paramormia polyscoidea</i> Krek 1970 | Psychodinae | Paramormiini | Paramormiina* | Telmatoscopini | IVa |
| <i>Paramormia ustulata</i> | <i>Paramormia ustulata</i> Walker 1856 | Psychodinae | Paramormiini | Paramormiina* | Telmatoscopini | IVa |
| <i>Sciria advena</i> | <i>Telmatoscopus advenus</i> Eaton 1893 | Psychodinae | Paramormiini | Paramormiina* | Telmatoscopini | IVa |
| <i>Clytocerus longicorniculatus</i> | N/A | Psychodinae | Pericomini* | | Pericomini | IVb |
| <i>Clytocerus ocellaris</i> | <i>Clytocerus ocellaris</i> Meigen 1818 | Psychodinae | Pericomini* | | Pericomini | IVb |
| <i>Clytocerus rivosus</i> | <i>Clytocerus rivosus</i> Tonnoir 1919 | Psychodinae | Pericomini* | | Pericomini | IVb |
| <i>Oomormia andrenipes</i> | <i>Mormia andrenipes</i> Strobl 1910 | Psychodinae | Mormiini | Mormiina* | Pericomini | IVc |
| <i>Jungiella procera</i> | <i>Jungiella procera</i> Krek 1971 | Psychodinae | Paramormiini | Paramormiina* | Telmatoscopini | IVd |
| <i>Jungiella valachia</i> | <i>Jungiella valachia</i> Vaillant 1963 | Psychodinae | Paramormiini | Paramormiina* | Telmatoscopini | IVd |
| <i>Parajungiella consors</i> | <i>Jungiella consors</i> Eaton 1893 | Psychodinae | Paramormiini | Paramormiina* | Telmatoscopini | IVd |
| <i>Panimerus denticulatus</i> | <i>Panimerus denticulatus</i> Krek 1972 | Psychodinae | Paramormiini | | Telmatoscopini | IVd |
| <i>Telmatoscopus brittini</i> | <i>Telmatoscopus brittini</i> Tonnoir 1940 | Psychodinae | Paramormiini | Paramormiina* | Telmatoscopini | IVd |
| <i>Berdeniella illiesi</i> | <i>Berdeniella illiesi</i> Wagner 1973 | Psychodinae | Pericomini* | | Pericomini | IVe |
| <i>Berdeniella stavniensis</i> | <i>Berdeniella stavniensis</i> Krek 1969 | Psychodinae | Pericomini* | | Pericomini | IVe |
| <i>Berdeniella unispinosa</i> | <i>Berdeniella unispinosa</i> Tonnoir 1919 | Psychodinae | Pericomini* | | Pericomini | IVe |
| <i>Pericoma blandula</i> | <i>Pericoma blandula</i> Eaton 1893 | Psychodinae | Pericomini* | | Pericomini | IVe |
| <i>Pericoma exquisita</i> | <i>Pericoma exquisita</i> Eaton 1893 | Psychodinae | Pericomini* | | Pericomini | IVe |
| <i>Pericoma rivularis</i> | <i>Pericoma rivularis</i> Berden 1955 | Psychodinae | Pericomini* | | Pericomini | IVe |
| <i>Pneumia compta</i> | <i>Satchelliella compta</i> Eaton 1893 | Psychodinae | Pericomini* | | Pericomini | IVe |
| <i>Pneumia gracilis</i> | <i>Satchelliella gracilis</i> Eaton 1893 | Psychodinae | Pericomini* | | Pericomini | IVe |
| <i>Pneumia nubila</i> | <i>Satchelliella mutua</i> Eaton 1893 | Psychodinae | Pericomini* | | Pericomini | IVe |
| <i>Pneumia pilularia</i> | <i>Satchelliella pilularia</i> Tonnoir 1940 | Psychodinae | Pericomini* | | Pericomini | IVe |
| <i>Pneumia plumicornis</i> | <i>Satchelliella plumicornis</i> Tonnoir 1922 | Psychodinae | Pericomini* | | Pericomini | IVe |
| <i>Pneumia stammeri</i> | <i>Satchelliella stammeri</i> Jung 1954 | Psychodinae | Pericomini* | | Pericomini | IVe |
| <i>Pneumia trivialis</i> | <i>Satchelliella trivialis</i> Eaton 1893 | Psychodinae | Pericomini* | | Pericomini | IVe |
| <i>Saraiella rotunda</i> | <i>Saraiella rotunda</i> Krek 1970 | Psychodinae | Pericomini* | | Pericomini | IVe |
| <i>Ulomyia cognata</i> | <i>Ulomyia cognata</i> Eaton 1893 | Psychodinae | Pericomini* | | Pericomini | IVe |
| <i>Ulomyia fuliginosa</i> | <i>Ulomyia fuliginosa</i> Meigen 1818 | Psychodinae | Pericomini* | | Pericomini | IVe |
| <i>Ulomyia undulata</i> | <i>Ulomyia undulata</i> Tonnoir 1919 | Psychodinae | Pericomini* | | Pericomini | IVe |

473 NB. An “*” indicates the best-match subtribal or tribal classification following Ježek (1984).

474 Table 2 – Constant (C), variable (V), parsimony informative (PI) and total number of sites,
 475 in base pairs. Percentage of total base pairs are indicated in parenthesis. Gaps for each
 476 region are also shown, and total sequence length and number of gaps are indicated
 477 considering both regions together.

| | C | V | PI | Total | Gaps |
|--------------|----------------|----------------|----------------|--------------|-------------|
| NADH-1 | 306 (51.34) | 289 (48.49) | 244 (40.94) | 596 | 54 |
| Cyt-b | 264 (40.49) | 388 (59.51) | 332 (50.92) | 652 | 33 |
| Total | | | | 1248 | 87 |

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480 Table 3 – Age of the Most Recent Common Ancestor (MRCA) of each group or subgroup
 481 identified as a monophyletic clade in the Bayesian inference analysis, with best-match
 482 taxonomic classification at the subtribal or tribal (between parentheses) levels, following
 483 Ježek (1984). When subtribes were revealed to be paraphyletic, a mention for each distinct
 484 group was added (§1 and §2). Bayesian Posterior Probability (BPP), Maximum Likelihood
 485 (ML) bootstraps and Maximum Parsimony (MP) Decay indexes are shown to inform on clade
 486 support. Groups or subgroups in bold indicate clades whose monophyly is supported by all
 487 phylogenetic criteria. Names of groups and subgroups correspond to those stated in Figures
 488 1 and 2.

| Group | Subgroup | Sub-tribal taxonomy according to Ježek (1984) | Age of stem of MRCA | BPP | ML support | MP Decay Index |
|------------|--------------------|---|---------------------|-------------|------------|----------------|
| I | | Trichopsychodina §1 | 85.6 | 0.99 | 73 | 12 |
| II | <i>Ila</i> | <i>(Psychodini)</i> | 64.48 | 0.57 | 40 | N/A |
| | <i>I Ib</i> | <i>Brunettiina</i> | 47.27 | 1 | 100 | 45 |
| | <i>I Ic</i> | <i>(Psychodini)</i> | 47.27 | 0.85 | 84 | 18 |
| | <i>I Id</i> | <i>(Psychodini)</i> | 54.11 | 0.93 | 26 | N/A |
| | <i>I Ie</i> | <i>(Psychodini)</i> | 57.03 | 0.98 | 41 | N/A |
| III | | Trichopsychodina §2 | 75.01 | 1 | 100 | 0 |
| IV | <i>IVa</i> | <i>Paramormiina §1</i> | 75.01 | 0.96 | 47 | N/A |
| | <i>IVb</i> | <i>(Pericomini)</i> | 59.97 | 0.99 | 87 | 6 |
| | <i>IVc</i> | <i>Mormiina</i> | 46.84 | 1 | 100 | 31 |
| | <i>IVd</i> | <i>Paramormiina §2</i> | 46.84 | 0.55 | 48 | N/A |
| | <i>IVe</i> | <i>(Pericomini)</i> | 53.19 | 0.93 | 29 | 1 |

489 Table 4 – a) Summary of genera *sensu* Ježek (1984) for which monophyly could be
 490 retrieved with Bayesian inference (BI), Maximum Parsimony (MP) and Maximum Likelihood
 491 (ML) approaches. Number of species sampled per genus are also indicated. N/A :
 492 monophyly not tested because only one species sampled. b) Counts of monophyletic and
 493 paraphyletic genera.

494 a)

| Genus | Number of species | Monophyly | | |
|-------------------------|-------------------|--------------------|-------------------|--------------------|
| | | Bayesian inference | Maximum Parsimony | Maximum Likelihood |
| <i>Atrichobrunettia</i> | 1 | N/A | N/A | N/A |
| <i>Berdeniella</i> | 3 | yes | yes | yes |
| <i>Clogmia</i> | 1 | N/A | N/A | N/A |
| <i>Clytocerus</i> | 3 | yes | yes | yes |
| <i>Copropsychoda</i> | 1 | N/A | N/A | N/A |
| <i>Jungiella</i> | 2 | yes | yes | yes |
| <i>Logima</i> | 3 | no | no | no |
| <i>Oomormia</i> | 1 | N/A | N/A | N/A |
| <i>Panimers</i> | 1 | N/A | N/A | N/A |
| <i>Parajungiella</i> | 1 | N/A | N/A | N/A |
| <i>Paramormia</i> | 2 | no | yes | no |
| <i>Pericoma</i> | 3 | no | no | no |
| <i>Pneumia</i> | 8 | no | no | no |
| <i>Psycha</i> | 1 | N/A | N/A | N/A |
| <i>Psychoda</i> | 3 | no | no | no |
| <i>Psychodocha</i> | 2 | yes | no | yes |
| <i>Psychodula</i> | 1 | N/A | N/A | N/A |
| <i>Psychomora</i> | 1 | N/A | N/A | N/A |
| <i>Saraiella</i> | 1 | N/A | N/A | N/A |
| <i>Sciria</i> | 1 | N/A | N/A | N/A |
| <i>Telmatoscopus</i> | 1 | N/A | N/A | N/A |
| <i>Threticus</i> | 3 | yes | yes | yes |
| <i>Tinearia</i> | 2 | yes | yes | yes |
| <i>Trichopsychoda</i> | 1 | N/A | N/A | N/A |
| <i>Ulomyia</i> | 4 | no | no | no |
| <i>Ypsydocha</i> | 1 | N/A | N/A | N/A |

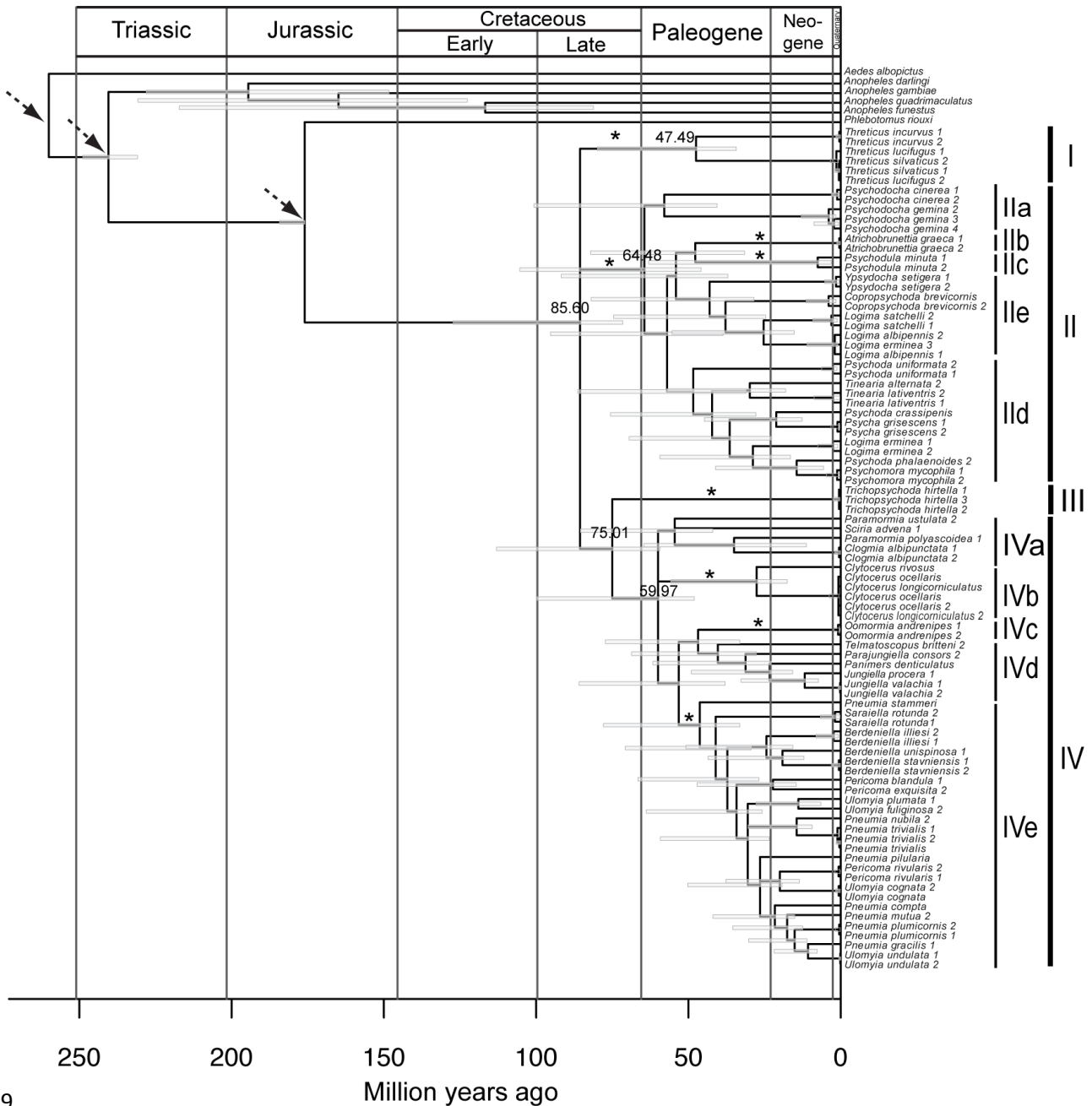
495

496 b)

| Monophyly | Count |
|-------------------------------|-------|
| yes for BI, MP or ML (or all) | 7 (5) |
| no for neither BI, MP and ML | 5 |
| N/A | 14 |
| Total | 26 |

497

498



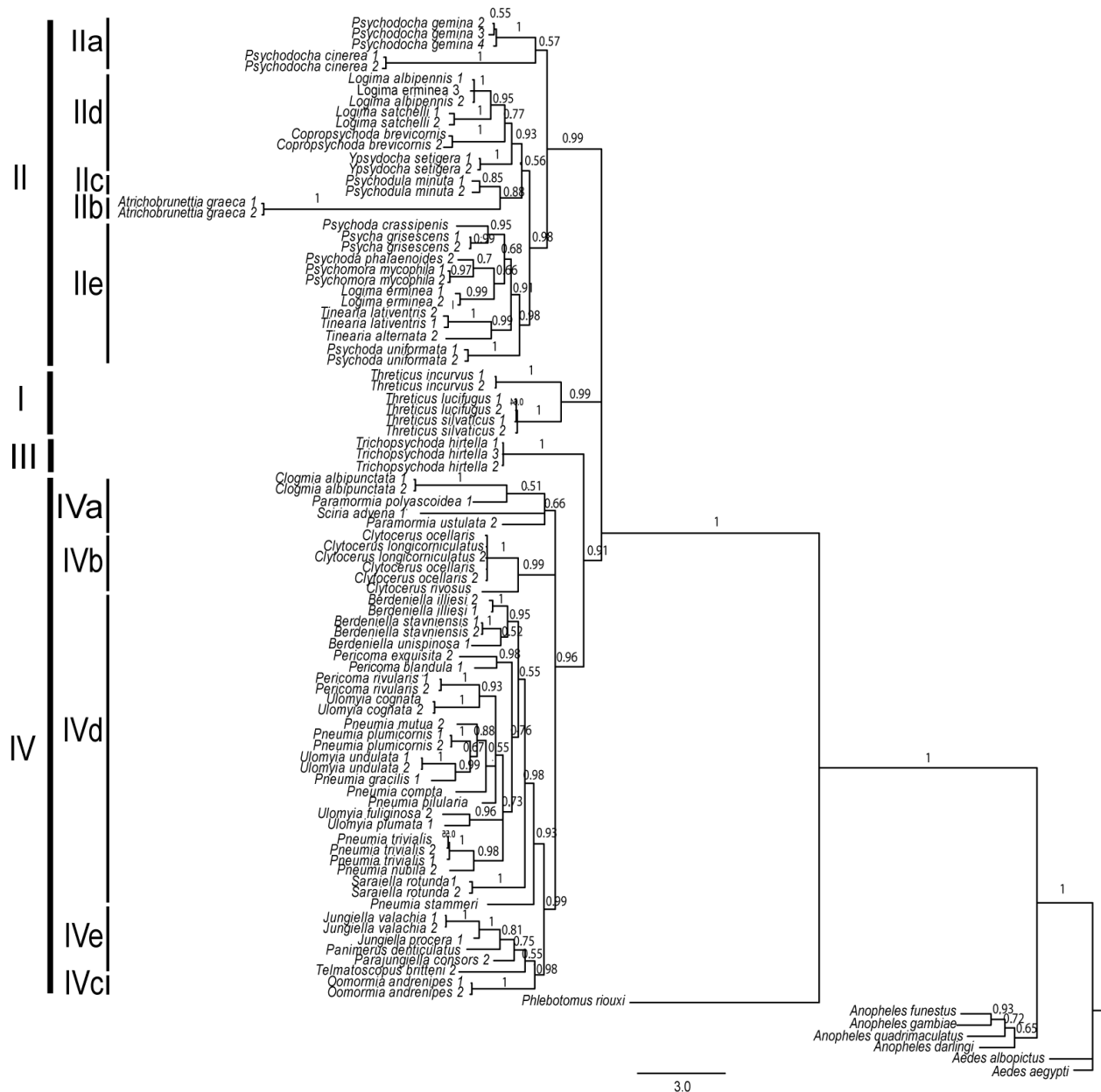
499

500 Figure 1 – Molecular dating of subfamily Psychodinae. Arrows indicate fossil calibration
 501 points. Dates on the topology indicate divergence times of the four main clades (I, II, III and
 502 IV). Vertical bars on the right illustrate the informal classification we propose to use here.

503 Stars on a stem indicate a node that is supported by a monophyletic status whatever
 504 phylogenetic criterion is applied.

505

506



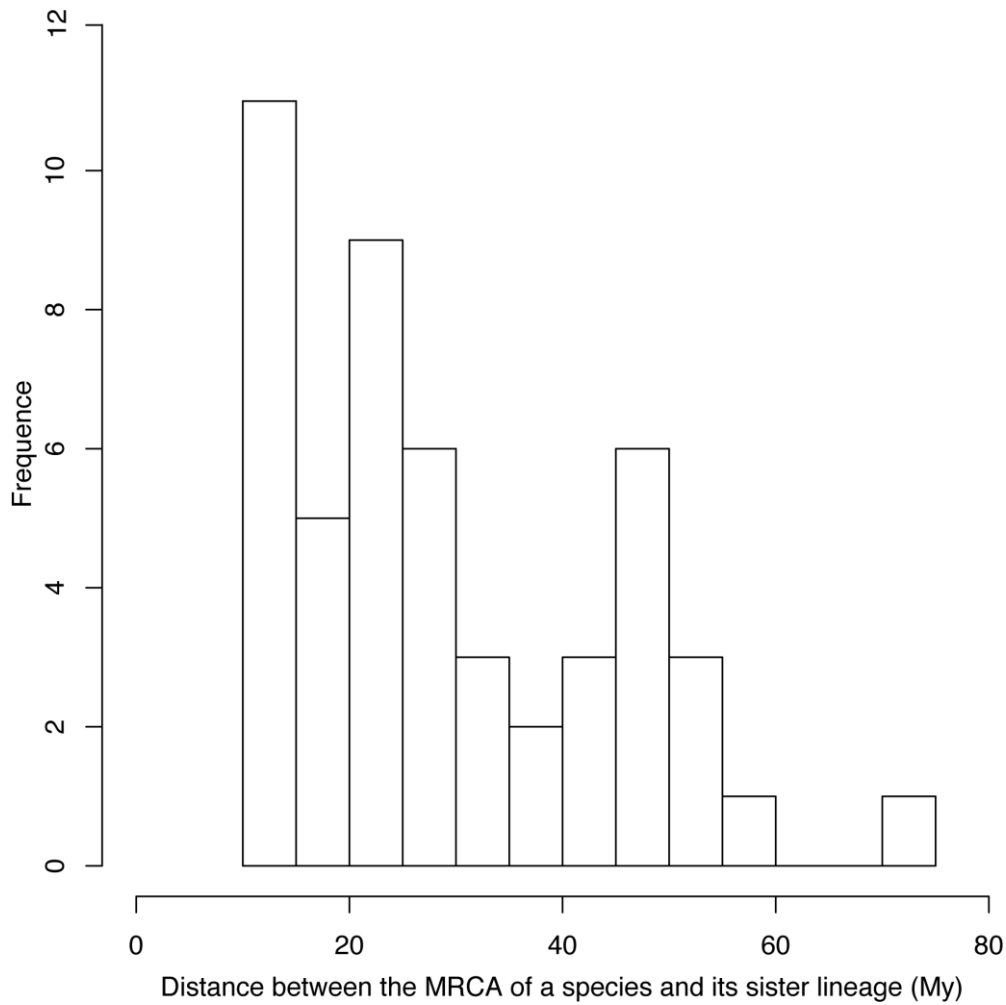
507

508 Figure 2 – Phylogeny of subfamily Psychodinae using Bayesian inference. Node supports

509 refer to Bayesian Posterior Probability (BPP) values. Vertical bars on the left illustrate the

510 classification considered in the present study.

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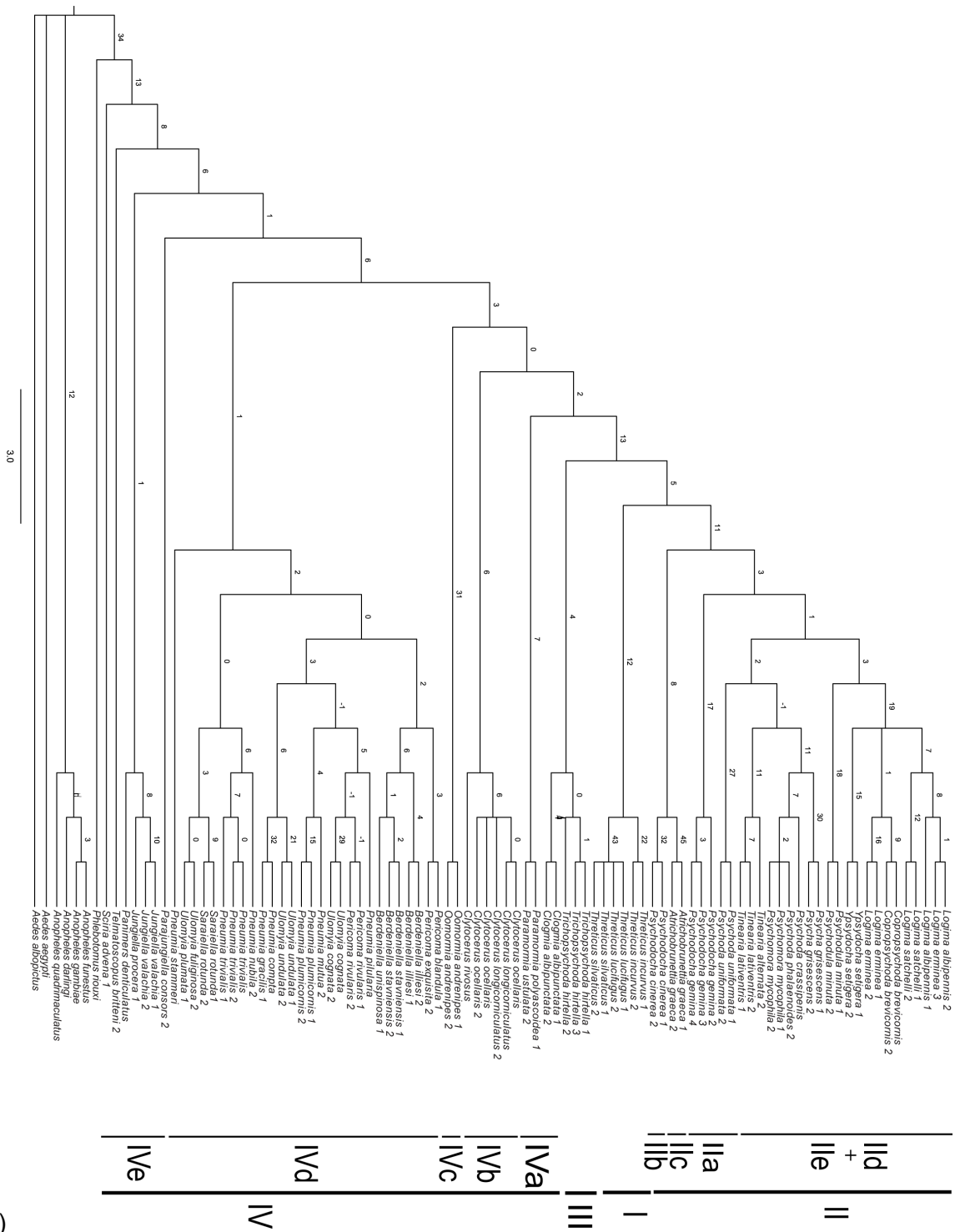
Figure 3. – Distribution in the timing of terminal-lineages diversification, as a function of the age of splitting with the sister lineage. Age is in Million years ago (Mya). Occurrences are counted according to five-million-years temporal classes (ranging from 0 to 75 Mya).

517 Supplementary Material 1 – Regions amplified for each sample (Genbank accessions to be
518 provided).

| Sample | NADH- ₄ | Cytochrome B |
|-------------------------------|--------------------|--------------|
| Berdeniella_illiesi1 | - | yes |
| Berdeniella_illiesi2 | - | yes |
| Berdeniella_stavniensis1 | - | yes |
| Berdeniella_stavniensis2 | - | yes |
| Berdeniella_unispinosa1 | - | yes |
| Clogmia_albipunctata1 | - | yes |
| Clogmia_albipunctata2 | - | yes |
| Clytocerus_longicorniculatus | yes | yes |
| Clytocerus_longicorniculatus2 | - | yes |
| Clytocerus_ocellaris | yes | yes |
| Clytocerus_ocellaris2 | yes | yes |
| Clytocerus_rivosus | - | yes |
| Copropsychocha_brevicornis | yes | yes |
| Copropsychocha_brevicornis_2 | yes | - |
| Jungiella_valachia1 | yes | yes |
| Jungiella_valachia2 | yes | yes |
| Logima_albipennis_1 | - | yes |
| Logima_albipennis_2 | yes | yes |
| Logima_erminea_1 | yes | - |
| Logima_erminea_2 | yes | - |
| Logima_erminea_3 | - | yes |
| Logima_satchelli_1 | - | yes |
| Logima_satchelli_2 | - | yes |
| Attrichobrunettia_graeca1 | - | yes |
| Attrichobrunettia_graeca2 | - | yes |
| Oomormia_andrenipes_1 | yes | yes |
| Oomormia_andrenipes_2 | yes | yes |
| Pericoma_blandula1 | yes | yes |
| Parajungiella_consors2 | yes | yes |
| Paramormia_polyascoidea1 | yes | - |
| Paramormia_ustulata2 | yes | yes |
| Panimerus_denticulatus | - | yes |
| Pericoma_exquisita2 | - | yes |
| Pericoma_rivularis1 | yes | yes |
| Pericoma_rivularis2 | - | yes |
| Pneumia_compta | - | yes |
| Pneumia_gracilis1 | yes | yes |
| Pneumia_mutua2 | - | yes |
| Pneumia_nubila2 | yes | - |
| Pneumia_pilularia | - | - |
| Pneumia_plumicornis1 | yes | yes |
| Pneumia_plumicornis2 | yes | yes |
| Pneumia_stammeri | yes | yes |
| Pneumia_trivialis3 | yes | yes |
| Pneumia_trivialis2 | yes | - |
| Pneumia_trivialis1 | yes | yes |

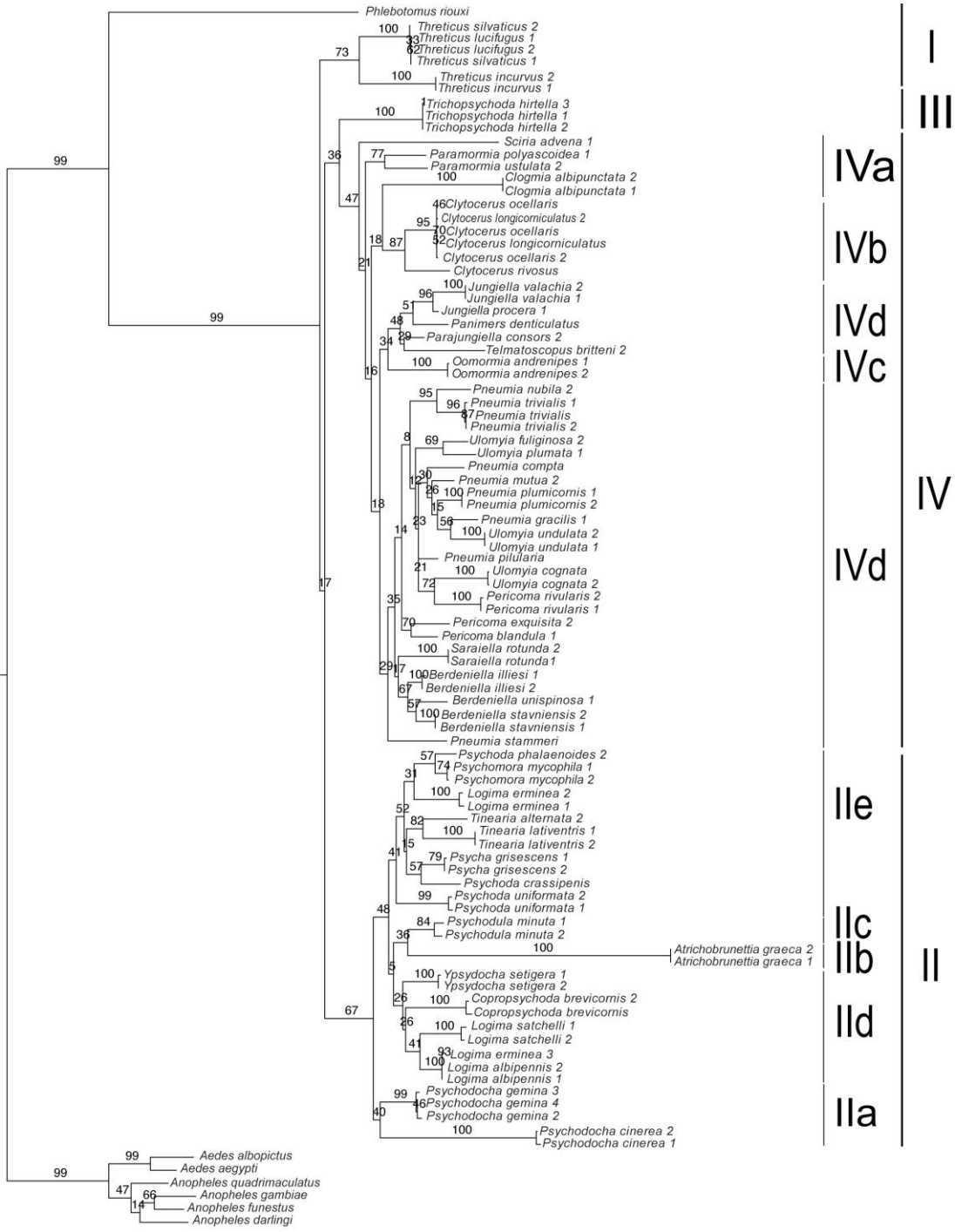
| Sample | NADH- ₄ | Cytochrome B |
|---------------------------|--------------------|--------------|
| Psychodocha_cinerea_1 | - | yes |
| Psychodocha_cinerea_2 | yes | yes |
| Psychodula_minuta_1 | - | yes |
| Psychoda_crassipenis | - | yes |
| Psychodocha_gemina_2 | - | yes |
| Psychodocha_gemina_3 | - | yes |
| Psychodocha_gemina_4 | - | yes |
| Psychocha_grisescens_1 | - | yes |
| Psychocha_grisescens_2 | yes | yes |
| Psychodula_minuta_2 | yes | yes |
| Psychoda_phalaenoides_2 | - | yes |
| Jungiella_procera1 | - | yes |
| Psychoda_uniformata_1 | yes | yes |
| Psychoda_uniformata_2 | yes | yes |
| Psychomora_mycophila_1 | yes | - |
| Psychomora_mycophila_2 | yes | yes |
| Saraiella_rotunda1 | - | yes |
| Saraiella_rotunda2 | yes | yes |
| Sciria_advena1 | yes | yes |
| Telmatoscopus_britteni2 | - | yes |
| Threticus_incurvus1 | yes | yes |
| Threticus_incurvus2 | - | yes |
| Threticus_lucifugus1 | yes | yes |
| Threticus_lucifugus2 | yes | yes |
| Threticus_silvaticus1 | yes | yes |
| Threticus_silvaticus2 | yes | yes |
| Tinearia_alternata_2 | yes | yes |
| Tinearia_lativentris1 | yes | - |
| Tinearia_lativentris2 | yes | yes |
| Trichopsychoda_hirtella2 | yes | yes |
| Trichopsychoda_hirtella3 | yes | yes |
| Ulomyia_cognata | - | yes |
| Ulomyia_cognata2 | - | yes |
| Ulomyia_fuliginosa_2 | - | yes |
| Ulomyia_plumata1 | - | yes |
| Ulomyia_undulata1 | - | yes |
| Ulomyia_undulata2 | yes | yes |
| Ypsydocha_setigera_1 | yes | yes |
| Ypsydocha_setigera_2 | yes | yes |
| Aedes albopictus | yes | yes |
| Anopheles funestus | yes | yes |
| Aedes aegypti | yes | yes |
| Anopheles darlingi | yes | yes |
| Anopheles quadrimaculatus | yes | yes |
| Anopheles gambiae | yes | yes |
| Phlebotomus riouxi | - | yes |

519 Supplementary Material 2 – Maximum Parsimony topology with Decay Indexes (a) and
 520 Maximum Likelihood phylogeny with bootstrap values (b).
 521



522 a)
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524 b)



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0.2

Chapter three

*Variation in the proportion of flower visitors of
Arum maculatum along its distributional range in
relation with community-based climatic niche
analyses*

Anahí Espíndola, Loïc Pellissier
and Nadir Alvarez

In press OIKOS

Variation in the proportion of flower visitors of *Arum maculatum* along its distributional range in relation with community-based climatic niche analyses

Anahí Espíndola, Loïc Pellissier and Nadir Alvarez

A. Espíndola (maria.espindola@unine.ch), Laboratory of Evolutionary Entomology, Inst. of Biology, Univ. of Neuchâtel, Emile-Argand 11, CH-2000 Neuchâtel, Switzerland. – L. Pellissier and N. Alvarez, Dept of Ecology and Evolution, Univ. of Lausanne, Biophore Building, CH-1015 Lausanne, Switzerland.

Because species-specific interactions between plants and insects require considerable physiological adaptations to establish and be maintained through time and space, highly specialized interactions are rare in nature. Consequently, even if some one-to-one interactions might appear locally specialized, additional partners may be involved at a wider scale.

Here, we investigate the geographical constancy in the specificity level of the specialized lure-and-trap pollination antagonism involving the widespread European *Arum maculatum* and its associated Psychodid pollinators. Until now, studies concurred in demonstrating that one single insect species, *Psychoda phalaenoides*, efficiently cross-pollinated plants; researches were, however, performed locally in western Europe. In this study we characterize for the first time the flower visitors' composition at the scale of the distribution range of *A. maculatum* by intensively collecting plants and insects throughout the European continent. We further correlate local climatic characteristics with the community composition of visiting arthropods.

Our results show that flowers are generally visited by *P. phalaenoides* females, but not over the whole distribution range of the plant. In some regions this fly species is less frequent or even absent and another species, *Psycha grisescens*, becomes the prevailing visitor. This variability is geographically structured and can be explained by climatic factors: the proportion of *P. grisescens* increases with higher annual precipitations and lower precipitations in the warmest trimester, two characteristics typical of the Mediterranean zone. Climate thus seems driving the specificity of this interaction, by potentially affecting the phenology of one or both interacting species, or even of volatile and heat production in the plant. This result therefore challenges the specificity of other presumably one-to-one interactions covering wide distribution ranges, and provides an example of the direct effect that the abiotic environment can have on the fate of plant-insect interactions.

In its early definition, the coevolutionary theory predicted that adaptive responses and speciation patterns in organisms were strongly influenced by one or a few interacting species (Ehrlich and Raven 1964). This has led to a simple view of coevolution, in which the evolutionary history of interacting organisms mostly occurs in a strict framework of coadaptation and cladogenesis, as formerly proposed in the case of figs and fig-wasps (Ramirez 1970, Jousselin et al. 2008). Establishing highly specific relationships would thus represent an evolutionary stable strategy, since it would imply a rise in the efficiency of exploitation, and would assure the constancy through time of the resource(s) exploited (Bronstein 2009). However, such cases of species-specific interactions between plants and insects are more the exception than the rule (Hoeksema and Bruna 2000) and the view of strict coevolution evolved towards a more complex and diffuse coevolutionary concept taking place at the community level (Stanton 2003). Reasons for the scarcity of one-to-one interactions include the fact that specific

biological associations require the development of particular ecological and physiological features in at least one of the interacting lineages (Brodie and Ridenhour 2002), which might entail substantial energy investments. Moreover, specialization could also cause a loss of efficiency in the exploitation of other non-specific associated species (Thompson 1994), which potentially makes these interactions sensitive to the loss of their specific partners. Thompson's theory on the geographic mosaic of coevolution (Thompson 2005) also predicts specialized interactions to be scarce in communities characterized by varying or disturbed environments: because biological interactions are expected to change along the distributional gradient, partner variations and multiple associations (McKinney 1997, Débarre and Gandon 2010) could be favoured and specialization might be then evolutionarily non-adaptive. Even if Thompson's view is sound, investigating simultaneously environmental variations and interactions is, in practice, a complex challenge. As a consequence, the environment-interaction relationship has been

only analyzed in a few multi-partners case-studies (reviewed by Laine 2009). In contrast, specific interactions provide simple and adequate natural laboratories for this type of studies, because of the restricted number of interacting species.

The genus *Arum* (Araceae) is characterized by the display of specialized antagonistic pollination interactions with arthropods, which have been documented for all but one species (Gibernau et al. 2004, Diaz and Kite 2006, Lobin et al. 2007): insects visit the plant but no service or resource is provided in exchange for pollination. One of the most studied pollination systems in the genus is that of lords-and-ladies *Arum maculatum*, a widely distributed European species (Boyce 2006, Espíndola et al. 2010). Its monoecious and protogynous inflorescences are formed by a spadix surrounded by a spathe, a characteristic feature in Araceae (Mayo et al. 1997). The lower part of the spathe is narrowed and encloses the fertile flowers, forming the floral chamber. During anthesis the chamber opening is covered by modified male flowers, the staminodes (Boyce 1993), which thus close the only aperture. In combination to these morphological features, plants of this species present a complex combination of heating cycles and odour production, which help lure small coprophilous Psychodid flies. Duped flies are attracted to the smelling flower and, after landing on the oily spathes, slip into the floral chamber. Because of the staminodes, they cannot fly away, being thus trapped for several hours. While trying to flee they move in the chamber and will – if they had been recently trapped by another *Arum* – passively pollinate the receptive female flowers. After the female phase, pollen release takes place and the trapped hairy flies are covered with it. Once pollen is liberated, staminodes wither, walls lose their oily nature and the captive insects are set free (Lack and Diaz 1991).

Among the *Arum* species known to show lure-and-trap pollination, *A. maculatum* has called the attention of ecologists for decades notably because it appears to display a highly specific pollination biology (Beck 1983, Lack and Diaz 1991, Diaz and Kite 2002). Whereas other *Arum* species are pollinated by a guild of visiting insects (Gibernau et al. 2004), investigations concurred in demonstrating that *A. maculatum* relied only on the fly species *Psychoda phalaenoides* (Diptera: Psychodidae) (Diaz and Kite 2002). Even if researchers have also identified other species (mainly *Smittia pratorum* (Diptera: Chironomidae) and *Psycha grisescens* (Diptera: Psychodidae)) trapped in floral chambers, these were present in very small proportions compared to *P. phalaenoides*, and because they were carrying significantly less pollen grains than the former it was assumed that their occurrence in the flower was accidental (Diaz and Kite 2002). Since at the several locations studied in western Europe the species composition of visiting insects was similar, and because in all sites *P. phalaenoides* was by far the most frequent visitor, it became tacitly accepted that the interaction between the plant and *P. phalaenoides* was species-specific (Gibernau et al. 2004) and constant all over the *A. maculatum* distribution range. However, the plant is distributed across a much wider area than that studied since it covers a large part of the occidental Palearctic from Spain to Denmark and from Ireland to Bulgaria (Boyce 2006) and thus potentially spans a wide diversity of climatic conditions and environments.

The theory of the geographic mosaic of coevolution would therefore predict that specialization should not be maintained throughout the plant's distribution range, and that the identity of pollinators might change in space, along environmental conditions. In order to test such predictions, we aim in the present work at: 1) evaluating the visitors composition of *A. maculatum* inflorescences with an unprecedented coverage of the whole plant's distribution range; 2) analyzing to what extent the putative spatial variation in visitors identities is correlated with environmental variations associated with climatic factors.

Material and methods

Sampling and insect identification

Fifty-six populations of *Arum maculatum* were sampled across the whole plant's distribution range (Supplementary material Appendix 1) during springs 2006, 2007 and 2008. To avoid biases related either to regional differences in the sampling effort or to local inter-annual variation, populations were sampled when possible based on a 200km grid (not shown), and all main European regions (i.e. central and northern Europe, Balkans, Italian Peninsula) were visited all years. Plant populations were sampled during blooming period and depending on populations sizes, up to ten floral chambers per population were randomly selected and collected (Supplementary material Appendix 1). Floral chambers and their visitors were independently kept in 70% ethanol. Insects were identified when possible to species with the help of entomological keys (Chinery 1976, Withers 1988, Ježek 1990, Oosterbroek 2006). Arthropods were counted, and proportions were calculated per population, per floral chamber and for the totality of the samples.

Spatial and environmental structure in the composition of visitors

Spatial variation of the floral chamber fauna was analyzed using ArcMap 9.3 (ESRI, Redlands CA, USA) by plotting and examining the compositions of all sampled visitors. To test for climatic factors influencing the composition of visitors found in the floral chambers, we performed a canonical correspondence analysis (CCA) (ter Braak 1986) using the library ade4 (Thioulouse et al. 1997) on R 2.10.0 (R Development Core Team 2010). The CCA is designed to extract synthetic environmental gradients from ecological data sets, by describing the differential habitat preferences (i.e. from a climatic point of view in our study) of taxa via an ordination diagram. Climatic values were extracted for each sampled point from seven WORLDCLIM layers at a resolution of 2.5 arc-minutes (Hijmans et al. 2005). Layers were chosen based on their ability to describe basic general characteristics of the environment (mean annual and seasonal temperatures and precipitations). Finally, the CCA was calculated on visitor abundances at different taxonomic levels (classes, orders, families, different species of Psychodids; Supplementary material Appendix 1).

Simultaneously, in order to identify the climatic factors explaining the variation in abundances of the two most

frequent and constant visiting species (i.e. *Psychoda phalaenoides* and *Psycha grisescens*), we calculated their proportion at each population, and adjusted a general linear model (GLM) with a binomial distribution considering the climatic variables previously extracted. Finally, the significance of the parameters estimated was tested by a Wald-z statistic, as implemented in the function glm in R 2.10.0.

Results

General composition of visitors

Two hundred and eighty-four floral chambers were collected from the 56 visited locations, containing a total of 11 036 arthropods. The mean number of arthropods trapped per population and per floral chamber were 187.05 (SE = 53.22) and 38.58 (SE = 8.03), respectively.

Absolute overall proportions were as follows: arthropods belonged to classes Arachnida (5.53%) and Insecta (94.47%). The totality of Arachnida was represented by the subclass Acarina. Insecta were composed of orders Diptera (92.68%), Collembola (1.21%), Hemiptera (0.38%) and Coleoptera (0.19%). The far most important proportion of Diptera was formed by the family Psychodidae (87.49%) in which main visitors were females of *Psychoda phalaenoides* (84.27%), followed by *Psycha grisescens* from both genders (2.06% females and 0.70% males); these two species together represented 87.03% of all arthropods trapped in the flower chambers. The remaining Psychodid species were very little represented and accounted for 0.46% of the total sampling.

Based on these results and in order to identify general patterns in the composition of visiting insects, sampled locations were classified according to the relative numbers of the two most abundant species *P. phalaenoides* and *P. grisescens*: locations comprising more *P. phalaenoides* than *P. grisescens* were defined as 'p' sites, while locations comprising more *P. grisescens* than *P. phalaenoides* were considered as 'g' sites (Supplementary material Appendix 2). As shown in Fig. 1, sites 'p' and 'g' not only differed in the relative abundance of the two most frequent species but also in the composition and overall abundance of visitors. In 'p' sites, the composition was strongly dominated by females of *P. phalaenoides*, with very small proportions of other groups, and a particularly high number of visitors (>70) in about half of them. In 'g' sites, the number of visitors was low (a maximum of 77), both males and females of *P. grisescens* were dominant, but other insect groups were also found in non-negligible proportions (Supplementary material Appendix 1).

Geographic variation in the composition of visitors

Figure 2 shows the variation in the composition of floral chamber visitors all over the plant distribution. *Psychoda phalaenoides* is almost found at every location (in blue on Fig. 2) but in varying proportions. Populations in the southern edge of the sampled area include a more diverse fauna, mainly represented by *P. grisescens* (red and pink portions in Fig. 2a). Moreover, when pies are resized proportionally to the total number of visitors (Fig. 2b), a pattern appears, with populations from the south being also less visited than those from the north, and this,

irrespective of the year of collection (data not shown, but see Supplementary material Appendix 1).

Influence of climate on visitors' composition

The composition of the floral chambers' fauna seems to be mainly determined by the first CCA axis (64.95% variance explained), with *P. phalaenoides*, Acarina and *P. setigera* showing an opposite ordination to that of the other groups (Fig. 3a). It is worth noting that the correlation between the abundances of the two most commonly found species (i.e. *P. phalaenoides* and *P. grisescens*) is strongly negative. Regarding climatic variables (Fig. 3b), 'p'-type and 'g'-type populations are differentiated by temperatures, which are negatively correlated to the amount of precipitations: 'g'-type populations are associated with higher temperatures and lower rain rates than are 'p'-type.

The GLM shows that proportions of the two main visitors *P. phalaenoides* and *P. grisescens* are explained by annual precipitations ($p = 0.023$; Table 1) and precipitations during the warmer quarter ($p = 0.007$; Table 1). The proportion of *P. grisescens* increases with higher annual precipitations and lower precipitations during the warmer quarter. The opposite effect is observed for *P. phalaenoides*.

Discussion

A more complete insight on the composition of *A. maculatum* pollinators

Species-specific interactions are rare, because their persistence strongly relies on tight and durable coevolutionary forces. Such conditions are particularly difficult to maintain

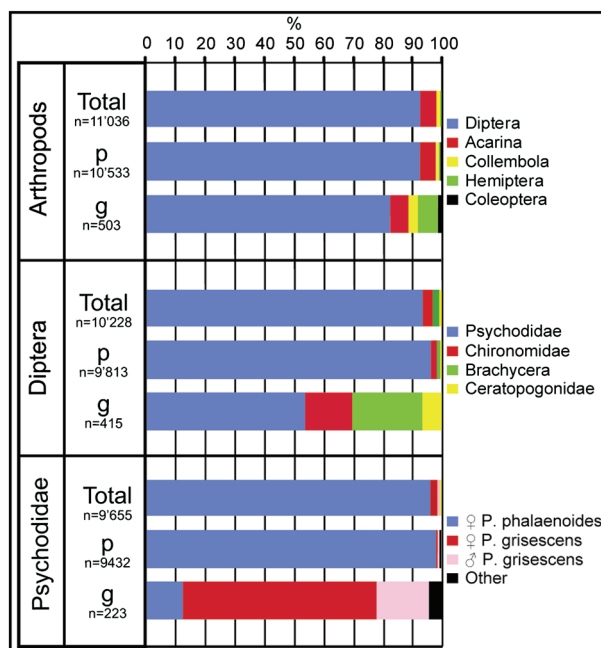


Figure 1. Proportions of Arthropods, Diptera and Psychodidae found in floral chambers. Total numbers according to the two pollinators' community categories are indicated (i.e. 'p': populations harbouring more *P. phalaenoides* than *P. grisescens*; 'g': populations harbouring more *P. grisescens* than *P. phalaenoides*; see text). Legend is shown at the right of the corresponding bars.

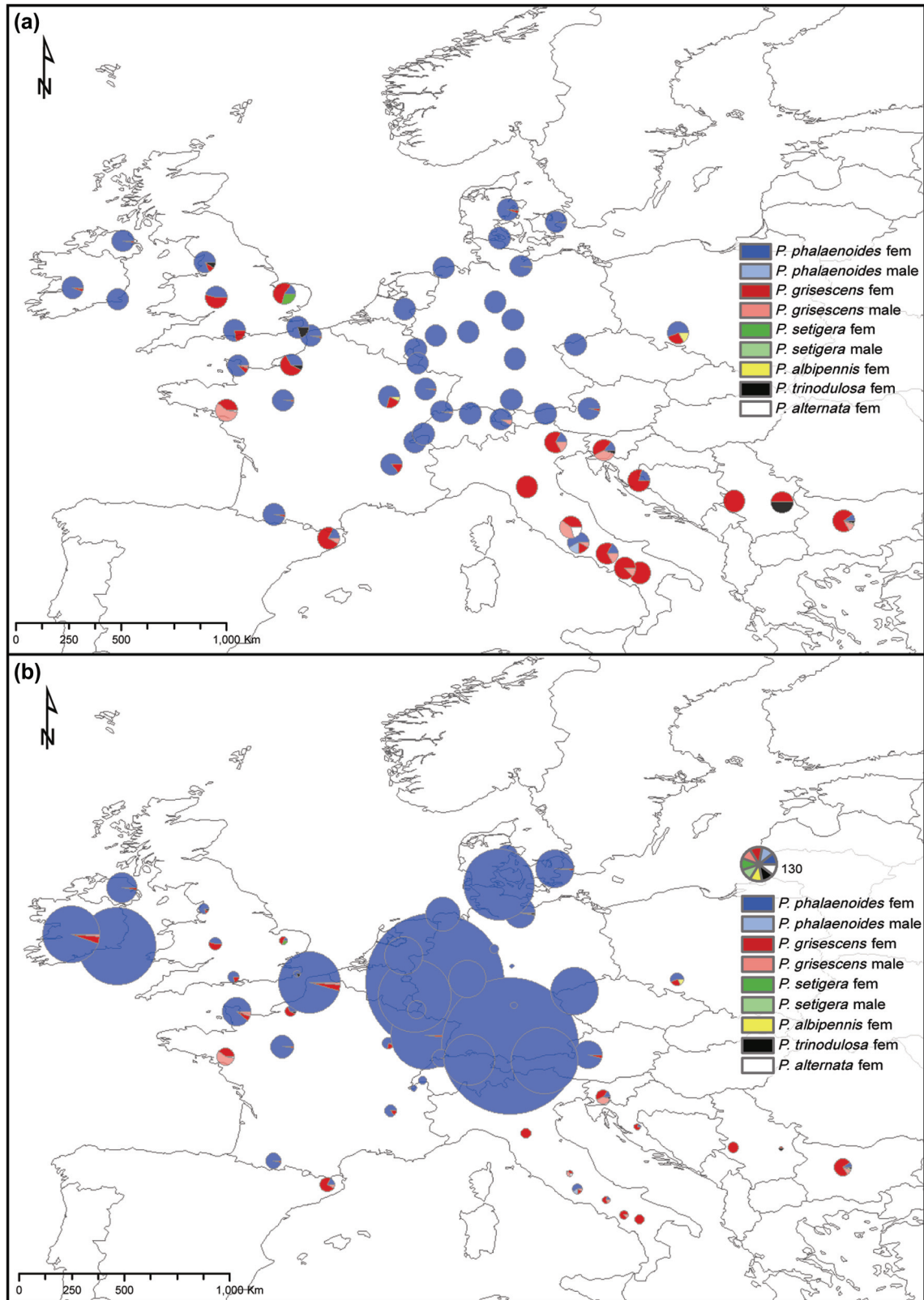


Figure 2. Distribution and composition of Psychodids found in floral chambers. Colours indicate species and sex. (a) proportions of Psychodids per population. (b) proportions resized in function of total number of trapped Psychodids (for a size reference, refer to map legend).

in species with wide distributions, in which environmental pressures are highly heterogeneous across their distribution ranges (Thompson 1994). Accordingly, our results indicate that the reported consistency in the strong and specific

interaction between *A. maculatum* and its main pollinator is not preserved throughout the whole distribution range of the plant: both the total number of visitors and their composition are structured spatially (Fig. 2). Overall, there was

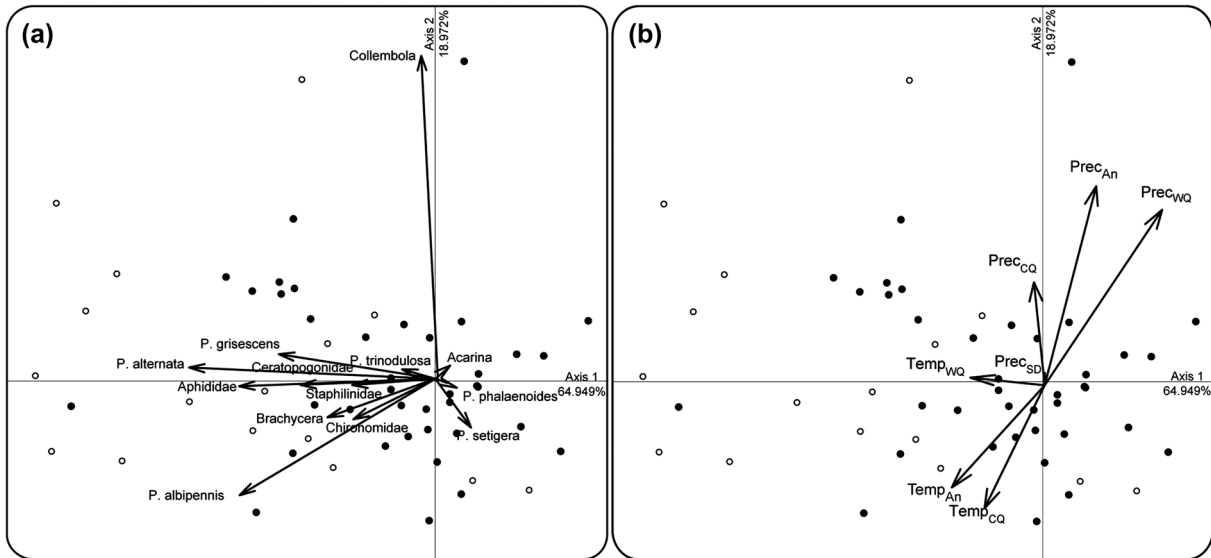


Figure 3. Canonical correspondence analysis for (a) the number and composition of arthropods found in floral chambers and (b) climatic variables. Variables are indicated on each arrow; direction of arrows indicates the sense of correlations. Filled circles represent type-‘p’ populations, while open circles refer to type-‘g’ populations. Percentage of variance explained is shown on axes. Refer to Table 1 for explanations on variables in (b).

a large variation in the abundance of *P. phalaenoides* specimens at each sampled site (ranging from zero to more than a thousand, Fig. 2b). Populations harbouring high numbers of *P. phalaenoides* females were mainly found in central and northern Europe, whereas those hosting a lower number of visitors – generally more diverse and mainly composed of *P. griseus* males and females (82.96%) – were found in the Mediterranean area as well as in northern France (Fig. 2, Supplementary material Appendix 2). The similar behaviours, morphologies and ecologies shared by *P. phalaenoides* and *P. griseus* probably attest to the ability of the latter in transporting *A. maculatum* pollen. However, the pollination effectiveness of *P. griseus* should be investigated in these populations before unambiguously confirming its identity as a real pollinator. Because *A. maculatum* can also reproduce by clonal means (Boyce 1993), further population genetics analyses in the plant all over its southern range of distribution could provide information on its level of cross-pollination and consequently, on the ability of *P. griseus* to efficiently transfer pollen from one plant to another. Finally, other arthropods captured by *A. maculatum* were less abundant and were not structured geographically (Fig. 2). They appear to be more casual and accidental visitors than

reliable pollen vectors, but this does not exclude their potential importance in cross-pollination when main visitors lack.

Variation in the attracted insect genders

Our study points out that *A. maculatum* flowers capture mainly females of *P. phalaenoides*, but both sexes of *P. griseus* (Fig. 1). This result is in agreement with the findings of Diaz and Kite (2002), who described a similar trend in southern England. It is therefore likely that the odour cues released by the plant are different or are interpreted differently by the two species. Whereas *P. phalaenoides* females are attracted by *A. maculatum* odours that mimic their laying substrate (i.e. cow-dung; Kite 1995), one could imagine that this same odour also represents a mating site for *P. griseus* (at least in males). This would be plausible since cases in which insects mate at their laying sites are not rare (Crowson 1981, Petit et al. 2007).

The effect of the environment on plant–insect interactions

Our results show that the composition of arthropods found in floral chambers of *A. maculatum* is correlated with climatic variables (Fig. 3, Table 1). The two most abundant visitors (i.e. *P. phalaenoides* and *P. griseus*) were negatively correlated in the CCA, indicating that there is a strong differentiation of populations visited by one or another species in response to environmental conditions (Fig. 3a): populations dominated by *P. griseus* (type ‘g’, open circles in Fig. 3) were mainly clustered in regions displaying higher summer temperatures combined with lower rain rates. This pattern is also confirmed by the GLM results, in which annual precipitations and precipitations during the warmest quarter were significantly correlated (positively and negatively, respectively) with the relative proportions of *P. griseus* and *P. phalaenoides*.

Table 1. Estimates, z-values and associated p-values of the GLM adjusted to the proportion of *P. griseus* and *P. phalaenoides*. *: $p < 0.05$.

| Climatic variable | Estimate | z-value | p-value |
|--|----------|---------|---------|
| Intercept | -4.793 | -0.74 | 0.460 |
| Mean annual temperature (Temp _{Ann}) | 0.131 | 1.026 | 0.305 |
| Annual precipitations (Prec _{Ann}) | 0.024 | 2.27 | 0.023* |
| Temp. warmest quarter (Temp _{WQ}) | -0.040 | -0.514 | 0.607 |
| Temp. coldest quarter (Temp _{CQ}) | -0.084 | -1.487 | 0.137 |
| Precip. SD (Prec _{SD}) | 0.062 | 1.155 | 0.248 |
| Precip. warmest quarter (Prec _{WQ}) | -0.052 | -2.79 | 0.005* |
| Precip. coldest quarter (Prec _{CQ}) | -0.048 | -1.801 | 0.072 |

It appears that precipitation-related factors modify the visitors' composition and do therefore affect the dynamics of the interaction between *A. maculatum* and its pollinators. These results tend to show that predictions from the theory of the geographic mosaic of coevolution (i.e. variation of the interaction in a variable environment) apply to the case of *A. maculatum*. This is, to our knowledge, one of the few studies (see also Toju 2008) in which the expectations of Thompson's theory match the observed pattern in the context of interacting species, and the first time it is demonstrated in plant–insect antagonistic interactions beneficial to the plant.

The proximal factors underlying this effect might be explained by three non-mutually exclusive hypotheses. First, because most 'g' sites were found in the Mediterranean region – which is characterized by rain regimes different from those of temperate Europe, with at least one marked dry season (Quézel 1985) – water stress might influence the plant phenology, leading it to flower at different time periods, uncoupled with the life cycle of *P. phalaenoides*. As a consequence, it is possible that when Mediterranean *A. maculatum* blossom, their main pollinator *P. phalaenoides* is not present yet/anymore. Second, differences in precipitations could have an impact on the physiology of volatile and heat production, which are known to be both energetically costly (Minorsky 2003) and a key-factor for the attraction of pollinators in this system. Since changes in rain regimes and rates can put plants under water stress and modify their metabolism (Gouinguene and Turlings 2002, Baldwin et al. 2006), one could imagine that *A. maculatum* plants from sites visited mainly by *P. grisescens* present different odour/heat patterns than those visited mainly by *P. phalaenoides*, the latter relying on a much more constant water supply. These differences could make 'g'-type populations less successful than 'p'-type populations in terms of attraction of the most efficient pollinator (i.e. *P. phalaenoides*), this would also be in agreement with both their lower number of visitors and higher arthropod species diversity. Finally, it is also possible that changes in water resources modify the insects' life cycles, limiting their phenological overlap with plants blossom. This seems however unlikely, since faunistic records indicate that both fly species are always present long before and after the flowering period, and over a larger distribution range than that of the plant (Ježek and Barták 2000, Fauna Europaea 2004). An alternative would be that even if *P. phalaenoides* is known to occur in the Mediterranean region, this area constitutes a marginal habitat of lower suitability, in which the species is rather rare compared to more northern habitats. In order to test this hypothesis, further intensive entomological collections should be performed all over the insect's distribution range.

Conclusion

In this study we show that arthropod visitors of *A. maculatum* are mainly restricted to the two Psychodids *P. phalaenoides* and *P. grisescens*. Overall, as previously observed at a more limited geographical scale (Beck 1983, Diaz and Kite 2002), *P. phalaenoides* females are the most abundant visitors of *A. maculatum* flowers. However, the abundance and

composition of the fauna captured in floral chambers strongly varies across the distribution range of the plant, with *P. grisescens* becoming notably more abundant in the Mediterranean region. As a consequence, diversity and abundance of arthropods are geographically structured (without any notable inter-annual effect), indicating that all over its distribution range the plant might exploit at least two species and not a single one, as previously assumed. Our results show that variation in environmental conditions, particularly in rain regimes, is correlated with changes in the visitors' composition. This climatic effect could be explained by the role played by water stress in modifying the phenology of either the plant or the insect, or the physiology of volatile and heat production in the plant. Precipitation-related factors therefore affect not only the visitors' composition of the plant, but also the specificity of the pollination interaction.

This study provides evidences confirming predictions of the theory of the geographic mosaic of coevolution, directly demonstrating that environmental variation does have an impact on antagonistic interactions. It finally points out the importance of applying large-scale surveys when challenging the existence of putative one-to-one interactions.

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Supplementary material (available online as Appendix O18937 at <www.oikos.ekol.lu.se/appendix>). Appendix 1–2.

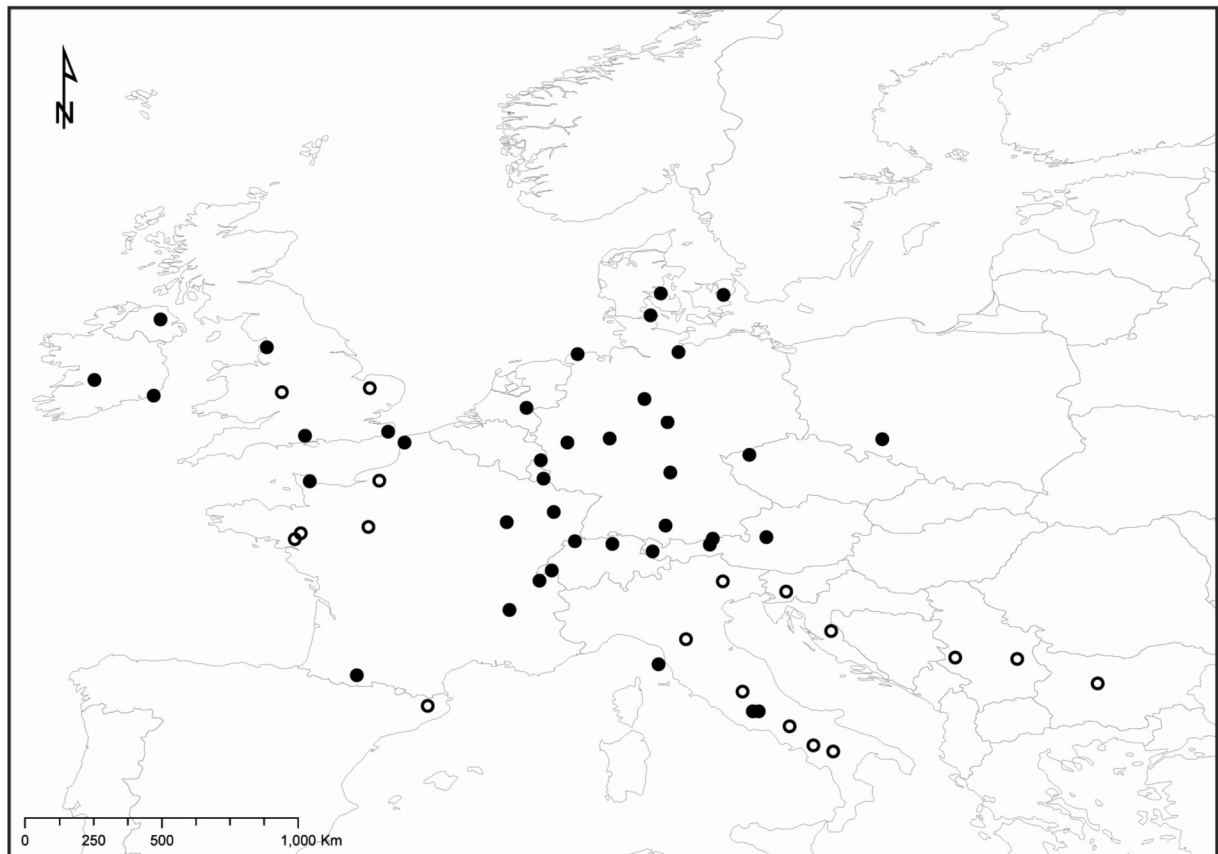
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Appendix 1

List of populations and visitors' composition.

Appendix 2

Sampling distribution of *A. maculatum* visitors. Open circles: type-‘g’ populations; full circles: type-‘p’ populations.



Chapter four

Size-advantage model and pollination strategies in plants

Natacha Revel, Marc Gibernau,
Nadir Alvarez and Anahí Espíndola

To be resubmitted to *Evolutionary Ecology*

1 **Size-advantage model and pollination strategies in plants**

2 Natacha Revel¹, Nadir Alvarez², Marc Gibernau³, Anahí Espíndola^{1 4}

3 ¹ Laboratory of Evolutionary Entomology, Institute of Biology, University of
4 Neuchâtel, Emile-Argand 11, 2009 Neuchâtel, Switzerland.

5 natacha.revel@gmail.com

6 ² Department of Ecology and Evolution, University of Lausanne, Biophore Building,
7 CH-1015 Lausanne, Switzerland. nadir.alvarez@unil.ch

8 ³ CNRS-UMR 8172 Ecologie des Forêts de Guyane, BP 709, 97387 Kourou, France.
9 Marc.Gibernau@ecofog.gf

10 ⁴ Corresponding author. Laboratory of Evolutionary Entomology, Institute of Biology,
11 University of Neuchâtel, Emile-Argand 11, 2009 Neuchâtel, Switzerland.

12 maria.espindola@unine.ch.

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26 Abstract

27 The size-advantage model (SAM) is aimed at explaining the temporal variation of
28 energetic investment on reproductive structures in long-lived hermaphroditic
29 organisms. The theory proposes that an increase in the resources available to an
30 organism induces a higher relative investment on the energetically most costly
31 sexual structures. The model has been demonstrated to apply to both animals and
32 plants, and different factors (e.g., habitat quality, capability of dispersion) appear to
33 shape the benefit curve of the SAM expression.

34 In plants, pollination interactions are known to play a role in the evolution of flower
35 features. Because the SAM directly concerns flower characters, pollinators are
36 expected to have an influence on the application of the model. This hypothesis has
37 however never been tested.

38 In the present work, we investigate whether the identity and diversity of pollinators
39 can be used as a proxy to predict the application of the SAM in exclusive zoophilous
40 plants. We present a new approach to understand the dynamics of the model and
41 test it on pollination systems involving widespread central-European *Arum* (Araceae)
42 species (*A. cylindraceum*, *A. maculatum* and *A. italicum*).

43 By identifying the species composition, abundance and spatial variation of
44 arthropods trapped in the inflorescences we show that *A. cylindraceum* displays a
45 generalist reproductive strategy, relying on the exploitation of a low number of
46 dipterans, similarly to what is found in *A. italicum*, but contrasting with the pattern
47 depicted in *A. maculatum*. Based on the simple model presented here, the
48 application of the SAM is predicted for the first two and rejected for the last species,
49 predictions being further confirmed by allometric measures.

50 This is to our knowledge the first time that this theory is both proposed and
51 empirically tested in zoophilous pollination systems. Its application on other non-
52 *Arum* systems is discussed, indicating its general biological importance.

53

54 Introduction

55 Energetic income is limited in all organisms and natural selection optimizes
56 resource allocation strategies between growth/survival and reproduction according to
57 physiological and environmental (biotic and abiotic) factors (Guillon et al. 2006; Lloyd
58 and Bawa 1984; Policansky 1982). As a consequence, it has been proposed that in
59 hermaphroditic and long-lived organisms the reproductive energetic allocation varies
60 with the size of the organism. In general, when available resources increase, the size
61 of an organism tends to augment, which favours the relative increase of the most
62 costly sexual structures. This model –namely the Size-Advantage Model (SAM)–
63 was first proposed (Ghiselin 1969) and tested (Warner 1988) in animals, but was
64 later applied to dioecious and hermaphroditic plants (Lloyd and Bawa 1984), *e.g.*, in
65 species within genera *Acer* L. (Sapindaceae) (Primack and McCall 1986), *Arisaema*
66 Mart. (Araceae) (Vitt et al. 2003) and *Arum* L. (Araceae) (Méndez 2001).

67 According to the model, floral resource allocation, *i.e.*, the quantity of energy
68 invested by a plant in the production of female vs. male sexual structures (Lloyd and
69 Bawa 1984), depends on the energy available at a certain time. When only the
70 amount of available resources varies, larger inflorescences -found in larger plants-
71 are predicted to harbour a larger proportion of female flowers (*i.e.*, the most
72 energetically costly sexual structure) than smaller inflorescences. Since its first
73 application, the comprehension of the SAM has been progressively enhanced by the
74 inclusion of additional factors such as local sexual competition and differential
75 gamete dispersion (Lloyd and Bawa 1984), wind-pollination syndromes (Burd and
76 Allen 1988), plant geometry (Bickel and Freeman 1993) or variation in environmental
77 quality (Guillon et al. 2006). However, even if pollination biology is known to strongly
78 shape floral characters (Raven et al. 2005), no studies have tested yet its influence
79 on the application of the SAM within the context of biotic pollen vectors. This gap

80 might be due to the scarcity of studies combining careful examination of pollinators
81 with accurate surveys of flower allometric characters, but also to the complexity of
82 some mathematical models developed to understand the SAM (Guillon et al. 2006).

83 Despite the apparent complexity of this topic, some simple predictions can be
84 made regarding the possible influence of both the efficiency and diversity of
85 pollinators on the investment towards a given gamete type. Lloyd and Bawa (1984)
86 first proposed that a gamete positive-bias is expected for the gamete dispersing
87 longer distances, since this would avoid intra-plant gamete competition. Here, we re-
88 visit this hypothesis from the point of view of the pollinators. In an exclusive
89 zoophilous plant, in which pollen is strictly dispersed by animals, pollen dispersion is
90 directly related to pollinator efficiency (a function of both the number of pollinators
91 visiting a flower and of their individual capability to effectively cross-pollinate). As a
92 consequence, if the probability of cross-pollination is low because pollinators are
93 scarce, present a low ability to cross-pollinate, or both, we predict that the SAM
94 should apply: gametes should be biased towards female function because pollen
95 dispersion is expected to be low. In addition, by investing on female flowers, the
96 number of pollinated ovules should be quickly increased as soon as an eventual
97 pollen-carrier visits the flower. Furthermore, an investment towards male gametes in
98 such a situation would translate into a large loss of these gametes. On the other
99 hand, if the probability of cross-pollination is high because pollinators are very
100 abundant, cross-pollination is efficient, or both, plants should not match to the SAM,
101 since it is likely that the pollen produced will present a high probability of reaching
102 another flower, diminishing a potential intra-plant gamete competition. Conforming to
103 the SAM in such a context is expected to be counter-selected because the plant has
104 the possibility to favour both its fruit set and male gamete dispersion (Chartier and
105 Gibernau 2009).

106 As stated above and in addition to pollinator efficiency, we also consider
107 pollinator diversity (in terms of number of different taxa visiting a flower) as a
108 possible factor influencing the application of the SAM. In fact, the overall probability
109 of cross-pollination is explained by the interaction of two main factors: pollinator
110 diversity and efficiency. Pollinator diversity in a given plant can be considered as a
111 proxy for the level of specificity in a pollination interaction (along a continuum
112 between generalist and species-specific pollination, Ollerton et al. 2007), and thus
113 indirectly informs on the probability that a given pollinator will carry specific pollen.
114 The nature of this interaction (*i.e.*, specific or generalist) combined with the individual
115 efficiency of the pollinator (*i.e.*, its visitation rate and constancy) finally shapes the
116 probability of cross-pollination for a given pollinator. We expect that very efficient and
117 specific pollinators would provide a final global high probability of cross-pollination,
118 which would not favour the application of the SAM (high pollen dispersion, low intra-
119 plant gamete competition), whereas selection towards SAM expression should be
120 advantaged under generalist and inefficient visits (low pollen dispersion and high
121 intra-plant gamete competition). Moreover, the shape of the probability curve for the
122 application of the SAM is expected to be different for cases in which specific *vs.*
123 generalist pollinators are involved (*i.e.*, with a faster decrease of intra-plant pollen
124 competition in the case of specific visitors): if pollen load gets saturated the visit of a
125 reduced number of pollinators from a few species will be translated into a higher
126 relative probability of receiving pollen from the “correct” species than if visitors were
127 generalists. Predictions regarding the application of the SAM as a function of both
128 pollinator diversity and efficiency are summarized in Figure 1.

129 Geophytic species of the Araceae are well-suited to test hypotheses within the
130 SAM framework since inflorescence and plant-size in this plant family are tightly
131 related to reserves stored in the tuber (Méndez and Obeso 1993; Vitt et al. 2003)

132 and increase with the age of the plant. Related to this point, it has been
133 demonstrated for example that *Arum italicum* Mill. and *A. idaeum* Coust. & Gand. do
134 not produce any inflorescence until the tuber mass has reached a threshold weight,
135 after which larger plants produce more inflorescences than smaller ones (Méndez
136 and Obeso 1993). This physiological feature would indicate that there are genetic
137 constraints intrinsic to the species which make the relationship bulb size -
138 reproductive allocation specific to a species and less variable at a regional level.

139 The simplest view of the SAM in the case of Araceae would thus predict a trend
140 towards feminization in larger inflorescences. Within bulbous Araceae, the genus
141 *Arum* is particularly suitable for testing hypotheses related to the effect of pollinator
142 efficiency and diversity on the SAM. It includes protogynous, thus generally self-
143 infertile species, which present similar ecologies and biologies (Boyce 1993) and
144 express insect-pollination syndromes with different degrees of pollinator specificity
145 (Gibernau et al. 2004). However, the application of the SAM is heterogeneous within
146 the genus (see below). The genus *Arum* comprises 28 species, most of them with a
147 peri-Mediterranean distribution (Boyce 1993; Espíndola et al. 2010). As all members
148 of the Araceae family, plants are characterized by singular floral structures, including
149 a spadix harbouring flowers, surrounded by a spathe, *i.e.*, a modified bract (Mayo et
150 al. 1997). A special feature of the genus is that this basic floral structure is adapted
151 to the attraction and trapping of pollinators (lure-and-trap antagonistic pollination). In
152 all but one species (*i.e.*, *A. creticum* Boiss. & Heldr.) a contraction of the spathe
153 takes place just above the fertile flowers, where sterile flowers transformed into hairs
154 (*i.e.*, staminodes) delimitate a closed floral chamber (Boyce 1993). Another
155 remarkable feature of *Arum* species is their ability to produce heat and odour during
156 anthesis in order to attract and temporarily capture potential pollinators (Gibernau et
157 al. 2004). Probably because of the substantial amount of energy involved both in the

158 characteristic floral structure and in the heat and odour production, a trend towards
159 specificity in pollinator attraction is generally detected (Gibernau 2003). However,
160 there is strong variation in pollinator diversity levels across the different *Arum*
161 species. At one end of the continuum are species such as the specialist *A.*
162 *maculatum* L., a widespread European species, which shows a highly specific
163 interaction with two species of the tribe Psychodini [*i.e.*, *Psychoda phalaenoides* L.
164 and *Psycha grisescens* Tonnoir (Diptera: Psychodidae); Espíndola et al. in press]. At
165 the other end are species such as the generalist *A. italicum* Mill. –also widely
166 distributed in Europe– whose efficiency in attracting pollinators is lower and is visited
167 by several arthropods from different families and orders (Albre et al. 2003). Whereas
168 the latter species has been shown to follow the SAM (Albre and Gibernau 2008;
169 Méndez 2001), the first does not do so (Chartier and Gibernau 2009). These two
170 species therefore confirm predictions illustrated in Figure 1: plants visited by efficient
171 and specific pollinators should not favour the SAM, while those visited by rare and
172 generalist pollinators should increase their fitness by following this model. However,
173 it is difficult to confirm this theory based on these two single cases. For this reason,
174 we propose to further test our predictions by investigating another widely-distributed
175 European *Arum* species, *A. cylindraceum* Gasp. The choice of this species is based
176 on its wide distribution range similar to those of the other currently well-known *Arum*
177 species, thus decreasing possible effects of other untested external factors (*e.g.*,
178 environmental features, local adaptations) on the application of the SAM.

179 *Arum cylindraceum* is a recently rediscovered and redescribed species (Bedalov
180 and Küpfer 2005; Fridlender 1999), whose ecology and biology are currently not well
181 known. Its centre of distribution is Eastern Europe, being present from Spain and the
182 Balkans to Poland and Denmark (Bedalov and Küpfer 2005). Until now, no studies
183 have addressed the question of the identity of pollinators in this species, even if it

184 likely presents some level of specificity, as shown in other species within *Arum*
185 (Gibernau et al. 2004). In the present work, we investigate the pollination strategy of
186 *A. cylindraceum* at the species level and use it as a third case study to test the
187 validity and wider application of our predictions on the SAM as a function of pollinator
188 efficiency and diversity. In order to identify an eventual specificity in the pollination
189 interaction and to get a large-scale overview of its geographical consistency, we
190 studied the species composition in arthropod visitors –and thus potential pollinators–
191 throughout the distribution range of the plant. This allowed us evaluating both the
192 reproductive strategy of the species (*i.e.*, specific vs. generalist) and the level of
193 efficiency in pollination as a function of the number of visitors attracted. Using these
194 cues and applying the predictions presented above (see Fig. 1) we compared results
195 in efficiency and diversity of pollinators with allometric measures taken on the
196 inflorescences of the sampled plants and tested the following expectations: i) a
197 combination of high insect diversity and low number of visitors is expected to be
198 found in plants conforming to the SAM, because the probability of cross-pollination is
199 expected to be low (as in *A. italicum*); ii) a low insect diversity combined with a high
200 number of visitors should be related with the non-conformity to the SAM, since the
201 probability of cross-pollination is expected to be high (like in *A. maculatum*); iii) a
202 combination of low insect diversity and number is expected in plants matching to the
203 SAM (low net probability of cross-pollination, especially if the number of pollinators is
204 extremely low); iv) a combination of high insect diversity and number is expected to
205 be present in plants which do not match to the SAM (high net probabilities of cross-
206 pollination).

207

208 **Material and methods**

209 **Sampling and pollinator identification**

210 *Arum cylindraceum* populations were sampled from April to May 2008, almost
211 completely covering its distribution range. Considering the generally small population
212 sizes (less than 80 plants) and highly aggregated distribution of the plants in
213 populations, we excluded the effect that clonal reproduction (a feature known for the
214 genus, Boyce 1993) could have in our results by restricting the number and
215 distribution of sampled individuals. This is why we selected up to five floral chambers
216 per population from plants distant by more than five meters from their closest
217 sampled neighbour. Because we studied the complete distribution range of the plant,
218 and since we were interested in having a view of the strategy displayed by the
219 species in general and not necessarily at each sampled location, we consider that
220 this sampling is representative of the main strategy displayed by the species and has
221 the advantage of not being biased by locally high clonal reproduction. Once floral
222 chambers were collected, they were labelled and independently preserved in a
223 Falcon tube filled with 70% ethanol. A plant voucher was collected at each location,
224 and geographic coordinates were registered using a GPS Etrex Garmin.

225 Arthropods were identified to the order level (Freude et al. 1964). Diptera
226 families were indentified at the species level. Because of the lack of updated
227 identification keys for several Diptera families, European specialists were consulted
228 for the identification of samples belonging to this group. An exception is the
229 Psychodidae family, for which keys are actually available (Jezek 1990; Withers
230 1988). Occurrences of each species/genus/family were recorded and numbers per
231 floral chamber, per population and in total were calculated, as well as the sex-ratio of
232 the most abundant visitors.

233 Because the identity and amount of pollinators have been shown to vary
234 geographically in other *Arum* species (Espíndola et al. in press) we tested for an

235 eventual spatial structure by analyzing the geographic visitor composition using
236 ArcMap 9.3 (ESRI, Redlands CA, USA).

237 **Allometric measures**

238 Out of all analyzed plants, a maximum of three inflorescences per population
239 (n=39) were selected to perform allometric measures (lengths of the spadix -see
240 below-, female and male zones). Numbers of male and female flowers were counted
241 and floral sex ratios calculated. The floral sex ratio was inferred as: number of male
242 flowers/(number of male flowers + number of female flowers), where 1 represents a
243 complete male, and 0 indicates a complete female. Following the approach
244 established by Chartier and Gibernau (2009), regions of the inflorescence were
245 divided into fertile male and female zones, and their lengths were measured with a
246 digital calliper (TESA, model CCMA-M). Because the addition of fertile and sterile
247 zones is correlated in a linear manner with the total size of the inflorescence (M.
248 Gibernau, unpublished), we measured the length from the base of the female fertile
249 zone to the last upper staminodes and used this value as a representative of the
250 total spadix length, a value correlated with the total plant size, and thus with
251 available resources (Albre et al. 2003).

252 Descriptive statistics of these measures and correlations (linear regressions for
253 lengths; logistic regressions for counts) were calculated, applying the functions `lm`
254 and `glm` implemented in R 2.9.0 (R Core Development Team, 2009). Because the
255 present study was interested on general floral patterns intrinsic to the species, and
256 because values presented no special population differentiation (see below), all
257 analyses were done at the interpopulation level.

258 **Testing predictions**

259 In order to test predictions made by our model, we applied a comparative
260 method. Based on the data obtained from the insect identification, we inferred their

261 diversity and efficiency. Diversity was inferred following the approach applied by
262 Ollerton et al. (2007), which defined the specificity as a measure of the number of
263 interacting species. Efficiency was estimated by considering the total number, and
264 the constancy in the composition of visitors from a spatial point of view, since it is
265 unlikely that a group of rare, diverse and inconstant visitors presents a high overall
266 pollination efficiency, while the opposite is true for abundant and specific visitors.

267 The correctness of the predictive model (Fig. 1) was thus inferred by using
268 indirect measures of diversity and efficiency to identify the theoretical expectations
269 corresponding to the different combinations addressed, and further comparing them
270 with the results obtained in the allometric survey. Congruence between these
271 allometric results and expectations would provide support to the model.

272

273 **Results**

274 **Visitors' diversity and distribution**

275 A total of 24 populations were sampled (Appendix 1) and 80 floral chambers
276 collected, containing 252 arthropods distributed in half of the populations (12
277 populations did not comprise any visitor). This represented a mean of 3.50
278 (SE=0.57) and 12.75 (SE= 3.64) arthropods per floral chamber and per population,
279 respectively.

280 Figure 2 shows the diversity in collected visitors, restricted to Arthropods and
281 represented by the classes Insecta (67.9%), Arachnida (Acarina, 18.2%), Diplura
282 (7.1%) and Collembola (6.7%). The two most frequent orders were Diptera (65.5%)
283 and Coleoptera (28.1%). The order Diptera comprised families Psychodidae
284 (57.1%), Ceratopogonidae (14.3%), Sphaeroceridae (10.7%), Sciaridae (7.1%),
285 Cecidomyiidae (3.6%), Chironomidae (3.6%), and Mycetophilidae and Phoridae
286 (3.6%).

287 Regarding the diversity found in each Diptera family, our results show that
288 Psychodids were represented by a large majority of *Psycha grisescens* (82.8%,
289 mainly males) and lower proportions of females of *Psychoda phalaenoides* (14.1%)
290 and *P. albipennis* Zett. (3.1%). Ceratopogonids were identified as belonging to the
291 species *Culicoides obsoletus* Meigen (94%) and *Atrichopogon oedemerarum* Stora
292 (6%). Species present in the family Sphaeroceridae were *Spelobia clunipes* Meigen
293 (58.3%), *Coproica ferruginata* Stenhammar (33.3%) and *C. vagans* Haliday (8.4%).
294 Concerning the Sciarids present in the samples, we could only identify them to the
295 two genera *Bradysia* Winnertz (63%) and *Phytosciara* Frey (25%). Samples of the
296 remaining families were very scarce (less than 11% in total): taxa were *Smittia*
297 *leucopogon* Meigen (Chironomidae), species belonging to genera *Azana* Walker and
298 *Boletina* Staeger. (Mycetophilidae) and to tribes Orthoclaadiinae (Chironomidae),
299 Lestremiinae and Cecidomyiinae (Cecidomyiidae). No sub-generic identification was
300 possible on the unique Phorid sample.

301 The spatial distribution of these groups is not geographically structured in terms
302 of composition (Fig. 3a) or of number of visitors trapped (Fig. 3b).

303

304 **Allometric measures**

305 There is a logical positive correlation between the length of each floral zone (*i.e.*,
306 male, female and sterile) and the total inflorescence length (Fig. 4a). The increase in
307 the length of the sterile floral zone was however the highest (slope=0.562), followed
308 by female (slope=0.334), and finally by male (slope=0.103) zones. Each factor
309 significantly increased the adjustment of the general linear model indicating that the
310 three slopes are different ($p < 0.001$ in all the cases, $R^2 = 0.946$, $F = 379.2$).

311 Another positive relation is observed for the number of fertile flowers and the
312 total inflorescence length (Fig. 4b). The slope is higher for the number of female

313 (slope= 0.462) than for male (slope=0.225) flowers, and both slopes are significantly
314 different ($p < 0.05$, $z_{(fem)} = 5.781$, $z_{(male)} = -2.425$).

315 The inflorescence floral sex-ratio is negatively correlated (slope= -0.063,
316 $R^2 = 0.094$, $F = 4.626$, $p < 0.05$) with the total inflorescence length (Fig. 4c), indicating a
317 significant feminization of the inflorescence with an increase of the total size.

318 There was no significant correlation between the number of trapped insects and
319 the total length of the inflorescence ($p > 0.1$; Fig. 4d). This result was maintained even
320 when samples presenting or not insects were considered (not shown) and despite
321 the fact that mean total lengths were significantly different between the two groups
322 ($F = 4.992$; $df = 1$; $p < 0.05$).

323 Correlations showed no population structure (data not shown), indicating that the
324 observed patterns are intrinsic to the species and not to a specific region.

325

326 **Interaction between SAM and visitors**

327 In a comparative framework, our results showed that *A. cylindraceum* presents a
328 visitor pattern combining the presence of high visitor diversities (Fig. 2) with low
329 numbers of spatially highly variable compositions of visitors (low expected efficiency,
330 Fig. 3), which represents the combination “high diversity – low efficiency” predicting
331 the application of the SAM (Fig. 1).

332 The survey on the variation of reproductive allocation indicates that the plant
333 modifies the proportion of one or the other flower type in function of the available
334 resources (Figure 4a-c), indicating that the SAM is applied in *A. cylindraceum*.

335

336 **Discussion**

337 **The diversity and efficiency of *A. cylindraceum* visitors**

338 Compared to other *Arum* species (Espíndola et al. in press; Albre et al., 2003),
339 our results show that visitors of *A. cylindraceum* are scarce (3.5 per floral chamber
340 and completely absent of half of the populations) and highly diverse comprising
341 species from several insect groups. Because the plant does not seem prone to self-
342 fertilization in the European continent (Chouteau et al. 2008; Fridlender 1999), such
343 a low number of trapped insects was not expected. Since several species within the
344 genus can reproduce by forming bulbs in a vegetative manner (Boyce 1993), it is
345 likely that *A. cylindraceum* can reproduce by clonal means (Fridlender 1999) and that
346 cross-pollination may rarely happen. However, during our sampling trips, it was
347 frequent to find *A. cylindraceum* plants displaying fruits (A. Espíndola, pers. obs.).
348 Another explanation for such a low number of sampled insects could be that this
349 species is effectively visited by more arthropods than those found in the present
350 study, but that the trapping system (*i.e.*, the number and length of staminodes) is not
351 highly efficient, releasing arthropods after a short span of time spent in the floral
352 chamber. Such a non-optimal strategy would require a smaller energetic investment
353 in terms of odour (*e.g.* less specific volatiles), heat and morphological features (*e.g.*
354 length of staminodes). It also might be sufficient to ensure pollination of several
355 female flowers during the lifespan of a plant, if at least one insect carrying pollen
356 visits the inflorescence (*i.e.*, as shown in *A. maculatum* (Lack and Diaz 1991)).
357 However, this strategy would bring the problem that if visitors are not trapped quickly
358 after the first trapping, pollen would lose its viability (Gibernau and Macquart 2003)
359 and pollination would not occur if the pollen carrier visited a second flower. Long-
360 lasting observation and sampling of all insects visiting the flowers during the whole
361 anthesis would allow testing these ideas. Additionally, investigating the variation of
362 pollen viability through time would also inform on the temporal range during which

363 pollen carry-over needs to happen, and would provide indirect information on the
364 natural dynamics of pollination.

365 Despite showing a low total number of visitors, our results still reveal a high
366 diversity of insects and especially of Diptera (Fig. 2), as shown for some other *Arum*
367 species (Gibernau et al. 2004). All Dipteran families recorded here (with the
368 exception of the three less abundant ones, *i.e.*, Cecidomyiidae, Mycetophilidae and
369 Phoridae) have been previously identified as visitors and thus potential pollinators of
370 several other species within the genus, as for example *A. dioscoridis* and *A. orientale*
371 (for a review, see Gibernau et al. 2004). Moreover, their simultaneous occurrence
372 was also recorded in the floral chambers of *A. italicum*, where they were confirmed
373 as effective pollinators (Albre et al. 2003). Considering these reasons, we feel
374 confident to consider that insects trapped in the present study are very probably
375 reliable pollinators of *A. cylindraceum*.

376 In contrast to a recent study on *A. maculatum* (Espíndola et al. in press), the
377 spatial distribution of visitors' composition does not show any specific geographic
378 pattern (Fig. 3), indicating that the pollination strategy applied by the plant is intrinsic
379 to the species and is not a result of local adaptation. Consequently, the plant
380 appears as a widespread generalist, presenting little efficiency in pollinator attraction.

381 ***Arum cylindraceum* in the SAM perspective**

382 Because of its low efficiency and generalist behaviour, *A. cylindraceum* should
383 be expected to conform to the SAM (Fig. 1). This prediction is confirmed by our
384 results on the variation in the resource allocation applied by the plant (Fig. 4): there
385 is an increasing investment towards female function as inflorescence size becomes
386 larger. As shown in Fig. 4b, the increase in the female floral zone is mostly due to an
387 increase in the number of female flowers, indicating that not only the size but also
388 the number of female flowers increases in larger plants. In agreement with the

389 similar pattern found in *A. italicum* (Albre and Gibernau 2008), the mean floral sex
390 ratio of the plant is biased towards the male function, but significantly feminizes
391 when the inflorescence size increases (Fig. 4c). This trend holds even if both flower
392 types increase in number (Fig. 4b). Our simple SAM-pollinator model is again
393 confirmed, indicating that its rules seem biologically correct.

394 In *Arum*, it has been demonstrated that the augmentation in plant size is
395 associated with an increase in the size of the organ involved in heat and odour
396 production -the appendix- (Gibernau and Albre 2008), and a subsequent raise of
397 pollinator attraction (Méndez and Obeso 1992). Our analyses however show that in
398 *A. cylindraceum* the increase in inflorescence size is not correlated with a gain in
399 attraction power (Fig. 4d). Moreover, this result corroborates our view of the
400 interaction between *A. cylindraceum* and its associated pollinators as a low-
401 efficiency pollination system.

402 In the same way that we could not identify any geographic structure in the
403 composition of insects, we could not find any local variation of floral characters
404 (samples from the same locations were randomly distributed in the point clouds; Fig.
405 4). Moreover, points from all locations were clearly distributed along a unique and
406 continuous regression line, which would not be expected if there were local
407 specializations. This would indicate –as previously thought- that the relationship
408 between bulb size and floral allocation is genetic and thus intrinsic to the species. A
409 confirmation of this relationship would however require measuring bulb weights and
410 floral allocations both at local and general scales.

411 The combination of pollinator composition and plant allometric measures
412 confirmed our expectations in long-lived zoophilous species. This is to our
413 knowledge the first time that this theory is both explicitly proposed and tested in an
414 empirical pollination system. It is important to note that our predictions in Araceae

415 might also apply to other plant groups, as for example to species of the genus
416 *Trillium* L. (Melanthiaceae) (Wright and Barrett 1999), in which low pollination
417 visitation with application of the SAM has been registered. Nevertheless, in order to
418 extend the understanding of the dynamics of the SAM as a function of pollinator
419 diversity and efficiency, it is necessary to analyze a wider number of plant-insect
420 systems taking into account not only the variation of sex-resource allocation, but also
421 pollinator diversity. In this context, the family Araceae is particularly suitable because
422 biological (e.g., long-lived plants) and ecological characters (e.g., generalised
423 zoophily) shared by its species greatly simplify the comprehension of SAM-related
424 floral allocation responses. Finally, the application and development of dynamic
425 mathematical models in the SAM/pollinator context could provide important
426 information on the exact expected shapes of the probability curves proposed by our
427 predictions, and would thus help deepen our understanding of conditions guiding the
428 variation of floral allocation.

429

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437

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511 Appendix 1 – Sampled locations.

| Locality | Country | Collection date | Altitude (m) | Latitude | Longitude |
|----------------------|-----------|-----------------|--------------|----------|-----------|
| Mont Iti Kato | Greece | 20.07.2007 | 1377 | 38.80973 | 22.20632 |
| Gross Disnack | Germany | 15.04.2008 | 42 | 53.72313 | 10.70769 |
| Schmilau | Germany | 15.04.2008 | -8 | 53.66833 | 10.7575 |
| Purkersdorf | Austria | 16.04.2008 | 283 | 48.20223 | 16.17298 |
| Herl'any | Slovakia | 20.04.2008 | 300 | 48.79885 | 21.48015 |
| Tylawa | Poland | 22.04.2008 | 350 | 49.46503 | 21.7323 |
| Sisteron | France | 26.04.2008 | 508 | 44.21825 | 5.91398 |
| Vondrou | Greece | 25.05.2008 | 685 | 41.27574 | 23.74667 |
| Chaliki | Greece | 26.05.2008 | 1132 | 39.68423 | 21.19218 |
| Delvinaki | Greece | 29.06.2008 | 827 | 39.92821 | 20.46182 |
| Anilio | Greece | 26.05.2008 | 1369 | 39.73474 | 21.19969 |
| Sokobanja Vrh | Serbia | 23.05.2008 | 917 | 43.58153 | 21.89266 |
| Kalvehale | Denmark | 24.04.2008 | 2 | 54.99954 | 12.16737 |
| Villetta Barrea | Italy | 28.05.2008 | 1258 | 41.79683 | 13.93833 |
| Gargano | Italy | 29.05.2007 | 703 | 41.7937 | 15.97829 |
| Zobor | Slovakia | 19.04.2008 | 310 | 48.33696 | 18.09889 |
| Gioia Vecchio | Italy | 29.05.2008 | 1390 | 41.90208 | 13.73339 |
| Bucje | Serbia | 22.05.2008 | 669 | 44.15512 | 22.11236 |
| Gylling | Denmark | 25.04.2008 | 25 | 55.84845 | 10.16711 |
| Traiskirchen | Austria | 17.04.2008 | 207 | 48.00996 | 16.29883 |
| Solosnica | Slovakia | 18.04.2008 | 170 | 48.4538 | 17.23844 |
| Muszkowice | Poland | 23.04.2008 | 226 | 50.64387 | 16.95121 |
| Church of Taxiarches | Greece | 30.04.2007 | 832 | 32.87914 | 20.67902 |
| Trpejca | Macedonia | 25.05.2008 | 1361 | 40.96949 | 20.85911 |

512

513 Table 1 – Descriptive statistics of allometric measures in inflorescences of *Arum*
 514 *cylindraceum*. SE: Standard Error.

| | | Mean | SE |
|--------------------------------------|----------------|-------|-------|
| Number of Flowers | female | 38.92 | 1.67 |
| | male | 71.61 | 2.70 |
| Floral zone length (cm) | female | 0.92 | 0.028 |
| | male | 0.46 | 0.012 |
| | sterile | 1.44 | 0.039 |
| Total length (cm)¹ | | 2.81 | 0.035 |
| Sex-Ratio | | 0.65 | 0.011 |

515 ¹from the base of the female zone up to the highest staminode (not including the
 516 stipe and the appendix).

517

518 Figures legends

519 Figure 1 – Predictions of application of the size-advantage model, considering
 520 pollinator diversity and efficiency in zoophilous plants. Because variables are
 521 continuous, grey scales indicate the expected variation of the probability of
 522 application of the model (dark: high probability, white: low probability). Note that in
 523 specific interactions, the application of the SAM is expected to be advantageous
 524 mainly at very low visitation rates (low net pollination efficiency), since specificity in
 525 plant-insect interactions is generally associated with highly efficient individual cross-
 526 pollination due to coevolutionary processes.

527

528 Figure 2 – Visitor composition of *Arum cylindraceum* floral chambers in 12
 529 populations at three taxonomic levels: Arthropod classes, Insecta orders, Diptera
 530 families. Total numbers of specimens are indicated below each group.

531

532 Figure 3 – Visitor composition in the sampled populations. a) among Arthropod
 533 orders; b) among Dipteran families. Pie sizes in b are proportional to numbers of
 534 trapped insects (the scale is referred in the legend).

535

536 Figure 4 – Allometric measures of *A. cylindraceum* inflorescences. a. Length of
 537 each floral region (male, female, sterile) in function of the total inflorescence length:
 538 male (filled black circles; $T_{\text{slope}}=7.225$, slope=0.103, $p<0.001$); female (empty circles;
 539 $T_{\text{slope}}=3.695$, slope=0.334, $p<0.001$); sterile (filled grey circles; $T_{\text{slope}}=10.722$,
 540 slope=0.562, $p<0.001$). b. Number of female (empty circles, $Z=5.781$, slope=0.462,
 541 $p<0.001$) and male (filled circles, $Z=-2.425$, slope=0.225, $p<0.05$) flowers in function
 542 of total inflorescence length. c. Floral ratio in function of total inflorescence length
 543 ($T=-2.151$, slope=-0.063, $p<0.05$). d. Number of insects trapped in function of total

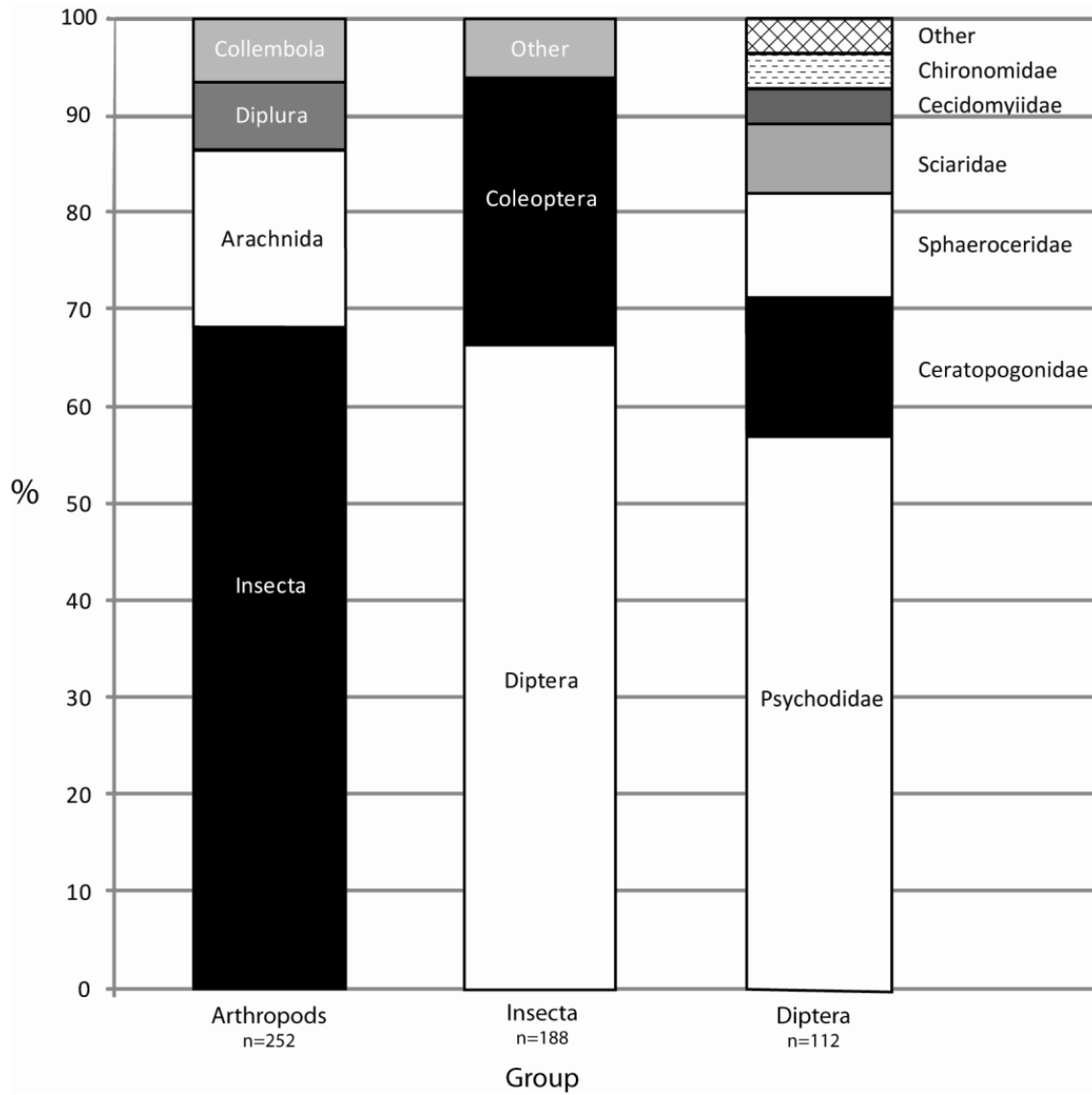
544 inflorescence length ($Z=2.819$, $p>0.05$; filled circles: inflorescences containing
545 visitors; empty circles: inflorescences not containing visitors). The total inflorescence
546 length corresponds from the base of the female zone up to the highest staminode
547 (not including the stipe and the appendix).
548

549

| | | Pollinator efficiency | |
|-----------------------------|---------------------------|--------------------------------------|---------------------------------------|
| | | low | high |
| Pollinator diversity | high (generalist) ↑ | Low probability of cross-pollination | High probability of cross-pollination |
| | low (specific) | Low probability of cross-pollination | High probability of cross-pollination |

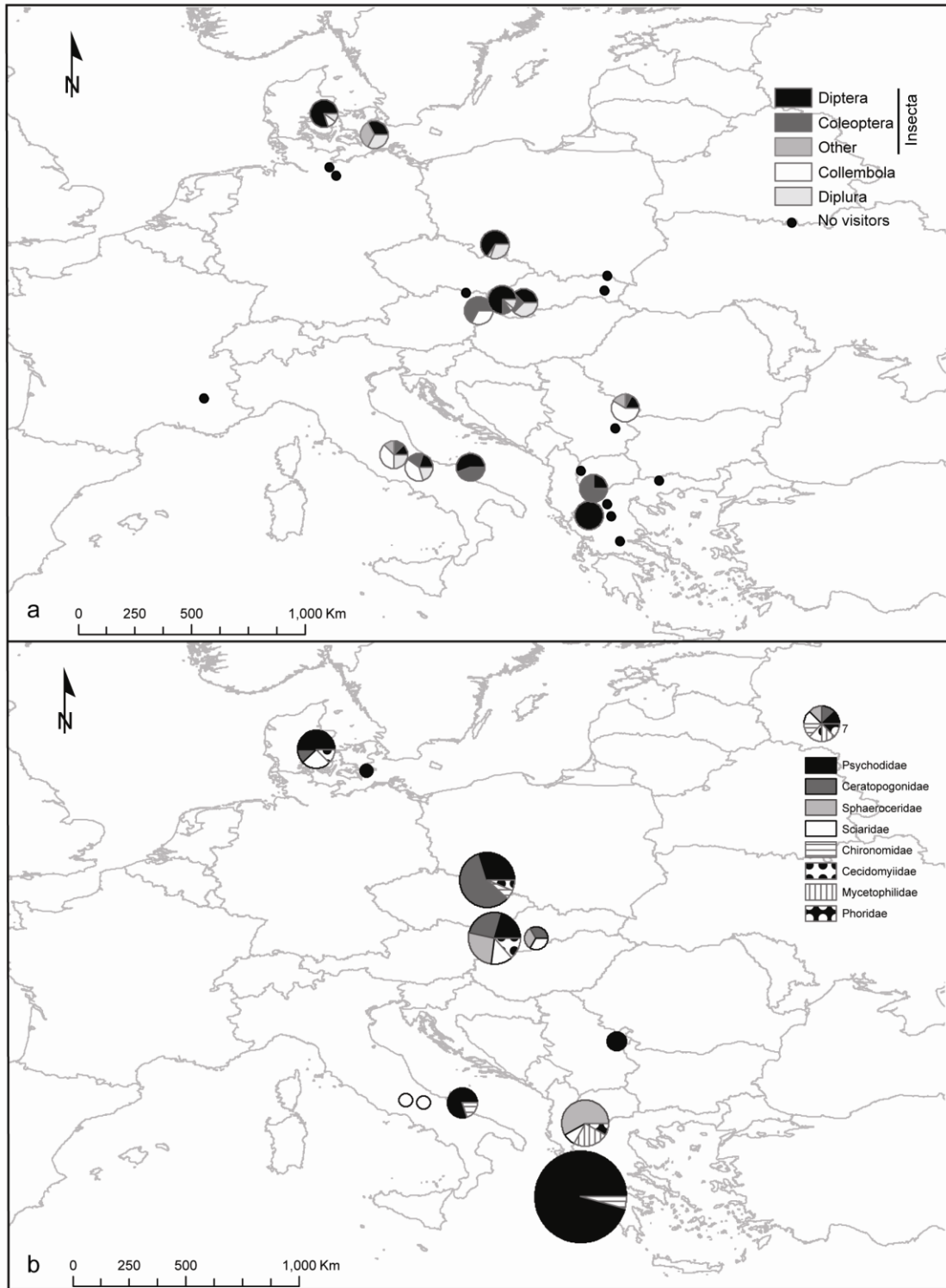
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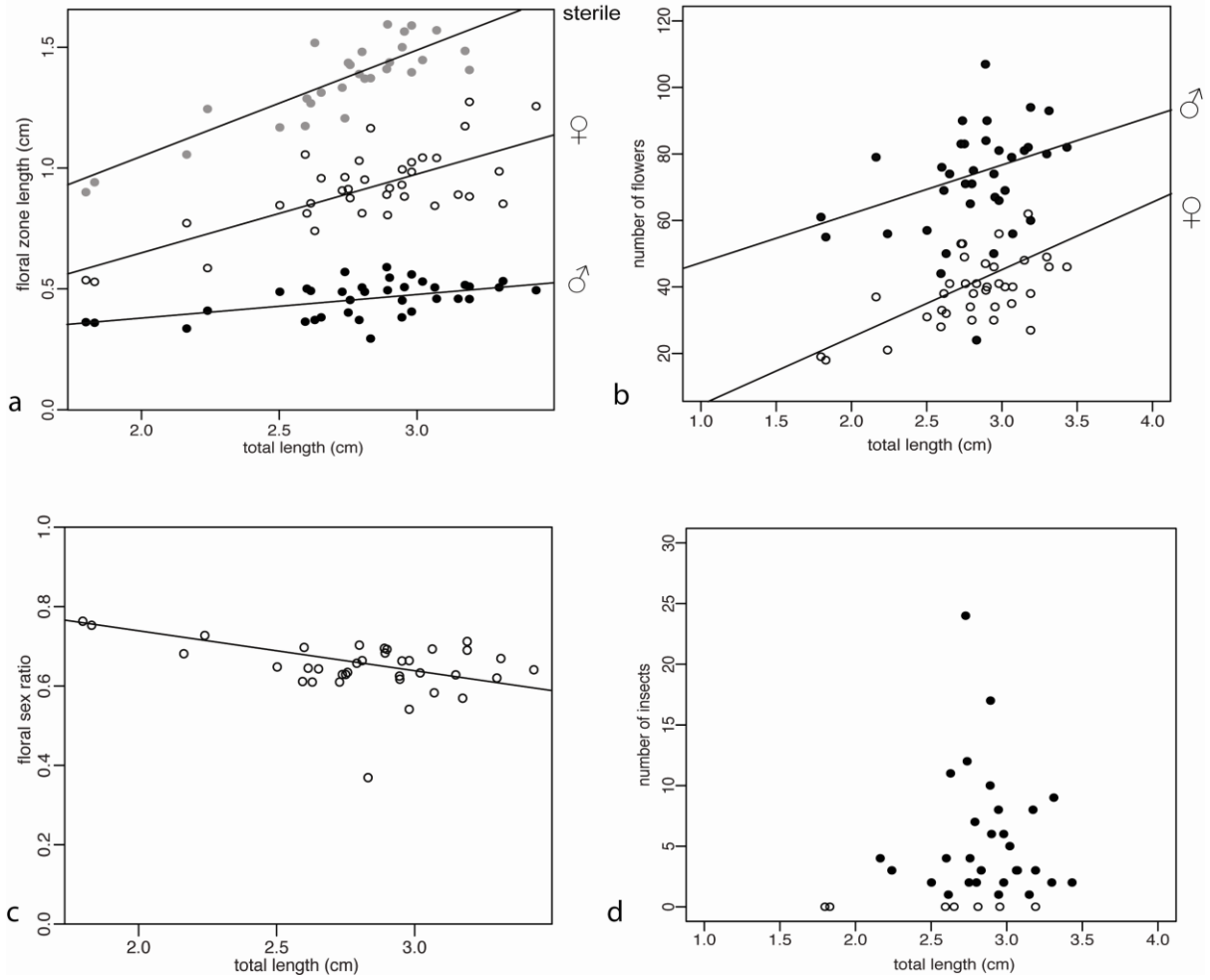
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Chapter five

*Comparative phylogeography in a specific
and obligate pollination antagonism*

Anahí Espíndola and Nadir Alvarez

1 Comparative phylogeography in a specific and obligate pollination
2 antagonism

3

4 Anahí Espíndola* and Nadir Alvarez†

5

6

7 * Laboratory of Evolutionary Entomology, Institute of Biology, University of Neuchâtel,
8 2009 Neuchâtel, Switzerland. † Department of Ecology and Evolution, University of
9 Lausanne, Biophore Building, CH-1015 Lausanne, Switzerland.

10

11 Keywords: AFLP, *Arum maculatum*, Psychodidae, spatial genetic structure, DNA
12 sequencing, plant-insect interactions

13

14 Corresponding author: Anahí Espíndola, Laboratory of Evolutionary Entomology, Institute
15 of Biology, University of Neuchâtel, 2009 Neuchâtel, Switzerland. Fax: +41 (0)32 718 30 01;
16 email :maria.espindola@unine.ch

17

18 Running title: Phylogeography of antagonistic interactions

19

20

21 **Abstract**

22 The niche of a species is defined not only by climatic and topographic features, but also
23 by biotic components. In specific and obligate pollination interactions, because of the
24 importance of the biotic factor, the presence or absence of a given ecological partner can
25 have important effects on the survival and population dynamics of associated organisms. In a
26 phylogeographic framework, we could thus expect that the fate of organisms interacting
27 specifically is also tightly interrelated. In the present study, we aim at testing such a
28 hypothesis by investigating an obligate plant-insect pollination antagonism. For this, we use
29 the European lure-and-trap system featuring *Arum maculatum* L. (Araceae) and its specific
30 Psychodid (Diptera) visitors *Psychoda phalaenoides* L. and *Psycha grisescens* Tonnoir as a
31 case-study. Based on mitochondrial data (in insects) and AFLP genome fingerprinting (in
32 plants) we first inferred the large-scale phylogeographies of each species and afterwards
33 compared their respective patterns.

34 Our findings indicate that while the plant presents a geographically well-defined genetic
35 structure, this is not the case of the insects, even though the latter harbour an important
36 amount of genetic variation. This suggests that the phylogeographic histories of the plant and
37 its pollinating insects are not congruent, a result that would indicate that insects move freely
38 during distributional shifts, while plants need either to find sites where insects are already
39 present, or change their pollinating strategy.

40

41 Introduction

42 The study of both recent (e.g., D'Andrea *et al.* 2009) and past (e.g., Nogués-Bravo *et al.*
43 2008) species distributional shifts has shown that in a changing environment organisms
44 follow the cline of their ecological niche. Environmental variations therefore drive changes in
45 distribution ranges. Because a large amount of data is available for both geological remains
46 and climatic records, we yet know that the Quaternary (2.6 – 0 million years ago -Mya) has
47 hosted series of glacial and interglacial periods, caused by orbital and tectonic events (Ehlers
48 & Gibbard 2007), which thus induced recurrent contractions and expansions of species'
49 distribution ranges (Stewart *et al.* 2010). Because such changes directly impact on
50 demographic population parameters, they can indirectly affect the genetic variation of
51 species through processes such as genetic drift that influence the allelic composition at each
52 location (Hewitt 1996). In addition, when contraction processes lead to fragmentation of a
53 species' distribution area, among-population gene flow might be restricted and local selection
54 could trigger lineages divergence (Stewart *et al.* 2010).

55 Phylogeographic studies aim at understanding these facets of the evolutionary history of
56 species, with the assumption that the spatial structure of taxa can be explained by historical
57 and geospatial (e.g., topological, hydrological) grounds (Avice 2009). In Europe for example,
58 phylogeographic patterns of organisms have been largely studied for the last 20 years, and
59 general postglacial phylogeographic paradigms have been addressed (see reviews in Hewitt
60 1999; Schmitt 2007; Taberlet *et al.* 1998). In this region, studies concurred in demonstrating
61 that during climate cooling, temperate species found refugia in the Southern mountainous
62 European peninsulas (i.e., Balkans, Italy and/or Iberia; Taberlet *et al.* 1998), and in some
63 cases also in northern areas (e.g., Carpathians; Stewart & Lister 2001). After ice retreat,
64 recolonization took place through several pathways, with contact zones appearing in several
65 central regions as for instance at natural boundaries such as mountain massifs (Taberlet *et*
66 *al.* 1998).

67 However, in a synecological context, it should be considered that species occupy
68 ecological niches that are not solely composed of abiotic factors, but which also present

69 biotic ecological components (Soberón & Peterson 2005). These latter can thus also
70 potentially shape the genetic structure of species impacting their survival, dispersal and
71 dynamics of populations. This effect could be particularly enhanced in the case of specific
72 and obligate interactions because of among-species ecological interdependence.

73 Nonetheless, only few studies (e.g., Anderson et al. 2004; Tsai & Manos *in press*) have
74 investigated large-scale phylogeographies of species interacting in specific and obligate
75 interactions. Among these, none has examined the comparative phylogeography of plants
76 and arthropods involved in specific antagonistic interactions, even though these are thought
77 to have driven diversification of several groups of angiosperms and insects in the last million
78 years (Farrell 1998; Farrell *et al.* 1992; Futuyma & Agrawal 2009). In this study, we aim at
79 investigating comparative phylogeography in a European plant-insect specific and obligate
80 antagonistic interaction, using the lure-and-trap pollination system in which the Araceae
81 *Arum maculatum* L. is specifically pollinated by the Psychodid (Diptera) sister species
82 *Psychoda phalaenoides* L. and *Psycha grisescens* Tonnoir (Diaz & Kite 2002; Espíndola *et*
83 *al. in press*)

84 In this Aroid, the volatile and heat production, and the presence of special morphological
85 floral structures (Boyce 1993) –a spadix and a spathe modified to create a closed floral
86 chamber, combined with a protogynous maturing dynamics– allows the plant to attract ready-
87 to-lay female Psychodid flies and trap them during anthesis (a maximum of one day), at the
88 end of which pollen release happens. Insects are then covered with pollen and set free;
89 some of them will be trapped a second time by another *Arum* flower, leading to cross-
90 pollination. Because a single plant experiences temporal delay in male and female flower
91 maturation, self-pollination is prevented and the plant completely relies on entomophilous
92 cross-pollination for seed production (Lack & Diaz 1991) during its short flowering period
93 (Ollerton & Diaz 1999). In the case of *Arum*, it is evident that the plant provides no rewards,
94 and insects are thus simply exploited.

95 Because i) a strong knowledge has been acquired on this European antagonistic lure-an-
96 trap specific and obligate interaction; ii) general phylogeographic patterns are currently well

97 understood at the scale of the European continent (e.g., Hewitt 1999; Taberlet *et al.* 1998);
98 iii) there is a lack in the understanding of the effect of ecological interactions on
99 phylogeographic patterns, we use here the pollination interaction between *A. maculatum* and
100 its associated Psychodids as a case-study to test hypotheses in the framework of
101 comparative phylogeography. Considering the unilateral obligate nature of the interaction
102 (i.e., the insect is ecologically independent from the plant), we hypothesize that while the
103 insect disperses freely, the plant is strongly constrained in its dispersal and long-term refugial
104 survival by the presence of its associated species; we thus expect that plants and insects
105 demonstrate incongruent phylogeographic patterns (*i.e.*, differences in the number and/or
106 distribution of refugia and recolonization pathways). The alternative hypothesis is that despite
107 this unbalanced dependence, these interacting species disperse in parallel, with the plant
108 following the insect footsteps concurring in a high level of overall congruence in
109 phylogeographic patterns (*i.e.*, similarities in the number and/or distribution of refugia and
110 recolonization pathways). We should, however, keep in mind that similarities or differences in
111 life-history traits (e.g., population sizes, generation times) of the interacting species might
112 blur these simple predictions (Alvarez *et al.* 2010).

113 In the present study, we test for the first time these hypotheses in a plant-insect
114 antagonistic and obligate interaction. Using mitochondrial markers in insects and genome
115 fingerprinting in plants, we infer the respective phylogeographies of *A. maculatum* and its two
116 main Psychodid visitors and further evaluate their congruence.

117

118 **Material and Methods**

119 **Sampling**

120 Plants and insects were sampled during springs 2006-2008, covering the whole range of
121 the plant (Boyce 1993; Espíndola *et al.* 2010). Up to 10 leaves per population were sampled
122 in 72 populations and dried in silica gel right after collection. All flower visitors were
123 preserved in separated Falcon tubes filled with 70% ethanol. Insects were identified to the
124 species following Espíndola and collaborators (*in press*) for 46 of the plant's sampling sites.

125 A maximum of three individuals per population of both *Psychoda phalaenoides* and *Psycha*
126 *grisescens* were selected for further genetic analyses (Sup. Mat. 1).

127 **Plant genetic analysis**

128 DNA was extracted using the QIAGEN DNeasy Plant extraction kit, following the
129 manufacturer's protocol (QIAGEN, Hombrechtikon, Switzerland). Alleles were produced with
130 the amplified fragment length polymorphism (AFLP) technique (Vos *et al.* 1995), assessed
131 with *EcoRI* and *MseI* digestion enzymes and further amplified with two primer pairs (E-
132 ACG/M-CTG and E-AGT/M-CAG). For digestion, 5µl of DNA were added to get a final
133 volume of 20 µl, containing 1X Buffer 2, 0.1mg/ml BSA and 1 unit/µl of *MseI* and *EcoRI* (New
134 England Biolabs); the mix was incubated at 37°C for 2h. Adaptors were ligated to the
135 digested products at 37°C for 2h, using 40µl of mix containing 1X Buffer T4, 0.45µM adaptor
136 E, 0.36µM adaptor M, 0.015 units/µl T4 ligase (Promega SA, Dübendorf, Switzerland) and
137 20µl of digestion product. Preselective PCR was done in a 20µl mix composed of 1X buffer
138 GoTaq (Promega SA, Dübendorf, Switzerland), 2µM MgCl₂, 250 µM dNTPs (Promega SA,
139 Dübendorf, Switzerland), 0.25µM primers EA and MC, 0.025 units/µl of *Taq*-polymerase
140 (Promega SA, Dübendorf, Switzerland) and 2µl ligated DNA. Conditions for this procedure
141 were 2min at 94°C, followed by 29 cycles of 45secs at 94°C, 45secs at 56°C and 1min at
142 72°C, and finishing with 10min of final extension at 72°C. The final selective step was done in
143 a 20µl mix composed of 1X buffer GoTaq (Promega SA, Dübendorf, Switzerland), 2µM
144 MgCl₂, 250 µM dNTPs (Promega SA, Dübendorf, Switzerland), 0.4µM primers EA and MC,
145 0.025 units/µl of *Taq*-polymerase (Promega SA, Dübendorf, Switzerland) and 3µl of a
146 solution 1/20 of the preselective PCR product. Conditions were 2min at 94°C, followed by 13
147 cycles of 30secs at 94°C, 30secs at 65°C with temperature lowering 0.7°C at each cycle and
148 1min at 72°C, followed by 23 cycles of 30secs at 94°C, 30secs at 65°C and 1min at 72°C,
149 and finishing with 5min of final extension at 72°C. AFLP profiles were genotyped by
150 MacroGen Inc. (South Korea). Allele binning was first automatically processed using
151 Genemapper 3.7 (Applied Biosystems) by applying a RFU threshold of 50 RFU in a range of
152 50bp to 350bp. Bins were afterwards visually checked and adjusted when necessary.

153 Because digestion and amplification problems can arise during the procedure and can highly
154 bias results (Arrigo *et al.* 2009), we randomly chose 36 individuals per plate to be used as
155 intraplate replicates, while five samples were selected out of the total to be used as interplate
156 replicates. The percentage of bands shared by all the replicates of the same sample was
157 analyzed and repeatability was assessed.

158 In order to identify the genetic structure of *A. maculatum*, 20 runs of 1.000.000
159 generations with a 200.000 burnin period for each *K* prior number of populations (set
160 between 1 and 20) were performed using a model-based assignment algorithm as
161 implemented in Structure 2.2 (Falush *et al.* 2007). Following recommendations of the
162 authors, data was coded as diploid and recessive alleles as present. The most probable *K*
163 was identified using approaches proposed both by Pritchard *et al.* (2000) and Evanno *et al.*
164 (2005). A non model-based algorithm was also applied to infer clustering of specimens, by
165 using the K-means method, following Hartigan and Wong (1979), and selecting for the best
166 number of groups using the inertia criterion (*i.e.*, the average of the distances between the
167 centroid of each cluster and each sample contained in it), following Kergoat & Alvarez
168 (2008). This method, recently applied in a phylogeographic framework by Burnier *et al.*
169 (2009) and Arrigo *et al.* (2010) is based on the allele composition of each sample and allows
170 the identification of the number of genetic clusters that optimises the grouping of samples.
171 The analysis was performed on R 2.9.1 (R Core Development Group, 2009), applying 10.000
172 replicates for each *K* cluster, for values going from 1 to 20. Scripts are available upon
173 request to the first author.

174 **Insect genetic analysis**

175 DNA from 152 flies belonging to the two visitor species *P. phalaenoides* and *P. grisescens*
176 was extracted using the QIAGEN DNeasy Blood and Tissue extraction kit (QIAGEN,
177 Hombrechtikon, Switzerland). In order to perform phylogeographic analyses, three (Cytb,
178 COI, 16s) and one (Cytb) mitochondrial regions were amplified for *P. phalaenoides* and *P.*
179 *grisescens*, respectively. PCR were done in 20µl of a mix composed of 0.5X buffer, between
180 1.25 and 2.5 mM MgCl₂, 10mM dNTP, 1 unit of GoTaq DNA polymerase (Promega,

181 Dübendorf, Switzerland), 0.5 μ M primers (Table 1) and 3 μ l DNA and run in a TGradient
182 thermocycler (Biometra, Goettingen, Germany). The program consisted of 2:30 min at 95°C,
183 followed by 35 cycles of 30 sec at 95°C, 40 sec at 57°C or 48.5°C (Table 1), 1 min at 72°C,
184 and finishing with a final elongation of 8 min at 72°C. Amplified fragments were sequenced
185 by Macrogen Inc. (South Korea) and Fasteris SA (Switzerland). Sequences were visually
186 corrected on Chromas Pro 1.41 (Technelysium Pty. Ltd.) and further aligned on BioEdit
187 7.0.4.1 (Hall 1999). Gaps were coded using the method of Simmons and Ochoterena (2000)
188 as implemented in FastGap 1.2 (Borchsenius 2009).

189 Phylogenetic relationships were inferred using Bayesian and Maximum Likelihood (ML)
190 approaches. Models of evolution were estimated using MrAIC v. 1.4.4 (Nylander, 2004) and
191 Bayesian analyses were performed on MrBayes 3.1.2 (Huelsenbeck & Ronquist 2001;
192 Ronquist & Huelsenbeck 2003) running 50.000.000 generations, with temperature 0.5 and
193 sampling every 1.000 generations. ML analyses were run on RaxML 7.2.6 (Stamatakis 2006)
194 based on 10.000 bootstraps. Complementary phylogenetic networks were constructed using
195 TCS v1.21 (Clement *et al.* 2000) applying a 95% connection limit.

196 In order to test for isolation by distance, correlations between genetic and geographic
197 distances were calculated for both species. For this, considering the models of evolution
198 previously inferred for each partition and each species, we calculated the pairwise genetic
199 distance between samples using the *Dset* function as implemented in PAUP* 4.0 (Swofford
200 2003). To account for multiple partitions in *P. phalaenoides*, distances were calculated for
201 each partition and were afterwards averaged. Geographic distances were calculated
202 between all samples and correlations were adjusted between both matrices using GenAlEx
203 6.3 (Peakall & Smouse 2006).

204 **Comparative analysis**

205 Phylogeographic patterns for both the plant and the insects were displayed on
206 geographical maps using ArcMap 9.3 (ESRI, Redlands CA, USA), with populations being
207 represented as pie charts showing the number of individuals assigned to each cluster or
208 haplotype.

209 Congruence was visually evaluated and was accepted if the spatial distribution and the
210 number of genetic clusters were similar for both the plants and the insects.

211

212 **Results**

213 **Phylogeography of *A. maculatum***

214 A total of 394 individuals were extracted, out of which 362 successfully amplified bands .
215 Of these, 62 did not present any band with primer pair E-ACG/M-CTG, while 4 did not
216 amplify with primer pair E-AGT/M-CAG. Finally, primer pair E-ACG/M-CTG provided 166
217 markers, while E-AGT/M-CAG provided 160. Reproducibilities were 0.963 and 0.957 for E-
218 ACG/M-CTG and E-AGT/M-CAG, respectively, calculated according to Bonin *et al.* (2004).

219 Structure runs showed that the most likely number of genetic clusters *K* for the *A.*
220 *maculatum* dataset was two. Genetic groups were geographically structured (Fig. 1a) with
221 one group spreading through the Balkans and Italy and the other covering the rest of Europe.

222 The non-hierarchical K-means approach indicated that K-values providing good inertia
223 values were three and six (Sup. Mat. 2). In the first case ($K=3$; Fig. 1b), one of the three
224 clusters appeared geographically structured: while two admixed groups were distributed
225 throughout the central and northern European region, the third one was restricted to the
226 Balkans and Italy. In the latter case ($K=6$; Fig. 1c) a similar pattern of admixture among four
227 clusters was retrieved for the central and northern European area, while in addition to the
228 group covering the Balkans and Italy, one supplementary cluster was exclusive of the
229 Carpathian region (pink in Fig. 1c).

230 **Phylogeography of *P. phalaenoides* and *P. griseocens***

231 A total of 104 samples of *P. phalaenoides* successfully amplified three regions (1428 bp),
232 while 45 *P. griseocens* amplified one region (679 bp) (Table 2; sequences will be submitted
233 to GenBank during the review process).

234 Both phylogenetic topologies (Sup. Mat. 3) and haplotype networks (Fig. 2) indicated that
235 no geographic structure of the genetic variation was retrieved in either of the two species

236 analyzed. For convenience of representation, we illustrate this result by focusing on the
237 haplotype network approach.

238 In *P. phalaenoides* (Fig. 2a), 24 different haplotypes were recovered, 21 of them being
239 unique to single specimens (white in Fig. 2a). The three others (pink, blue and green, in Fig.
240 2a) were present in 80, three and two specimens, respectively. While the most frequent
241 haplotype (pink) was separated of the others by at least four mutational steps, the remaining
242 types were more interrelated. Even if unique haplotypes appeared to be relatively more
243 abundant in the Eastern edge of the sampled zone, two main haplotypes (pink and blue)
244 were present homogeneously across the sampling range and the remaining haplotype
245 (green) was present only at the Eastern edge of the distribution, but included only two
246 samples.

247 In *P. grisescens* (Fig. 2b), 22 different haplotypes were retrieved, 19 being unique to
248 single specimens (in white on the Fig. 2b). The three others (pink, blue and green) were
249 present in 13, nine and three specimens, respectively. All 22 haplotypes were highly
250 interrelated with 20 of them showing distances to the closest haplotype equal to one step,
251 and only two demonstrating distances equal to two steps. Here again, the spatial distribution
252 of the main haplotypes was not structured.

253 When testing isolation by distance (Fig. 3), results indicated that in both species there was
254 no correlation between genetic and geographic distances ($R^2=0.003$ and 0.006 for *P.*
255 *phalaenoides* and *P. grisescens*, respectively, with *P* values > 0.05 in both cases).

256 **Comparison of phylogeographic patterns**

257 The phylogeographic patterns of *Arum* and each of the two Psychodids presented no
258 congruence: neither the number nor the distribution of genetic groups were similar between
259 the interacting species. Whereas the plant showed a trend towards a spatial structure of its
260 genetic variation, insects' haplotypes were distributed irrespective of any geographic
261 organization.

262

263 **Discussion**

264 **Phylogeography of *A. maculatum***

265 Our results demonstrate that *A. maculatum* shows a trend towards a spatial structure of
266 the genetic variation (Fig. 1), with one [*Structure* and K-means (K=3) approaches] or two [K-
267 means (K=6) approach] clusters exclusive of the Balkan and Italian region and of the
268 Carpathians, and the others widespread throughout Europe (Fig. 1a-c). This pattern is similar
269 to those unravelled in other European organisms, as for instance, *Ursus arctos* or *Sorex*
270 *araneus* (Hewitt 1999), where one widespread European genotype was segregated from two
271 or one clusters endemic of south-eastern Europe. The observed split of the south-eastern
272 region into two groups (i.e., Italy and Balkans on one hand, and Carpathians on the other
273 hand) at higher values of K (K=6) is also congruent with climatologic records, which indicate
274 that during the Last Glacial Maximum (LGM; around 18Kya) the Northern Adriatic was dry
275 land because of lowered sea levels and a communication between its Eastern and Westerns
276 coasts was possible (Mussi 1990), whereas gene flow with the Carpathian area was probably
277 more restricted.

278 It is worth noting that a large part of genetic variation identified by the non-model based
279 approach seems to be little or not spatially structured. This is the case of two and four
280 clusters in the K=3 and K=6 analyses, respectively. However, because the model-based
281 approach (i.e., *Structure*) did not identify these groups, it is difficult to conclude whether the
282 admixed pattern at the central and northern part of the European distribution area is
283 artefactual or not. Splitting overestimation by the K-means approach could result from a main
284 principle of the method, which aims at minimizing among-sample distances. As a
285 consequence, the more clusters it creates, the smaller the distances between the centroid of
286 each cluster and each included sample are, a bias similar to overfitted correlations. Following
287 this hypothesis, the multiple central-European clusters identified would be more likely an
288 indication of the difficulty to define a best *K* than a biological reality.

289 Nonetheless, if we assume that this admixture result reflects a biological characteristic, we
290 could imagine that specimens from this region are carrying a large amount of genetic
291 diversity present in pre-LGM populations of *A. maculatum*. In this scenario, plants from the

292 different genetic clusters identified in the K-means approach would have, at some time,
293 shared Pleistocene refugia, and among-lineage cross-reproduction might have occurred. The
294 pattern currently observed would thus be a consequence of ancient polymorphism combined
295 with massive gene flow among lineages. After the last glacial retraction (around 13 Kya),
296 immigrants from these refugia would have reinvaded the region, carrying in their genomes
297 both the signature of ancient polymorphisms, but also the effect of more recent refugial
298 genetic admixture. A result that supports this view is the pattern observed in the distribution
299 of Nei diversities (Sup. Mat. 4), which indicates centres of high diversity in both Italian and
300 south-eastern areas and in regions where several of these central-European clusters are
301 shown to be admixed.

302 **Insect Phylogeography**

303 Despite that the markers used revealed a high level of genetic variability, with more than
304 20 haplotypes identified in both cases (see Table 2 and Fig. 2), no geographic structure
305 could be retrieved in either of the species. Considering the large among-population distances
306 (up to 3.800 km), this pattern could indicate that gene flow between far distant populations is
307 possible in Psychodids despite the flies' small sizes, which is in agreement with the
308 significant lack of isolation by distance (Fig. 3). Moreover, because of their high population
309 sizes and number of generations per year, it is possible that substitution rates are extremely
310 high (see below). Although it is empirically difficult to quantify dispersal by a direct approach
311 in these species, *P. phalaenoides* was recently sampled in aerial netting surveys in England,
312 where insects were collected at around 200m above ground, a height at which any insect
313 found is considered to be engaged in migratory –and thus potentially large-scale– movement
314 (Chapman *et al.* 2004). Moreover, in at least one previous study (Hardy & Cheng 1986),
315 Psychodids were shown to be able to perform extended flights of at least 160km, since
316 members of this family were collected alive from boats navigating at this distance offshore in
317 the North Sea. Such biological evidences are highly compatible with the absence of spatial
318 genetic structure observed in our results, in which we indirectly reached the same
319 conclusion. It is also interesting to point out that high level of gene flow –with its associated

320 lack of spatial genetic structure- has been also observed in other insect species, as
321 demonstrated in the case of *Ips typographus* (Coleoptera: Scolytidae) at a regional scale in
322 Switzerland (Gugerli *et al.* 2008).

323 Another explanation for this lack of phylogeographic structure in combination with high
324 dispersal rates could also lay in the biology of these species, which could have been
325 associated to human activities since the Holocene. It is known that these insects are tightly
326 related to agricultural and anthropized environments, since they develop in rich substrates as
327 cow dung, sewage works and decaying matter (Satchell 1947; Vaillant 1971). Because of
328 this characteristic, it is possible that a part of the mixture of genetic types could be due to
329 both old and recent human activities, as rapid (fuelled) long distance movements currently
330 used for the transport of cattle and waste. Such an effect of human-mediated dispersal on
331 the spatial genetic pattern of insect species has already been observed in other taxa
332 associated with human activities, such as the bean pest beetle *Acanthoscelides obvelatus*
333 (Alvarez *et al.* 2007).

334 **Comparative Phylogeography**

335 The respective presence and absence of genetic structure in plants and insects indicate
336 that there is no congruence between both genetic structures. Not finding any evidence of
337 shared spatial genetic structure would confirm our first hypothesis: both species appear to
338 present independent refugial survival and recolonization pathways. The antagonistic
339 interaction seems thus to have persisted through Pleistocene climatic oscillations even if
340 lineages of the independent species (*i.e.*, the insect) do not systematically find shelter in the
341 same refugia as the dependent species (*i.e.*, the plant). However, the maintenance of the
342 interaction, and of the dependent plant species if it harbours a low level of plasticity, requires
343 that at least one refugium is shared by the two species. Similarly, the interaction can persist
344 despite differences in recolonization routes, as far as the independent species is frequent
345 enough to allow the dependent species to find both suitable abiotic and biotic habitats.

346 It should be noted that a recent post-glacial recolonization simulation-based study
347 performed by Borer *et al.* (submitted) proposed that a pattern similar to the one obtained in

348 the present study (*i.e.*, presence and absence of genetic structure in one and the other
349 partner, respectively) could be retrieved when the two interacting species disperse at very
350 different rates. This could be the case in the interaction studied here as well, since it is
351 known that Psychodid flies can reach enormous population sizes [several thousand
352 individuals per square meter (Arshad & Moh Leng 1991)] with extremely short generation
353 times when compared to those of *A. maculatum* [12-18 generations per year in the flies
354 (Vaillant 1971) vs. one generation every 10-15 years in the plant (Boyce 1993)]. This can
355 allow for higher dispersal rates through time, increase the mutation rate and enhance the
356 probability of genotype admixture.

357 Despite this incongruence in spatial genetic structures, recent large-scale studies on the
358 biogeography of pollinators associated with *A. maculatum* (Espíndola *et al.* in press)
359 indicated that respective densities of the two Psychodid flies were geographically structured
360 (Fig. 1d), roughly matching the plant spatial genetic structure identified in the current study
361 (Fig. 1a-c). The relative abundance of *P. griseescens* as a visitor of *A. maculatum* is much
362 higher in south-eastern Europe, an area in which the plant is characterized by a marked
363 spatial genetic structure. If this correlation is not random, this would indicate that the spatial
364 genetic structure of the plant could be currently shaped by the spatial variation of the
365 biological characteristics of pollinators in the different regions, the main principle of the theory
366 of geographic mosaic of coevolution (Thompson 2005). Another option could be that the
367 current genetic structure of the plant reflects selective and adaptive processes related to
368 pollinator attractions having happened during the last glacial period. In the future, coalescent-
369 based hypothesis testing on AFLP data could provide a statistical answer to the question of
370 whether or not the phylogeographic structure found in the plant is defined by history (*i.e.*, in
371 relation to its hindcasted LGM distribution) or pollination ecology (*i.e.*, defined by the
372 Psychodid composition).

373

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380

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499
500

501 **Author information box**

502 Anahí Espíndola is a PhD student at the University of Neuchâtel (Switzerland), interested
503 in ecological, spatial and phylogeographic aspects of the evolution, establishment and
504 dynamics of plant-insect interactions. Nadir Alvarez is a research associate at the University
505 of Lausanne (Switzerland), interested in the evolution of plant-insect interactions at different
506 scales of time and space.

507 Table 1 – Primer sequences and respective annealing temperatures used to sequence
 508 portions of the insects' mtDNA.

509

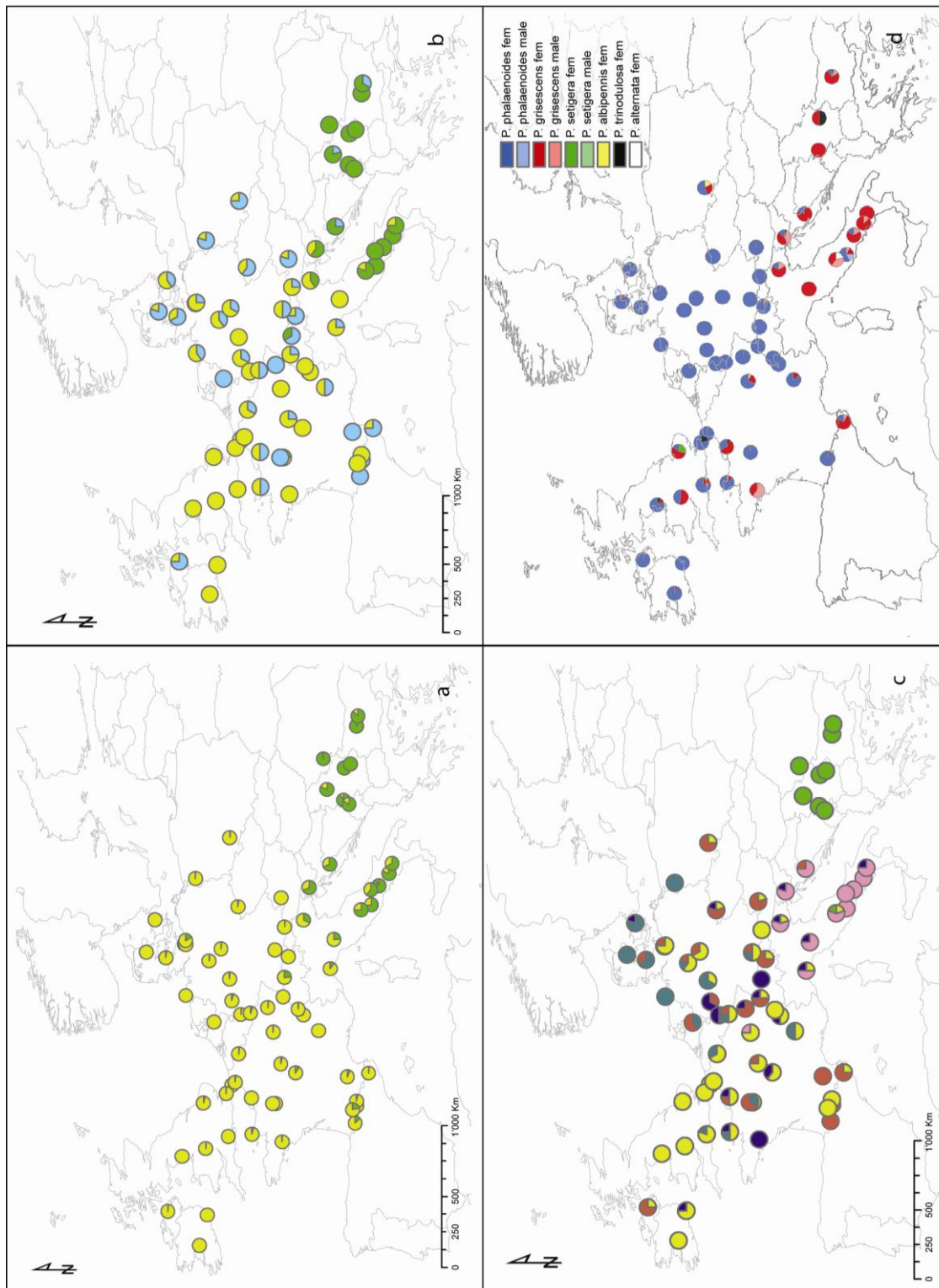
| Region | Primers | Sequence (5' -> 3') | Annealing (°C) |
|--------|------------|------------------------------------|----------------|
| COI | C1-J-1718 | GGG GGG TTT GGA AAT TGA TTA GTG CC | 48.5 |
| | TI-2-N3014 | TCC ATT GCA CTA ATC TGC CAT ATT A | 48.5 |
| Cytb | CB-J-11338 | CAC ATT CAA CCA GAA TGA TAT TT | 57 |
| | N1-N-12051 | GAT TTT GCT GAA GGT GAA TCA GA | 57 |
| 16s | LR-N-13398 | CGC CTG TTT AAC AAA AAC AT | 57 |
| | LR-J-12883 | CCG GTT TGA ACT CAG ATC ATG T | 57 |

510

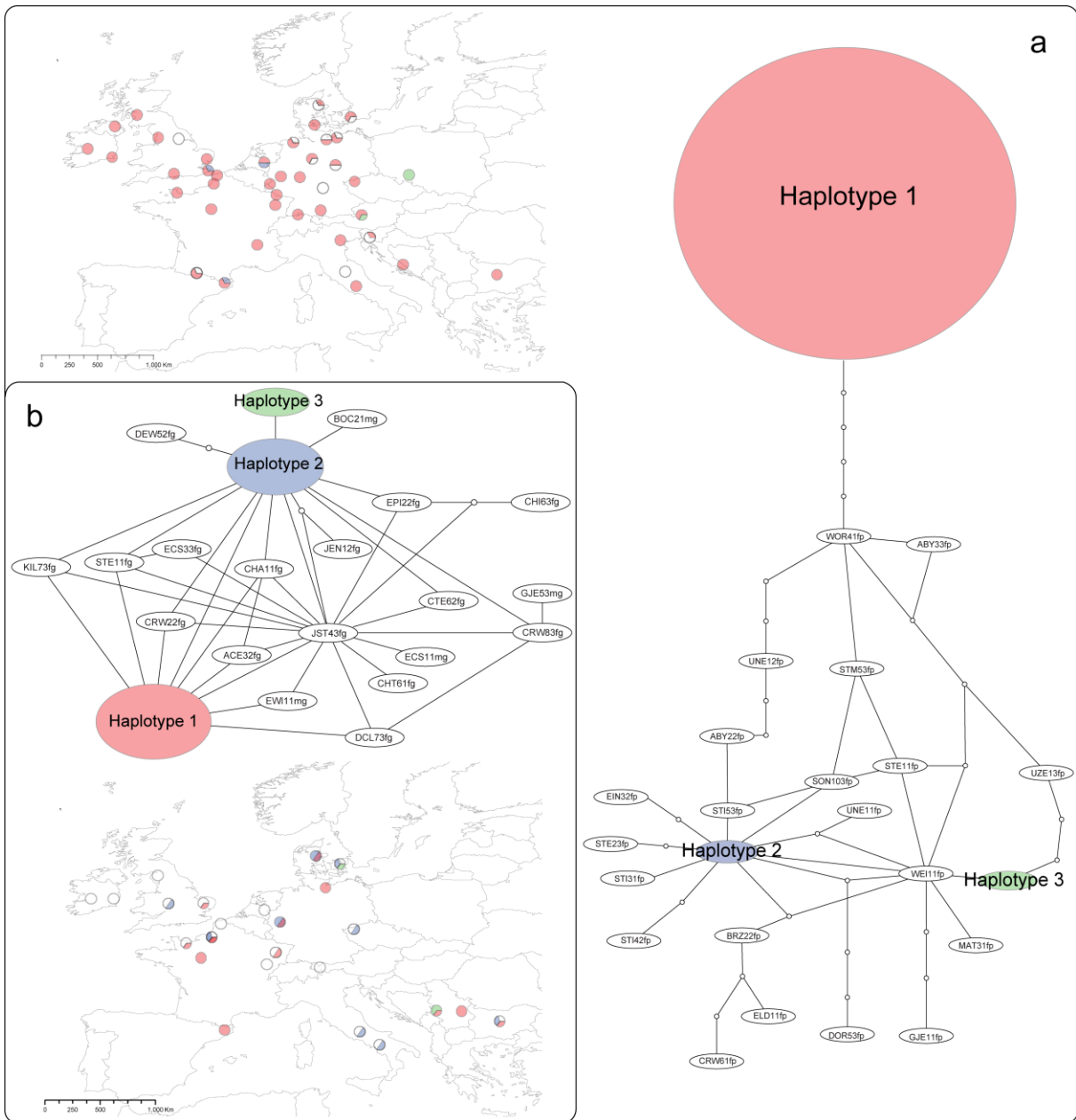
511 Table 2 – Total, variable (V), parsimony-informative (PI) and constant (C) sites per
 512 species and sequenced region, in base pairs, for the three mtDNA regions sequenced in
 513 insects.

| Species | Region | C | V | PI | Total |
|------------------------|--------------|-------------|-----------|-----------|-------------|
| <i>P. phalaenoides</i> | 16s | 209 | 14 | 0 | 223 |
| | Cytb | 646 | 34 | 14 | 680 |
| | COI | 509 | 16 | 5 | 525 |
| | Total | 1364 | 64 | 19 | 1428 |
| <i>P. grisescens</i> | Cytb | 611 | 68 | 13 | 679 |

514



515
 516 Figure 1 – a-c) Genetic clusters inferred under Bayesian (a) and non model-based (b and c)
 517 frameworks. Colours indicate clusters; portions in pies indicate probabilities of assignment
 518 (for Bayesian) or number of individuals (for K-means) assigned to each cluster. d)
 519 Composition of Psychodid visitors –and putative pollinators– of *A. maculatum* (from
 520 Espíndola *et al. in press*; reproduced with permission of the editors). Refer to figure legend
 521 for identity of species.



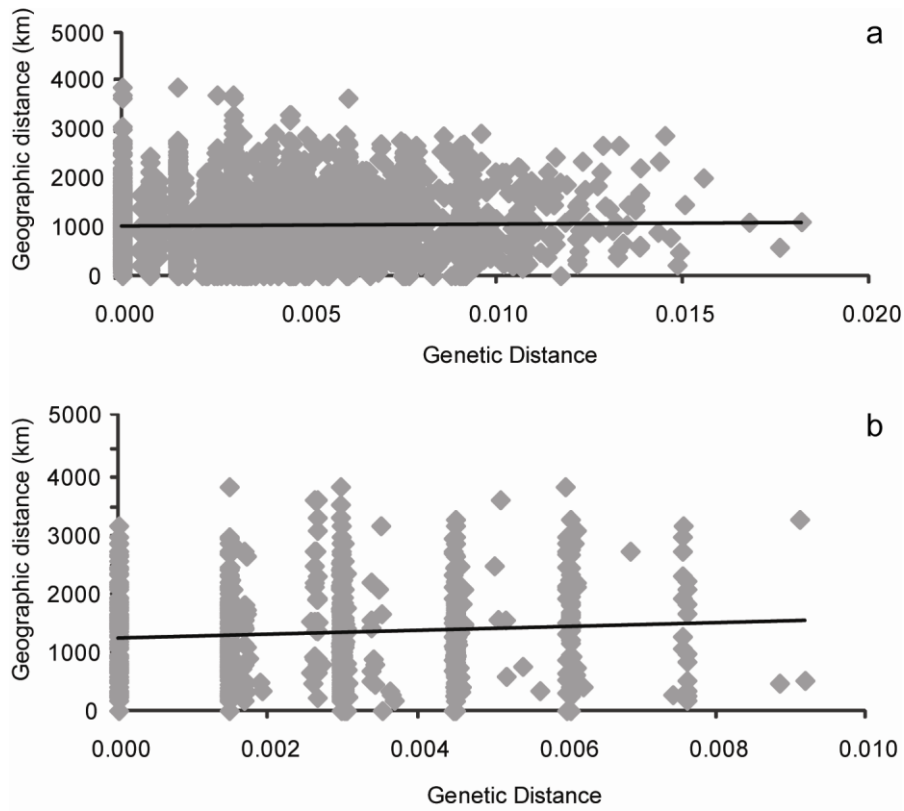
522

523

524

525

Figure 2 – Haplotype networks and respective geographic distribution of most-frequent haplotypes obtained for *Psychoda phalaenoides* (a) and *Psycha grisescens* (b).



526

527 Figure 3 – Correlations between genetic and geographic distances for *P. phalaenoides* (a)528 and *P. grisescens* (b). Adjusted regression lines are shown ($R^2=0.003$ and 0.006 , P -values:529 0.350 and 0.170 , respectively).

530

531 Supplementary Material 1 – List of sampled locations with geographic coordinates.

532 Number of plants, *Psychoda phalaenoides* and *Psycha grisescens* collected at each location.

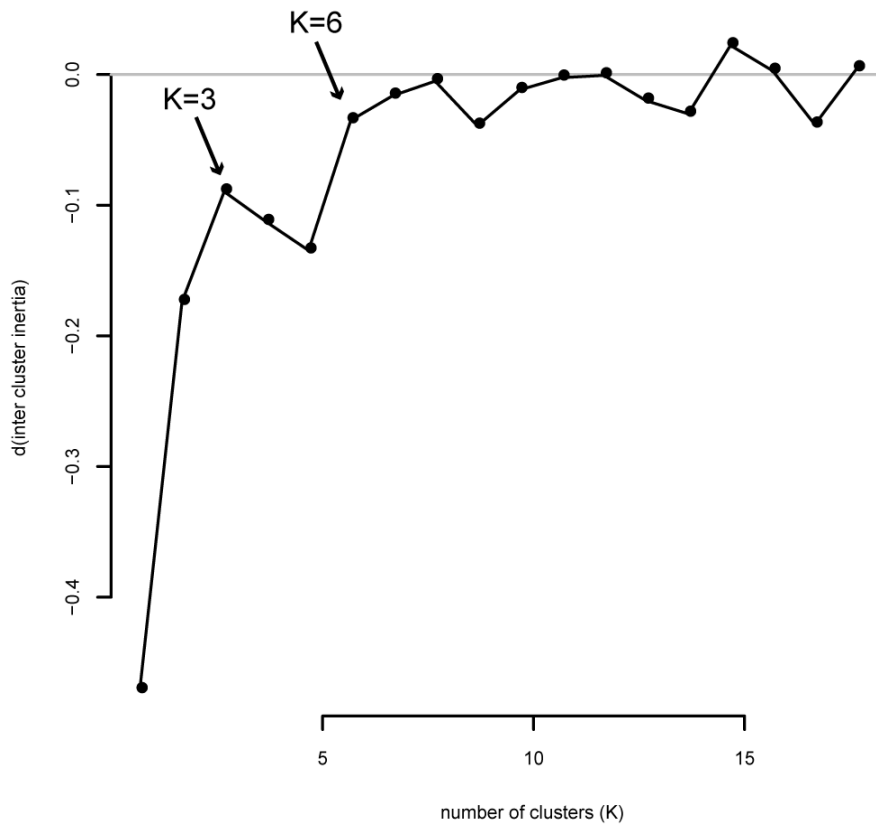
| Location | Code | Longitude | Latitude | Altitude (m) | <i>A. maculatum</i> | <i>P. grisescens</i> | <i>P. phalaenoides</i> |
|------------------|------|-----------|-----------|--------------|---------------------|----------------------|------------------------|
| Aby | ABY | 10.16801 | 56.14328 | 0 | 5 | 2 | 3 |
| Acerno | ACE | 15.16854 | 40.77018 | 928 | 5 | 2 | |
| Avala | AVA | 20.51481 | 44.69189 | 468 | 5 | | |
| Bausen | BAU | 0.7239 | 42.82485 | 610 | 5 | | |
| Baile Herkulane | BHK | 22.4586 | 44.93176 | 300 | 5 | | |
| Blankenheim | BLA | 7.098333 | 50.73389 | 531 | 5 | 2 | 3 |
| Boc | BOC | 13.06557 | 50.34522 | 364 | 5 | 2 | 2 |
| Bouloire | BOU | 0.56135 | 47.97096 | 115 | 5 | 1 | 1 |
| Brzeg | BRZ | 17.4509 | 50.85675 | 336 | 5 | | 2 |
| Bois St. Pierre | BSP | 3.71207 | 50.29306 | 130 | 5 | | |
| Chaumont | CHA | 5.09475 | 48.11508 | 296 | 5 | 1 | 3 |
| Chiflik | CHI | 24.52836 | 42.813 | 786 | 5 | 3 | 1 |
| Churchtown | CHT | -2.78686 | 53.88121 | 3 | 5 | 1 | 3 |
| Cranwich | CRW | 0.60818 | 52.52699 | 24 | 5 | 3 | 1 |
| Conteville | CTE | 1.73872 | 50.73731 | 60 | 5 | 1 | 3 |
| Duclair | DCL | 0.92016 | 49.48517 | -1 | 5 | 3 | 2 |
| Dewley | DEW | -2.2918 | 52.38652 | 35 | 5 | 2 | 2 |
| Dornum | DOR | 7.43117 | 53.65095 | 0 | 5 | | 3 |
| Ecausseville | ECS | -1.37439 | 49.46218 | 0 | 5 | 3 | 2 |
| Einhaus | EIN | 10.73472 | 53.71355 | 21 | 5 | 1 | 3 |
| Eldgasen | ELD | 9.62534 | 52.17533 | 101 | 5 | | 2 |
| Epinal | EPI | 6.654722 | 48.45028 | 400 | 5 | 2 | 3 |
| Ewijk | EWI | 5.74856 | 51.8763 | 21 | 5 | 1 | 3 |
| Falkstone | FLK | 1.19223 | 51.09506 | 78 | 5 | | 3 |
| Font de la Salut | FON | 2.49728 | 42.06183 | 1100 | 5 | 3 | 3 |
| Foret d'orient | FOR | 0.56135 | 48.1518 | 295 | 5 | | |
| gabrovnitza | GAB | 25.17856 | 42.7295 | 731 | 4 | | |
| Gross Disnack | GDI | 10.70769 | 53.72313 | 42 | 13 | | |
| Chene-Bourg | GE | 6.18533 | 46.192782 | NA | 2 | | |
| Genos | GEN | 0.40551 | 42.81207 | 975 | 5 | | |
| Gjeddesdal | GJE | 12.23064 | 55.60755 | 32 | 5 | 3 | 3 |
| Gostilje | GOS | 19.83549 | 43.65561 | 785 | 5 | 3 | |
| La Groutte | GRO | 2.51343 | 46.69078 | 137 | 5 | | |
| Jenne | JEN | 13.18219 | 41.88142 | 824 | 6 | 2 | 2 |
| Johnstown Castle | JST | -6.50412 | 52.29245 | -18 | 5 | 1 | 3 |
| Kamena Gora | KAG | 19.56567 | 43.2983 | 1201 | 5 | | |
| Killaloe | KIL | -8.44438 | 52.7946 | 44 | 5 | 1 | 2 |
| Laccio | LAC | 9.13009 | 44.48817 | 629 | 4 | | |
| Lescun | LEC | -0.6709 | 42.91148 | 1000 | 5 | | |

| | | | | | | | |
|----------------------|-----|-----------|----------|------|----|---|---|
| Les | LES | 0.7237 | 42.82462 | 610 | 5 | | |
| Balzers | LIE | 9.886944 | 47.15111 | 494 | 4 | | 3 |
| Hohenweiler | LIN | 10.315556 | 48.00694 | 486 | 5 | | 3 |
| La Loubatiere | LOU | 2.2568 | 43.40768 | 720 | 4 | | |
| Tintesmühle | LUX | 6.223889 | 50.15611 | 305 | 4 | | 3 |
| Maiche | MAI | 7.339722 | 47.485 | 613 | 4 | | |
| Lago Matese | MAT | 14.40561 | 41.40639 | 1098 | 10 | | 1 |
| Montese | MON | 10.98371 | 44.25523 | 707 | 5 | | |
| Morro Reatino | MOR | 12.85742 | 42.53585 | 759 | 6 | | |
| Monte Pizi | MPI | 14.15127 | 41.92002 | 925 | 3 | | |
| La Mignonnais | MSG | -1.86466 | 47.55066 | 16 | 3 | | |
| Quilen | QUI | 1.91746 | 50.52071 | 112 | 5 | | |
| Randalstown | RDT | -6.28349 | 54.79447 | 58 | 5 | | 3 |
| Rifreddo | RIF | 15.82473 | 40.57235 | 1172 | 5 | | |
| Col de Roanza | ROA | 12.20989 | 46.17638 | 816 | 5 | | 1 |
| Romsey | ROM | -1.52498 | 50.96424 | 52 | 5 | | 2 |
| St. Fargeau | SFG | 3.09205 | 47.62233 | 226 | 5 | | |
| Sitno | SIT | 14.84701 | 53.03406 | 66 | 5 | | 3 |
| Schmilau | SMI | 10.7575 | 53.66833 | 2 | 5 | | |
| Sokobanja Vrh source | SOK | 21.88755 | 43.60373 | 844 | 10 | 1 | |
| Sonderborg | SON | 9.82699 | 54.91554 | 10 | 7 | | 2 |
| Steeg | STE | 13.63328 | 47.61349 | 513 | 5 | 1 | 3 |
| Sankt Margrethen | STM | 11.77574 | 47.3825 | 527 | 5 | | 3 |
| Suva Planina | SUV | 22.13166 | 43.21021 | 1114 | 4 | | |
| Udbina | UDB | 15.76134 | 44.53128 | 810 | 6 | | 1 |
| Unec | UNE | 14.28609 | 45.82845 | 525 | 5 | | 3 |
| Ecublens | VD | 6.56637 | 46.52655 | NA | 2 | | |
| Verne | VER | 5.1905 | 45.22675 | 502 | 5 | | 1 |
| Hautes Pyrenées | VZE | 0.17868 | 43.07732 | 580 | 5 | | 3 |
| Weilburg | WEI | 8.475556 | 50.88361 | 158 | 3 | | 3 |
| Worbis | WOR | 10.38198 | 51.42088 | 370 | 4 | | 2 |
| Yutz (Thionville) | YUT | 6.303889 | 49.55139 | 225 | 4 | | 3 |
| Hönggerberg | ZH | 8.56637 | 47.40812 | NA | 5 | | 3 |

533

534

535 Supplementary Material 2. - Inertia computed, following Kergoat & Alvarez (2008), for
536 each K value in the K-means computations on the plant AFLP dataset.

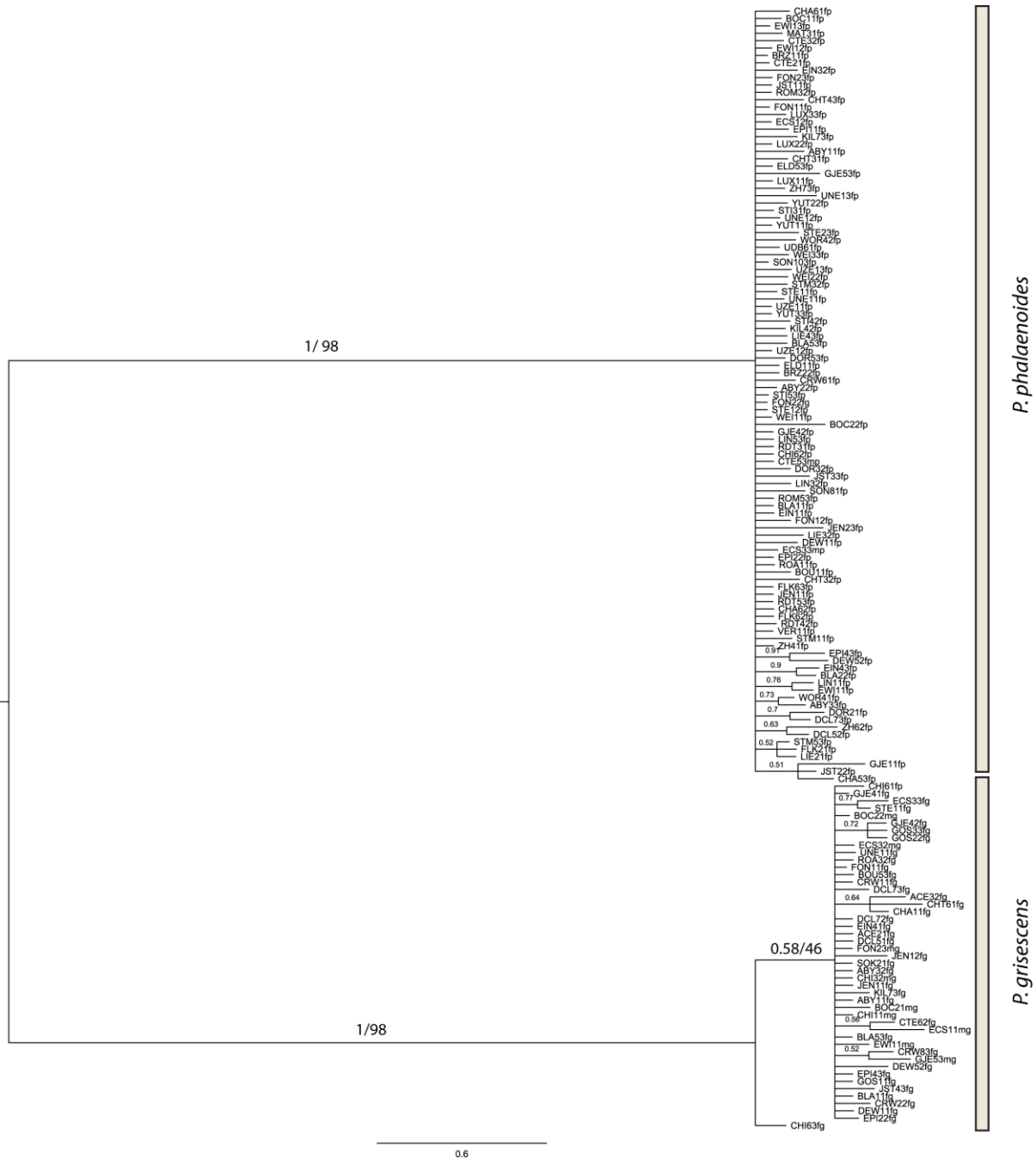


537

538

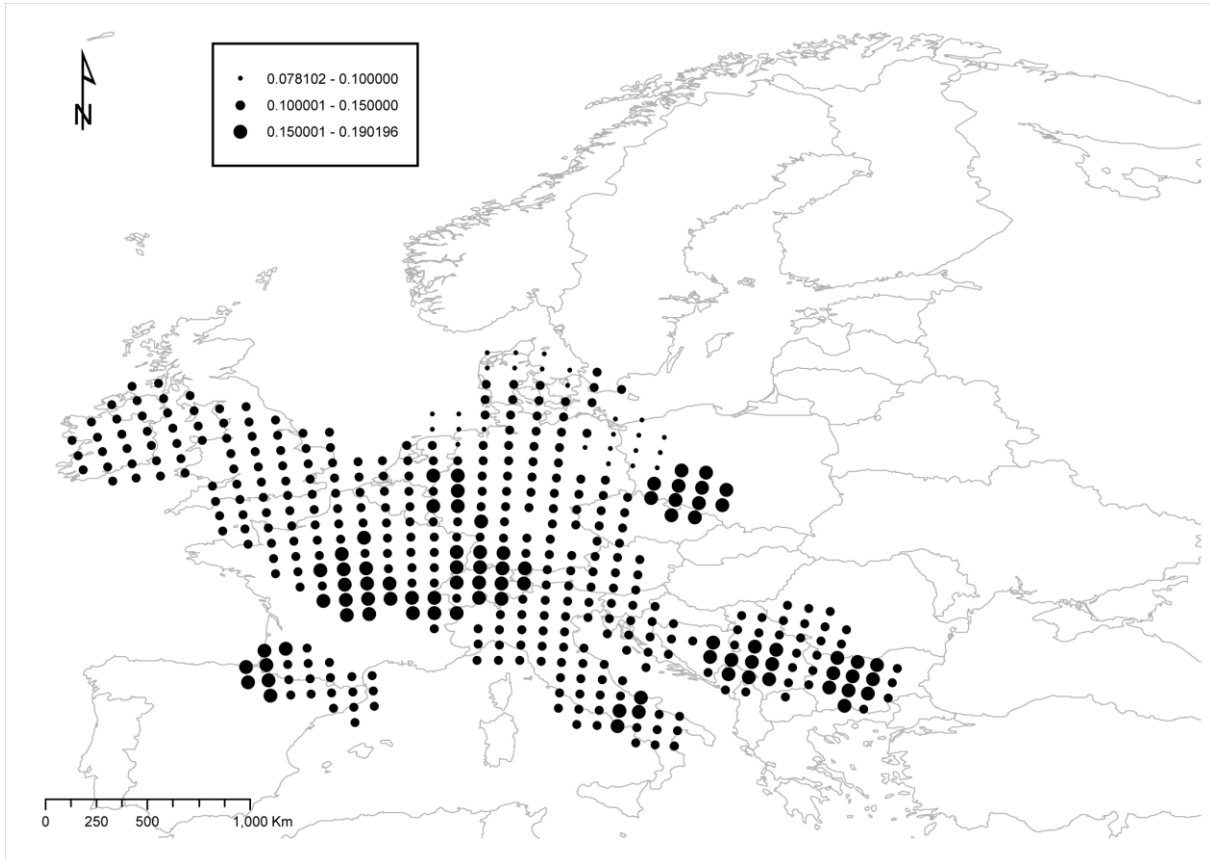
539 Supplementary Material 3 - Phylogeny of the two Psychodid sister species inferred applying
 540 Bayesian and Maximum Likelihood approaches. Support values are shown on branches,
 541 Bayesian/Maximum Likelihood.

542



543
 544 Both species are clearly separated, but no intraspecific structure is observed in any of the
 545 species.

546

547 Supplementary Material 4 - Nei diversities of *Arum maculatum*.

548

549 Nei genetic diversities (Nei & Li 1979) in *Arum maculatum* were calculated for samples
 550 having successfully amplified fragments with both AFLP primer pairs, applying a geographic
 551 moving window run on R 9.2.1 (R Core Development Group, 2009). The method consists of
 552 the following steps: i) the region of study is defined by the most extreme sampled locations,
 553 ii) a grid of fixed cell size (in our case 75 km) and their associated centroids are calculated,
 554 iii) starting at each centroid a fixed maximum number of samples is randomly selected (three
 555 in our case) in a user-defined perimeter (150 km in this study), iv) Nei diversities are
 556 calculated for each centroid, v) points iii and iv are repeated 10'000 times, vi) mean values
 557 are calculated for each centroid. This method allows not only to calculate the Nei diversity
 558 values at each location, but also to observe their variation and distribution throughout the
 559 area of study. Moreover, it allows for a correction of eventual local sampling biases since the
 560 analysis is grid-based. Scripts are available upon request to the first author.

561

Chapter six

Evolutionary history of Chiastocheta species interacting with Trollius europaeus

Anahí Espíndola, Sven Buerki
and Nadir Alvarez

1 **Evolutionary history of *Chiastocheta* species interacting with**
2 ***Trollius europaeus***

3

4 Anahí Espíndola¹, Sven Buerki² and Nadir Alvarez³

5

6 ¹Laboratory of Evolutionary Entomology, Institute of Biology, University of Neuchâtel.

7 Emile-Argand 11, 2000 Neuchâtel, Switzerland.

8 ²Molecular Systematics Section, Jodrell Laboratory, Royal Botanic Gardens, Kew,

9 Richmond, Surrey TW9 3DS, United Kingdom.

10 ³Department of Ecology and Evolution, University of Lausanne, Biophore Building, 1015

11 Lausanne, Switzerland

12

13 **Abstract**

14 The nursery pollination mutualism between *Trollius europaeus* and the species complex
15 of pollinating flies of the genus *Chiastocheta* has been studied for several years from both
16 behavioural and ecological perspectives. However, the species circumscription of several
17 pollinating flies is currently not clearly established, though this is a key point for further
18 investigating evolutionary forces driving the fate of this interaction.

19 Here, we aim at defining the identities and boundaries of the different *Chiastocheta*
20 species, giving special attention to those involved in the interaction with *T. europaeus*.
21 Based on a large-scale sampling, we apply a morphological survey and molecular-based
22 (nuclear and cytoplasmic) phylogenetic inference to i) date the origin of the genus
23 *Chiastocheta*, ii) infer its most likely biogeographic history, iii) define and clarify the
24 boundaries of taxa, iv) present a model explaining congruences and incongruences between
25 results and v) propose some guidelines for future taxonomic researches.

26 Our results demonstrate that the genus is relatively young (Pleistocene), and originated in
27 Europe. Only three species (*C. rotundiventris*, *C. dentifera* and *C. lophota*) involved in the
28 interaction with *T. europaeus* showed congruence between morphological and phylogenetic
29 inferences. The remaining species appear to present genetic admixture, with *C. setifera*
30 showing the highest level of hybridization.

31 This study demonstrates the importance of identifying potential genomic transfers
32 between morphospecies. In the framework of the study of coevolutionary processes, our
33 results warn researchers about the effect hybridization could have in the interpretation of
34 ecological results, since selective forces would be probably more diffuse than previously
35 thought, at least for those species frequently hybridizing.

36

37

38 Introduction

39 Understanding the nature of species and speciation are fundamental and much discussed
40 topics in biology (Mallet 2010). The classical species concept is based on Linnaeus'
41 typological philosophy (Ruse 1969), which considers species as static entities that can be
42 described based on shared morphological characters. This sight is still of high value today,
43 being particularly important in the field of applied conservation biology (Agapow, Bininda-
44 Emonds et al. 2004). Other theories such as the ecological (Van Valen 1976) or biological
45 (Mayr 1942) concepts are much more relevant from the theoretical point of view, but are also
46 more difficult to apply for direct organismal identification. In between these two extremes is
47 the phylogenetic species concept, which is based on molecular distances and evolutionary
48 histories, and which under its most extreme form considers that a species should be
49 delimited to all deeper taxa recognized in a phylogenetic classification (Mishler and Theriot
50 2000). Despite being sound, this concept is however also difficult to be used, because levels
51 of divergence considered to circumscribe species are variable among biological groups (de
52 Queiroz 2005) and because it does not necessarily allow any direct field identification of
53 entities (a goal of diversity studies). As clearly stated by de Queiroz (2005), the fact that
54 each concept analyzes evolutionary entities at different temporal scales, explains that
55 taxonomic units are not necessarily congruently delimited when different approaches are
56 used. Moreover, most concepts generally match when dealing with well-distinguished
57 lineages, but they are of limited application when treating complexes of species, especially if
58 they comprise lineages with similar ecologies and/or distributions, or which are experiencing
59 recent or ongoing genetic divergence processes. However, it is in those cases that it
60 becomes important to adopt combined approaches to define species, since the different
61 information each method provides could help disentangle complex situations.

62 Speciation processes can be influenced both by biotic and abiotic features (Ridley 1997).
63 When factors responsible for incomplete reproductive boundaries are related with abiotic
64 components (e.g., changes in topography driving vicariance of lineages followed by quick
65 admixture of lineages), scenarios are relatively simple to envisage. In contrast, when biotic

66 factors are the main drivers or limiting forces of among-lineage gene flow, understanding the
67 evolutionary history of these lineages might require a more integrative approach. The case
68 of the pollination mutualism between *Trollius europaeus* (Ranunculaceae) and its associated
69 *Chiastocheta* flies species complex (Diptera: Anthomyiidae) (Pellmyr 1989) is potentially one
70 of these biotic-influenced example. In this nursery pollination system, plants are pollinated by
71 insects, which lay eggs on flowers and whose larvae eat the developing floral structures
72 (Dufaÿ and Anstett 2003). Several adaptive traits are observed both in the plant and the flies:
73 on the one hand, the plant displays flowers presenting a closed shape that excludes an
74 important number of potential generalist pollinators and attracts high numbers of
75 *Chiastocheta* flies, providing them a mating site and a shelter against external conditions
76 (Pellmyr 1989). Studies have demonstrated that seed set is higher in closed than in open
77 flowers, and that visits by non-specific visitors are less efficient than those done by
78 *Chiastocheta* (Ibanez, Dujardin et al. 2009). It has also been observed that coloration of
79 flowers (orange-yellow) is adapted to optimally attract *Chiastocheta* flies (Pellmyr 1989), and
80 that the high number of carpels present in the flower appears to be related to predation costs
81 associated with nursery pollination (Pellmyr 1992). Finally, it has been shown that *T.*
82 *europaeus* produces feeding deterrent compounds that accumulate in its carpel walls, which
83 increase in concentration when *Chiastocheta* larvae feed on the seeds (Gallet, Ibanez et al.
84 2007; Ibanez, Gallet et al. 2009), and which have been proposed as a regulatory method of
85 the number of developing larvae per fruit and as a plausible starting point for explaining the
86 high diversification (in number of species) and specialization observed in this mutualism
87 (Ibanez and Després 2009). From the pollinators' side, fly lineages also present adaptations
88 to the interaction, which might be a consequence of their obligate development in *T.*
89 *europaeus* seeds: they display specific oviposition behaviors, with among-species
90 differentiation on the pattern (Pellmyr 1992) and timing of oviposition (Pellmyr 1989;
91 Pompanon, Pettex et al. 2006), and in the resource exploitation by developing larvae,
92 probably a consequence of interspecific competition and toxin escape (Pompanon, Pettex et
93 al. 2006).

94 This mutualistic system has been well studied in the European region, and it has been
95 demonstrated that at least seven *Chiastocheta* species, described based on morphological
96 grounds, contribute to pollination of the plant (*C. rotundiventris*, *C. dentifera*, *C. macropyga*,
97 *C. trollii*, *C. setifera*, *C. inermella*, *C. lophota*) (Pellmyr 1992). Even if researchers have been
98 working on Anthomyiidae for many years, the circumscription of fly species has been only
99 partially recovered. First, morphological identification keys exist for eggs of all species
100 (Pellmyr 1992), but only for some adult males and some females (Hennig 1976), while there
101 are up to now no possibilities of morphological identification of larvae. Second, because
102 rearing has been impossible *in vitro*, life cycles are not very well known, a situation that
103 hinders our understanding of the biology of the species. Finally, molecular studies performed
104 on this fly group have been focused on high taxonomic levels, comprising several –but not
105 all– species of the genus (Després, Pettex et al. 2002). Only three species interacting with *T.*
106 *europaeus* were shown to be monophyletic (*C. rotundiventris*, *C. macropyga* and *C.*
107 *dentifera*), whereas the other were para-/polyphyletic (*i.e.*, *C. inermella*, *C. lophota*, *C. trollii*
108 and *C. setifera*). Moreover, some taxa (*e.g.*, *C. inermella*) showed strong evidence of gene
109 flow or of incomplete ancestral polymorphism sorting. However, the survey lacked extensive
110 sampling, which might have allowed more robust conclusions on the nature of species,
111 especially considering their large distribution range (western Palearctic).

112 Because the system appears to present a complex history, understanding how
113 coevolutionary dynamics shape lineage divergence in pollinating flies might require a large-
114 scale and dense sampling, together with the use of various molecular markers. This might
115 allow recognizing and defining the associated fly species involved in this mutualism, as well
116 as the nature and intensity of the interaction itself. Since the distribution range of the plant is
117 important, going from the mountainous regions of southern Europe to Scandinavia, the
118 British Islands and occupying the north-eastern Eurasian plains up to the Ural (Meusel,
119 Jäger et al. 1965), and because the mutualism has been until now deeply studied only in the
120 Alps and eastern Scandinavia (*e.g.*, Pellmyr 1989; Jaeger and Després 1998; Hemborg and
121 Després 1999; Pompanon, Pettex et al. 2006; Ibanez, Dujardin et al. 2009), we cannot

122 assume that the same pollinator visitation patterns will be found everywhere, as has been
123 shown in the case of *Arum maculatum* and its fly pollinators (Espíndola, Pellissier et al. in
124 press). The aim of the present study is thus to provide new keys for the general
125 understanding of the identity, relationships and biogeography of *Chiastocheta* species
126 associated with *T. europaeus*. We propose here to first analyze a reduced dataset based on
127 samples of each morphological species (hereafter referred to as *morphospecies*) of the
128 genus *Chiastocheta* [including also those not interacting with *T. europaeus*, as in Després et
129 al. (2002)] in order to identify main lineages and clades and to infer the spatial and temporal
130 components of the biogeographic history of the genus. Afterwards, we aim at analyzing a
131 large-scale European sampling of fly species associated with *T. europaeus*. The
132 combination of these two approaches will let us identify limits among taxa using molecular,
133 morphological and spatial methods. We thus expect that the complementarity of these
134 approaches will allow i) identifying the dynamics and timing of colonization of the main
135 Eurasian regions by *Chiastocheta* lineages, ii) circumscribing species interacting with *T.*
136 *europaeus*, iii) defining the fly species' distribution ranges, based on adult identification and
137 iv) explaining possible reasons for barriers to hybridization between morphospecies.

138

139 **Material and methods**

140 ***Sampling***

141 Fly samples covering the whole European distribution range of their associated plant *T.*
142 *europaeus* were collected during springs and summers 2006, 2007 and 2008. Flies and/or
143 larvae were sampled directly from flowers and further preserved in 70% ethanol. All insects
144 were initially identified following Hennig (1976), which allowed a first sight on the general
145 specific diversity in each location. Based on this raw identification, a selection of up to three
146 individuals of the same morphospecies per sampled site was done, and determined using
147 expert knowledge (V. Michelsen, unpublished data) even for individuals in which
148 identification keys do not formally exist yet (*i.e.*, females in *C. macropyga*, *C. inermella*, *C.*

149 *lophota*; males in *C. lophota*). All double-checked samples were kept for genetic analyses
150 and were used to produce species' occurrence maps.

151 ***Molecular analyses***

152 Based on material obtained from the legs of insects (for adults) or from total extraction
153 (for 25 larvae), DNA was extracted using the QIAGEN DNeasy Animal tissue extraction kit
154 (QIAGEN, Hombrechtikon, Switzerland) following the protocol of the manufacturer. The
155 remaining parts (abdomen, rest of thorax and head) of samples were preserved in case of
156 further need to confirm species identification.

157 Nuclear (ITS) and mitochondrial (COI, COII, D-loop) regions were amplified and further
158 sequenced. Mitochondrial data was obtained for all samples, while ITS sequencing was
159 done only for a subset of all samples. PCR amplifications were performed using primers
160 shown in Table 1.

161 All reactions were prepared in a 20 µl mix, containing 0.5X buffer, between 1 and 2.5 mM
162 MgCl₂, 10mM dNTPs, 1 unit of GoTaq DNA polymerase (Promega, Dübendorf, Switzerland),
163 0.5 µM primers, 3 µl DNA and ran in a TGradient Thermocycler (Biometra, Goettingen,
164 Germany). Conditions were different depending of the amplified region. For COI, COII and
165 ITS the program started with 1:30 min at 95°C, followed by 40 cycles of 35 secs at 95°C, 1
166 min at 52°C, 45 secs at 72°C, and finishing with a final elongation of 8 min at 72°C. Because
167 the amplification of the D-loop provides a longer fragment harboring A/T-rich sequences, the
168 program consisted of 5 min at 95°C, followed by 35 cycles of 1 min at 95°C, 1 min of
169 annealing at 55°C and 2 min of elongation at 60°C; the program finished with 5 min of final
170 elongation at 60°C. PCR products were sequenced at Macrogen Inc. (South Korea) and
171 Fasteris SA (Switzerland). Chromatograms were visually corrected on ChromasPro 1.41
172 (Technelysium Pty. Ltd.) and sequences were aligned using the Clustal-Wallis algorithm for
173 the COI, COII and ITS regions, as implemented in BioEdit 7.0.4.1 (Hall 1999) and
174 “moderately accurate” strategy implemented in online MAFFT version 6 (Kato and Toh
175 2008; D-loop). All alignments were visually corrected using BioEdit 7.0.4.1. Gaps were
176 coded using FastGap 1.2 (Borchsenius 2009), which uses the simple gap-coding method of

177 Simmons and Ochoterena (2000). Number of constant, variable and parsimony informative
178 base pairs were calculated using MEGA 4.0 (Tamura, Dudley et al. 2007).

179 ***Divergence time estimation***

180 Four samples from each morphospecies associated with *T. europaeus*, and from
181 locations distributed throughout the whole sampling range covered by this study, were
182 chosen in order to complement those previously published by Després et al (2002). The
183 outgroup was *Delia radicum* L. (Diptera: Anthomyiidae), following Després et al. (2002).
184 Because we wanted to decrease the percentage of missing data as much as possible for the
185 dating approach, we chose to work on a matrix that comprised only COI and COII regions,
186 for which best-fit models were calculated using MrAIC (Nylander 2004) considering the
187 Akaike's Information Criterion (AIC; Akaike 1973). Specimens used in the dating approach
188 are listed in Supplementary Material 1.

189 Once the alignment was prepared, a partitioned coalescent Bayesian analysis was
190 performed on a reduced dataset (see above for more details) with BEAST 1.5.4 (Drummond
191 and Rambaut 2007). The COI and COII partitions were unlinked for the substitution model,
192 whereas they were linked for the estimation of the molecular clock (see below). The best-fit
193 model for both regions was the Hasegawa, Kishino and Yano (HKY) model with a parameter
194 to account for a gamma among-site rate heterogeneity. One run of 100×10^6 generations
195 was performed, sampling one tree every 1000 generations. Due to the lack of fossils for the
196 genus *Chiastocheta*, direct calibration of the tree topologies was not possible and branch
197 lengths and node ages were estimated by applying gene-specific mtDNA substitution rates.
198 According to Papadopoulou et al. (2010), the two most extreme insect mitochondrial
199 molecular clocks based on relaxed methods and fossil data correspond to means of 0.9%
200 MY^{-1} (0.78% MY^{-1} - 1.02% MY^{-1} ; Zakharov, Caterino et al. 2004) and 2.28% MY^{-1} (1.66%
201 MY^{-1} - 2.9% MY^{-1} ; Wahlberg 2006). We favoured an exploratory approach considering
202 these two drastically different substitution rates independently to investigate the effect of this
203 parameter on divergence time estimation as well as on the biogeographic history of
204 *Chiastocheta* (see below). For each of the two substitution rate values, an analysis was run

205 following Borer *et al.* (in press). A relaxed clock with log-normal branch length distribution
206 was used and a Yule speciation model was applied to model population size through time
207 (other prior parameters were set as default; Drummond, Ho et al. 2006). For each
208 parameter, convergence of the independent runs was confirmed by the examination of their
209 respective distributions in TRACER 1.4 (Rambaut and Drummond 2004). After removing a
210 10% burn-in period, 95% divergence time confidence intervals were plotted on a maximum
211 clade credibility tree using TreeAnnotator 1.5.4 (Drummond and Rambaut 2007).

212 ***Biogeographic analyses***

213 Following recommendations of Buerki *et al.* (in press), areas were defined based on the
214 sympatry (*i.e.*, distribution of the species) and geological criteria (*sensu* Sanmartín and
215 Ronquist 2004). Four areas were thus considered (map caption in Figure 1): i) Northeastern
216 Asia (occupying the Siberian plains and limited to the South by the mountain chains of
217 south-eastern Russia); ii) Europe (including all the western Eurasian territory and extending
218 as far east as the Ural); iii) Japan; iv) Southeastern Asia (the area to the south of
219 southeastern Russian mountains). Due to computational limitations and to improve the
220 convergence of Bayesian divergence time estimations (see above), a representative subset
221 of the main clades was selected and biogeographic analyses were performed on the
222 maximum clade credibility trees retrieved from the BEAST analyses. Terminals were coded
223 as in Espíndola *et al.* (2010) according to the location of each sample.

224 The dispersal-extinction-cladogenesis (DEC) likelihood model implemented in Lagrange
225 v.2.0.1 (Ree, Moore et al. 2005; Ree and Smith 2008) was used to investigate the
226 biogeographic history of the *Chiastocheta* spp. This method is a parametric, extended
227 version of the dispersal-vicariance analysis (Ronquist 1997) that estimates ancestral ranges,
228 transition rates between ranges, and biogeographical scenarios of range inheritance for a
229 group of taxa in a maximum likelihood framework (Ree and Sanmartín 2009). An advantage
230 of the DEC model is its ability to adapt a transition matrix (*i.e.*, Q matrix) to reflect the
231 changing palaeogeography, connections between areas (*e.g.*, land bridges) through time, or

232 dispersal capabilities of the group of interest (Buerki, Forest et al. in press). By taking
233 advantage of this flexibility, the Q matrix for analyses performed in this study was
234 constrained according to dispersal possibilities between areas (Supplementary Material 2). A
235 dispersal probability of 1.0 was thus set when areas were directly connected, whereas -in
236 order to maintain the reducibility of the Markov Chain (Buerki, Forest et al. in press)- it was
237 defined to 0.01 instead of 0 when this was not true. Considering this Q matrix and the two
238 dated maximum credibility clade trees obtained from the application of the different
239 substitution rates, two biogeographic analyses were run on Lagrange v.2.0.1. Finally,
240 following the approach described in Buerki et al. (in press), ancestral area reconstructions
241 for each node were plotted on the trees using pie charts. The collection of R scripts (R
242 Development Core Team, 2009) to achieve this task are available on request to SB.

243 ***Phylogenetic inferences in Chiastocheta species associated with T. europaeus***

244 Because topological searches were time- and resource-consuming (*i.e.*, due to the large
245 number of samples and the length of alignments), analyses were performed at different
246 levels and following complementary strategies. First, a dataset including all mitochondrial
247 sequences of the 399 *Chiastocheta* samples (see above) was analyzed under both
248 Maximum Likelihood (ML) and Maximum Parsimony (MP) approaches. It was for this dataset
249 impossible to perform Bayesian inferences, since analyses did not reach the required
250 parameter convergence. Second, a dataset including all nuclear sequences (237 samples)
251 was analyzed based on the ML, MP and Bayesian algorithms. For Bayesian inference, the
252 ITS model of evolution was estimated using MrAIC (Nylander 2004).

253 The ML searches were done using RAxML 7.2.6 (Stamatakis 2006) with 10'000 rapid
254 bootstrap analyses followed by the search of the best-scoring ML tree in one single run. The
255 three mtDNA regions were considered as one single partition (by applying the predefined
256 GTR+G model). Analyses were done at the CIPRES cluster (San Diego CA, USA).

257 Non-model based analyses in MP were performed using PAUP* 4b10 (Swofford 2003),
258 applying the parsimony Ratchet approach (as implemented in PAUPrat; Sikes and Lewis
259 2001) and based on ten independent searches of 200 iterations, as recommended by Nixon

260 (1999). For mitochondrial data, because searches identified only one most parsimonious
261 tree, we worked on the –unique- best tree, which had the advantage of carrying information
262 on branch lengths. For nuclear data, a majority-rule consensus topology was calculated from
263 the most parsimonious trees. Decay indices [or Bremer supports, (Bremer 1988)] were
264 calculated based on this topology, as implemented in TreeRot 3.0 (Sorenson and Franzosa
265 2007). Decay indices indicate the number of mutational steps needed to collapse a given
266 node, and thus indicate the strength of the clade.

267 Bayesian analyses were performed only for nuclear data (see above) on MrBayes 3.1.2
268 (Ronquist and Huelsenbeck 2003), which implements Monte Carlo Markov Chain (MCMC)
269 searches of topologies and parameters for which the data fits best. Six cold and six hot
270 chains were run for 50'000'000 generations in two independent runs. Convergence was
271 accepted when standard deviations reached values below 0.01 and when the Potential
272 Scale Reduction Factor index (Gelman and Rubin 1992) approached 1.0. We considered the
273 MCMC sampling sufficient when the Effective Sampling Size (ESS) was higher than 200 –
274 checked on TRACER v1.4 (Rambaut and Drummond 2004). Topologies were thus
275 calculated applying a burnin of 5'000'000 generations (considering thus a total of 270'006
276 trees) and a 50% majority-rule consensus tree was calculated.

277 Because one of our aims was to test the phylogenetic basis of each morphospecies,
278 congruence between taxonomic identifications (*i.e.* based on genital characters) and
279 monophyly was done by analyzing the position of morphospecies in the phylogeny.

280 In order to identify an eventual geographic structure of the different clades, their spatial
281 distribution was plotted on maps of the region, by exploiting the geographic coordinates
282 associated to each sample. This was done using ArcMap 9.3 (ESRI, Redlands CA, USA).

283

284 **Results**

285 ***Distribution of morphological species***

286 1611 insect samples (196 larvae and 1415 adults) were collected and identified at 53
287 locations from the whole distribution range of the plant. We double-checked a total of 372

288 samples, among which 72 were *C. rotundiventris*, 67 *C. inermella*, 54 *C. trollii*, 52 *C. setifera*,
289 49 *C. lophota*, 41 *C. macropyga* and 38 *C. dentifera*. Along with 27 larvae, we performed
290 molecular analyses on a total of 399 samples (Supplementary Material 3).

291 Distribution ranges of morphologically identified species corresponding to the 373 adults
292 analyzed are depicted in Supplementary Material 4. While some species are distributed over
293 a wide part of the sampled area (*C. rotundiventris*, *C. inermella*, *C. setifera*, *C. dentifera*, *C.*
294 *trollii*), others present either fragmented distribution ranges (*C. macropyga*) or are restricted
295 to a specific area of the sampled zone (*i.e.*, *C. lophota* covers the southern part of Europe).

296 ***Divergence time estimations and biogeographic history***

297 47 samples provided 1134bp for the mtDNA-based analysis at the whole genus level.

298 Gaps were retrieved neither in COI nor in COII.

299 Because dating was done applying two independent substitution rates, ages of inferred
300 nodes differed between analyses (Figure 1). The origin of the *Chiastocheta* genus thus goes
301 back as far as 2.1My or 830 Ky, when applying a mutation rate of 2.28% or 0.9%,
302 respectively. Phylogenetic reconstruction inferred using both mutation rates provided
303 congruent topologies, which only differed in the position of the clade comprising *C. pellmyrii*
304 and *C. curvibasis*, two species not associated with *T. europaeus* (Pellmyr 1992).

305 Biogeographic ancestral reconstructions indicated that the most probable ancestral area
306 at the origin of the genus was Europe, with different numbers of identified dispersal events
307 as a function of the mutation rate applied. When setting a low substitution rate (0.9%) we
308 inferred five probable dispersal events (*i.e.*, three from Europe to Northeastern Asia, one
309 from central Asia to Japan and one from Northeastern Asia to eastern Asia). Alternatively,
310 we found four likely dispersal events when fixing a higher mutation rate (2.28%) (*i.e.*, two
311 from Europe to northeastern Asia, one from northeastern Asia to Japan and one from
312 northeastern Asia to eastern Asia).

313 ***Phylogenetic analyses of the dataset comprising Chiastocheta spp. associated*** 314 ***with T. europaeus***

315 399 and 237 samples provided 2304bp for mitochondrial and 441bp for nuclear regions,
316 respectively. While nuclear data contained seven gaps, mitochondrial sequences comprised
317 237 gap positions (*i.e.*, all in the D-loop region). Numbers of constant, variable, parsimony-
318 informative and total base pairs are presented in Table 2. While all mitochondrial regions
319 had a high level of variability (up to 44.05%), the nuclear region presented a particularly low
320 number of variable (3.63%) or potentially parsimony informative (2.49%) sites. The best
321 model of evolution suggested by the Akaike's Information Criterion (AIC) for ITS was F81
322 (Felsenstein 1981).

323 As shown in Figure 2, topologies obtained based on nuclear and mitochondrial data were
324 only partially congruent (Table 3a; see below), and none showed a high level of compatibility
325 between retrieved clades and morphospecies (Table 3b and c).

326 The nuclear topology (Figure 2a, Table 3b) showed the presence of at least five groups.
327 Only one clade ("rotundiventris", ML bootstrap value = 96; Bayesian Posterior Probability =
328 1) fully corresponded to the morphospecies *C. rotundiventris*, including all and exclusively
329 samples having been identified as this species. Clade "trollii" (only supported in the ML
330 topology; Bootstrap value = 84) comprised all but two samples of those assigned to *C. trollii*,
331 as well as four *C. setifera*. Clan "nl" included all samples of *C. dentifera*, *C. macropyga*, all
332 but two of *C. inermella* and around one quarter of all *C. setifera* samples. Clan "lophota"
333 contained all but one *C. lophota* and one *C. setifera*. Clade "setifera" (ML bootstrap value =
334 91, Bayesian Posterior Probability = 0.91, MP Decay Index = 1) was mainly formed by *C.*
335 *setifera*, even if it also included two samples of *C. inermella*.

336 Topologies inferred from mitochondrial data (Figure 2b, Table 3c) indicated that there
337 were five clades or groups, more or less supported depending on the analytical approach. *C.*
338 *rotundiventris* was well identified as a genetically different group (ML bootstrap value = 99),
339 completely included in clade "rotundiventris" and comprising also 17 of the larvae analyzed.
340 The same was true for *C. dentifera*: clade "dentifera" harbored all but one sample of this
341 species (ML bootstrap value = 100); and to a lesser extent for *C. lophota*: all but four
342 samples of this species were included in clade "lophota" (ML bootstrap value = 100), which

343 also included one larva. The two remaining clades comprised a mix of the other species: on
344 the one hand clade “mtI” (ML bootstrap value = 91) included samples belonging to different
345 morphospecies (mainly *C. setifera*, *C. trollii* and *C. inermella*) as well as three larvae. Group
346 “mtII” included all *C. macropyga* but also presented an important mixture of morphospecies,
347 especially *C. inermella*, *C. setifera*, and to a lesser extent *C. lophota*.

348 Distribution of the different mitochondrial and nuclear clades, in relation to morphospecies
349 are shown in Figures 3 and 4. Because in the cases of *C. rotundiventris* (Figure 3a), *C.*
350 *dentifera* (Figure 3b) and *C. lophota* (Figure 3c) mitochondrial clades and morphospecies
351 presented similar compositions, their distributions were the same. While *C. rotundiventris* is
352 widespread across Europe, *C. dentifera* is not present in the British Isles, and *C. lophota* is
353 only present in southern mountain massifs.

354 Mitochondrial clade “mtI” (Figure 4a-c) is present in the centre and north-west of the
355 sampled region, and is especially absent of the Balkans. Samples from this clade were
356 distributed in three nuclear clades (Table 3a) and included the totality of clade “trollii” (Figure
357 4a), some individuals from “nl” (Figure 4b), and a portion of samples from clade “setifera”
358 (Figure 4c). The latter is restricted to the (central and Eastern) Alps, and includes all *C.*
359 *setifera* morphospecies from this region. The only two Scandinavian *C. setifera* samples
360 present in this cytoplasmic clade are included in “trollii”.

361 Mitochondrial group “mtII” (Figure 4d-f) had representatives in almost every sampled site,
362 and was the only homogeneously occupying the Balkans. Samples from this clade were
363 included in geographically structured nuclear clades (Table 3a). Clade “nl” (Figure 4e) was
364 present in Scandinavia, central and Eastern Europe, and in one population of the British
365 Isles. Nuclear clade “setifera” (Figure 4f) is on the other hand distributed in the Southern
366 regions and is clearly dominant in the British Isles. Only two sites presented samples
367 included in nuclear “lophota” clade (Figure 4d). Finally, *C. macropyga* is completely included
368 in clade “nl” (pink, Figure 4e).

369

370 Discussion

371 While topologies retrieved with the large dataset (*i.e.*, comprising 399 *Chiastocheta*
372 samples from seven species; Figure 2) were compatible at the level of main clades whatever
373 phylogenetic criterion was applied (*i.e.*, differences in the relationships among main clades
374 are only due to one single polytomy in the ML analysis), the topology retrieved in the
375 Bayesian inference analysis involving 47 representatives from twelve *Chiastocheta* species
376 (Figure 1) was substantially different, probably because this analysis comprised more taxa
377 but less mtDNA regions (only COI and COII were analyzed). However, despite the position
378 of four out of the five main clades was different, all topologies concurred in placing *C.*
379 *rotundiventris* as the most basal lineage. Relationships among the other four main lineages
380 thus require further analyses of other DNA regions. Nonetheless, it is worth noting that
381 several similarities were found when this topology was compared with that obtained by
382 Després et al. (2002): *C. rotundiventris* appears monophyletic and basal, while *C. dentifera*,
383 *C. latispinigera*, *C. curvibasis* and *C. pellmyri* are monophyletic. In contrast, we could not
384 retrieve the monophyly of *C. macropyga*, indicating that an increase in the number of
385 samples per species appears to have an effect on the retrieved level of taxon monophyly.
386 *Chiastocheta lophota*, *i.e.*, the only species lacking in previous phylogenetic surveys of the
387 genus, was placed in our chronogram as the sister lineage of *C. latispinigera* (Figure 1).
388 Furthermore, considering the important genital morphological similarities, it is highly likely
389 that the sample identified as *C. inermella* (inermella4) by Després et al. (2002) belongs in
390 fact to *C. lophota*.

391 ***Divergence time estimations***

392 Recent studies (Papadopoulou, Anastasiou et al. 2010) have pointed out the importance
393 of the use of molecular clocks that take into account the heterogeneity of substitution rates
394 among markers when performing temporal inferences. Because we did not have the
395 possibility to use fossil data to calibrate our analysis, we applied two extreme substitution
396 rates obtained from recent fossil-based molecular clock calibration studies (*i.e.*, Zakharov,
397 Caterino et al. 2004; Wahlberg 2006). Even if doing so did not allow us to consider variation
398 between markers, it provided a good way to compare two extreme biogeographic and dating

399 scenarios for the genus *Chiastocheta*. Our results thus associate nodes to a range of ages
400 that varies between the two analyses, but which place the root of the genus between the
401 Ionian Middle Pleistocene (around 830 Kya) and the Gelasian Lower Pleistocene (around
402 2.1 Mya) for high (2.28%) or low (0.9%) mutation rates, respectively (Figure 1). Irrespectively
403 of the kind of substitution rate used, our results concurred in placing the origin of genus
404 *Chiastocheta* sometime during the Pleistocene, indicating that it is relatively young. A similar
405 conclusion was also reached by Després et al. (2002) when applying a standard molecular
406 clock (Brower 1994) which estimated the origin of the genus at 1 Mya.

407 ***Biogeographic history of genus Chiastocheta***

408 Biogeographic analyses in a Lagrange framework (Ree, Moore et al. 2005; Ree and
409 Smith 2008) consider branch lengths for the estimation of the most probable ancestral area.
410 Practically this means that vicariance, extinction and dispersal events have more chances to
411 happen when inferred on long rather than on short branches. In this study, while analyses
412 based on the two different mutation rates provided very similar inferences of ancestral areas,
413 they informed on different numbers of dispersal events. Our inferences (Figure 1) indicate
414 that an early event of colonization by a lineage ancestral to *C. rotundiventris* appeared to
415 have invaded Asia and to have diversified in that region during the Calabrian age (1.8 - 0.7
416 Mya). Later (between the Calabrian and Ionian ages; 1.8 – 0.12 Mya), while a European
417 lineage survived in Europe, another (or two others in the 0.9 % analysis) dispersed and
418 established in northeastern Asia, and from there dispersed to southeastern Asia and Japan
419 afterwards.

420 The genus appears to have diversified during the Pleistocene, a period recognized by its
421 marked glacial ages (Ehlers and Gibbard 2004). In the northern hemisphere, the distribution
422 of biomes was shifted to southern regions during these ages, and arctic species appeared to
423 be much more widespread than today (e.g., Espíndola et al., submitted; Brochmann,
424 Gabrielsen et al. 2003). It is possible that during these distributional expansions experienced
425 throughout glacial periods, flies of this genus could have been able to disperse eastwards
426 and lineages might have been isolated through vicariance processes.

427 ***Phylogeny, distribution of clades in relation to morphospecies and reinterpretation***
428 ***of species***

429 A comparison of species assignments in both mtDNA and ITS analyses indicated some
430 similarities in the phylogenetic inferences, but also several substantial differences. The main
431 common pattern was that both analyses placed *C. rotundiventris* as a clearly defined entity
432 (Figure 2, Table 3). It is important to note that all samples having been identified as
433 belonging to this species were included in this supported clade. This clear molecular
434 assignment could be explained by an early divergence in the genus (Figure 1), which
435 allowed increasing reproductive barriers with other lineages. From a morphological point of
436 view, however, *C. rotundiventris* should be sister to *C. abruptiventris*, its supposed
437 Scandinavian vicariant (Pellmyr 1992). Nonetheless, even if morphological differences have
438 been found between the two entities (V. Michelsen, unpublished data), our phylogeny
439 provided no support to this taxon (no special clustering of samples from Scandinavia). Based
440 on this, it seems thus more likely that the identified morphological differences represent local
441 variations/adaptations eventually related to phenotypic plasticity.

442 The nuclear-mitochondrial congruence presented in the case of *C. rotundiventris* has
443 been observed in none of the other clades/morphospecies, indicating that either an
444 important rapid radiation in this group is happening, or/and there is/has been intensive
445 hybridization between several lineages.

446 Among those species presenting incongruence between morphological and molecular
447 identities, an important proportion of samples identified as *C. lophota* have also been
448 grouped by molecular analyses (yellow, in Figure 2, Table 3): 92% of samples described as
449 *C. lophota* are included in the “lophota” mitochondrial clade, while only two samples of
450 morphospecies *C. setifera* and *C. dentifera*, respectively are also contained by this clade.
451 Moreover, all *C. lophota* samples were included in the “lophota” nuclear clan; however, this
452 clan also comprises some samples of other species (two *C. macropyga*). Even if the nuclear
453 phylogeny did not provide any clade harboring exclusively *C. lophota*, it seems that either
454 the age of the group (this clade is the third oldest in the genus; Figure 1) or/and its

455 geographic isolation (the taxon is currently only found in southern European mountains)
456 might have allowed genetic isolation despite rare events of hybridization.

457 Another well-supported mtDNA clade was the one comprising virtually only and almost all
458 (97%) *C. dentifera* morphospecies (green in Figure 2, Table 3). All samples from this clade
459 fell in the “nl” nuclear group. It is interesting noting that the mtDNA clade comprising *C.*
460 *dentifera* samples follows temporally that of *C. rotundiventris*. Similar to the treatment of *C.*
461 *lophota*, we consider that there was enough morphological (Hennig 1976), phylogenetic
462 (Figure 1 and 2) and behavioral arguments (i.e., this is the only species laying eggs in
463 central carpels at the end of flowering, Pellmyr 1989) to accept the consistency of this
464 species, despite rare occurrences of hybridization.

465 Mitochondrial “clade mtI” included samples of all species (blue-purple in Figures 2 and
466 4a-c, Table 3), except *C. rotundiventris* and *C. macropyga*. All but one *C. trollii*, 79% of *C.*
467 *inermella*, 52% of *C. setifera* and one sample of *C. dentifera* and *C. lophota* were included in
468 the group. A remarkable point is that all *C. trollii* samples of this clade (93% of all those
469 analyzed with nuclear markers) were restricted to nuclear clade “trollii” (purple in Figure 2a),
470 which indicates some genetic consistency of this morphospecies, at least from a nuclear
471 point of view, and which would provide some support to the validity of this taxon.
472 Contrastingly, an important number of the Alpine *C. setifera* included in this clade (black in
473 Figures 4d-f), were placed in nuclear clade “setifera”, while the remaining were included in
474 clan “nl”. It seems thus correct to consider that all *C. setifera* samples included in clade
475 “setifera” are morphospecies carrying non-hybridized genomes, while the remaining
476 probably resulted from crosses between *C. setifera* and other species (see below). It is
477 plausible that at some point in the history of diversification of the species, *C. setifera* was
478 isolated of all other taxa, which allowed a certain differentiation. It could be possible that this
479 diversifying group came afterwards into contact with other species, as *C. trollii* and *C.*
480 *inermella*; hybrids could be today represented by all samples morphologically identified as *C.*
481 *setifera* but included in clan “nl” and clade “trollii”. However, because of the extremely high
482 levels of admixture, the identity of *C. setifera* should be redefined, at least from a

483 morphological point of view. While doing so, it would be highly recommended to consider
484 especially those samples having been identified as this species *and* being included in the
485 “setifera” nuclear clade; since it is likely that they are harboring non-hybridized genomes.

486 Another interesting point concerning clade “mtI” is that an internal subclade harboring
487 exclusively Scandinavian *C. inermella* samples was observed in the ML approach (Figure 2b
488 and Figure 4e). Even if this clade had a support value falling below 50%, it is worth noting it,
489 since i) these samples also clustered together in the MP approach, and ii) this is one of the
490 only subclades of “mtI” containing exclusively samples of one morphospecies. If this clade
491 was correct, it would validate the formerly proposed idea of *C. inermella* as an exclusive
492 Scandinavian species (Pellmyr 1992). It is however important to realize that nuclear results
493 did not confirm this point (Figure 2a), which is why we recommend the redefinition of this
494 entity.

495 Mitochondrial clan “mtII” (Figure 4d-f, Table 3) contained all *C. macropyga* but also
496 samples assigned to morphospecies *C. inermella* (16%), *C. setifera* (28%) and *C. lophota*
497 (3%) indicating that this group is far from being homogeneous. Despite this admixture, when
498 analyzed with nuclear markers, all samples identified as *C. macropyga* were comprised in
499 nuclear clan “nI” (pink in Figure 2a, Figure 4e, Table 3), while all remaining *C. lophota* were
500 included in clan “lophota” (Figure 4d), and all *C. setifera* and two *C. inermella* were
501 comprised in clade “setifera” (Figure 4f). This could indicate that in this clan *C. macropyga* is
502 the most consistent genetic entity, clustering uniformly with both types of genetic markers
503 (Table 3). On the other hand it appears, as mentioned above, that *C. setifera* and *C.*
504 *inermella* are experiencing genetic admixtures, which would confirm our doubts about their
505 identities.

506 ***Genetic boundaries of morphospecies***

507 Considering the complementary phylogenetic (Figure 2), spatial (Figures 3 and 4) and
508 morphological (Table 3b and 3c) approaches applied to the analysis of the *Chiastocheta*
509 species complex associated to *T. europaeus*, we propose here some ideas on the genetic
510 interspecific dynamics of the group (Figure 5). We find three different situations: i) species

511 are well defined and no hybridization is observed (*C. rotundiventris*); ii) morphology and
512 phylogenetics are congruent, but some rare hybridization events are observed (*C. dentifera*
513 and *C. lophota*); iii) hybridization is observed, which can be explained by spatial and
514 ecological reasons; species borders are in such cases less clearly circumscribed (*C.*
515 *macropyga*, *C. trollii*, *C. setifera* and *C. inermella*).

516 *Well defined species, no hybridization*

517 There are no doubts about the delimitation of *C. rotundiventris* (Figure 2a-b). It has been
518 demonstrated that this species is the most basal in the genus (Figure 1 and Després, Pettex
519 et al. 2002), which indicates that the genetic distance with other lineages is the largest,
520 which could have consequences on limiting genetic compatibility with other species.
521 Moreover, behavioral studies have demonstrated that this species visits flowers significantly
522 earlier than the other species (Després and Jaeger 1999), which could probably define a
523 temporal barrier for mating. Finally, there are clear male and female genital morphological
524 differences between this and the other species (Hennig 1976, V. Michelsen pers. com.),
525 which could represent a physical barrier to hybridization.

526 *Well defined species, rare hybridization events*

527 The specific identity of *C. dentifera* is relatively clear, especially regarding the mtDNA
528 phylogeny (Figure 2b). An important amount of samples (97%) identified as *C. dentifera* falls
529 in the same exclusive mitochondrial clade. The differential genetic identity of this species
530 could be explained both by historical, morphological and behavioral reasons. Previous works
531 (Després, Pettex et al. 2002) and this study demonstrate that this species diverged early,
532 which could indicate that it had been evolving for enough time before other *Chiastocheta*
533 lineages radiated, which has led to a high level of genetic isolation. Moreover, behavioral
534 studies have also demonstrated that this species visits the flower at the very end of the
535 flowering time (Després and Jaeger 1999), when other species have generally already
536 visited *T. europaeus* inflorescences, which could directly diminish the probability of mating
537 with other species and thus reduce hybridization rate. Finally, even if there are genital
538 differences between this and the other species, they are not as important as those presented

539 by *C. rotundiventris*, which could also explain that some hybridization seems to happen
540 (Figure 2 and Figure 4b). It is in our point of view very probable that the largest barrier to
541 hybridization is in this case ecological/genetic and not necessarily morphological. However,
542 in order to confirm this, developmental studies and hybridization tests should be performed
543 on this species.

544 The case of *C. lophota* is similar to that of *C. dentifera*, with the difference that very little is
545 known in what concerns flower visitation patterns. We can however confirm that this species
546 is not *C. inermella*'s vicariant as previously proposed (Pellmyr 1992). Even if our study has
547 not investigated behavioral topics, we imagine that there should be some behavioral
548 differences that would enhance the separation of species, especially considering that
549 morphological genital differences with non-*rotundiventris* types are not very important
550 (Michelsen V., unpublished data), and that, despite sharing an important part of its
551 distribution range with other species, it only rarely hybridizes.

552 *Morphological differences, hybridization observed*

553 This category corresponds to *C. macropyga*, *C. trollii*, *C. setifera* and *C. inermella*. Among
554 these four taxa, those harboring stronger phylogenetic support are *C. macropyga* -
555 completely included in groups “mtII” and “nl”-, and *C. trollii* -found only in clades “mtI” and
556 “trollii”. Interestingly, both *C. macropyga* and *C. trollii* harbor very distinctive genital
557 differences (males of *C. macropyga* present big surstili; females of *C. trollii* have small and
558 short ovipositors, morphologically closer to *C. rotundiventris* than to the others; Hennig
559 1976).

560 One might ask whether or not it was possible to predict hybridization among these four
561 morphospecies. From a morphological point of view, females among three out of the four
562 types (*C. setifera*, *C. inermella* and *C. macropyga*) are similar, while males of three of them
563 have similar types of genitalia (*C. setifera*, *C. inermella* and *C. trollii*) (Hennig 1976; V.
564 Michelsen, pers. comm.). This morphological similarity could, if not explain hybridization, at
565 least allow it, since no important physical barriers to mating would be present between
566 morphospecies. From a genetic point of view, the four species appear closely related in the

567 two mitochondrial clades “mtI” and “mtII” (Figure 2), indicating that speciation processes
568 could be recent, and that genetic incompatibility and/or differentiation is probably still an
569 ongoing process.

570 A coupled analysis of nuclear-cytoplasmic incongruences (Figure 2, Table 3) and of the
571 phylogeography of the four morphospecies (Figure 4) informs us on the direction and type of
572 crossings that seem to be happening in the group. This combined analysis indicated that
573 hybridization does not appear to occur at the same rate among all species (see below).
574 Based on this, we defined a simple model (Figure 5) which takes this fact into account and
575 explains the incongruences observed in our results.

576 The presence of *C. setifera* in both mitochondrial groups “mtI” and “mtII” (Figure 2b),
577 associated with its wide distribution range (Figure 4), suggests that this species is
578 hybridizing with the other four, and that in some cases it is also acting as a “genome carrier”
579 between the remaining species (especially demonstrated in an internal clade of clan “mtII” in
580 Figure 2b, where no admixture is observed between *C. macropyga* and *C. inermella*).
581 Moreover, the only one among the four species showing some geographic structure is *C.*
582 *setifera* (*i.e.*, all Alpine samples of nuclear “setifera” are in clade “mtI”, while all non-Alpine
583 are in clade “mtII”; Figure 4) which indicates that this species is the only one observed in two
584 mitochondrial groups. Based on these arguments we propose here two hybridizations
585 “types”, which seem to be concerning two different *C. setifera* lineages: one Alpine (“mtI”)
586 and one non-Alpine (“mtII”).

587 The Alpine lineage hybridizes with species available in that region: *C. inermella* and *C.*
588 *trollii* (Figure 4a-c and Figure 5). It is interesting noting that *C. trollii* was very well identified
589 by nuclear markers and is highly represented in the Alpine zone, while this is less the case
590 for northern locations, which probably provides a higher probability of hybridization in the
591 Alps than in northern areas.

592 On the other hand, the non-Alpine lineage of *C. setifera* is only present in “mtII”, where it
593 is admixed with *C. inermella* (Figure 4d-f), an entity much more widely distributed here than
594 in the Alps (Figure 4 and 5). It also admixes here with *C. macropyga*, but considering the

595 homogeneity in the distribution and genetic assignments of morphospecies of this species
596 (Table 3, Figures 2 and 4), this seems to be happening at a lower rate, probably due to the
597 more important genital differences found between species. Moreover, the absence of *C.*
598 *inermella* in the two internal –non supported- clades of “mtII”, could indicate that no gene
599 flow is happening between this and *C. macropyga*, or that if some gene flow is present it
600 happens indirectly, through *C. setifera* (Figure 5).

601 ***The species concept in Chiastocheta***

602 As suggested by de Queiroz (2005), it appears that the example of *Chiastocheta* species
603 represents one of those puzzling taxonomic cases in which speciation and diversification are
604 still going processes, and that depending on the approach applied (*i.e.*, morphology,
605 ecology, molecular differentiation), the delimitation of entities varies. In this study however,
606 we demonstrate that the simultaneous evaluation of complementary approaches allows the
607 definition and a better understanding of the speciation process happening in *Chiastocheta*
608 flies associated with *T. europaeus*.

609 Based on our analyses we recommend that the classic systematics of the group is
610 maintained for all species, besides *C. setifera* and eventually *C. inermella*. In spite of this,
611 future ecological studies should not ignore the fact that genetic limits of typological species
612 (*i.e.*, morphospecies) are not as strict as formerly supposed, which could have a non-
613 negligible influence on the interpretation of ecological and coevolutionary studies.

614 Moreover, as demonstrated in this survey, there is a spatial effect in the genetic dynamics
615 of at least one morphospecies (*C. setifera*). This effect should also be tested at an ecological
616 level, since it is also possible that isolation and especially long-term cohabitation with
617 different sets of species could induce taxa to behave differently in different geographic
618 areas. In order to better understand such interspecific interactions, it is necessary to
619 investigate behavioral features of all species and at different localities of their distribution
620 ranges. It is possible that once this is done, we will be able to better understand the variation
621 of the relationship at a spatial scale, not only in terms of number and identities of
622 morphological species (as presented in this work), but also in terms of ecological functions.

623

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630

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764 (Lepidoptera : Papilionidae)." Systematic Biology **53**(2): 193-215.
- 765
- 766

767 Table 1 – Regions, primer sequences, annealing temperatures and references used
 768 for PCR amplification.

| Region | Primer | Sequence | Annealing | Reference |
|--------|---------------------------|-----------------------------------|-----------|--------------------------|
| COI | COI-2171 | TTG ATT TTT TGG TCA YCC NGA AGT | 52 | Després and Jaeger, 1999 |
| | tRNA ^{Leu} -3048 | TGG AGC TTA AAT CCA TTG CAC | 52 | Després and Jaeger, 2000 |
| COII | tRNA ^{Leu} -3023 | GAT TAG TGC AAT GGA TTT AGC TC | 52 | Després and Jaeger, 2001 |
| | COII-3683 | CCR CAA ATT TCT GAA CAT TGA CC | 52 | Després and Jaeger, 2002 |
| D-loop | TM-N-193 | TGG GGT ATG AAC CCA GTA GC | 55 | Simon et al. 1994 |
| | SR-J-14612 | AGG GTA TCT AAT CCT AGT TT | 55 | Simon et al. 1994 |
| ITS2 | ITS2DF | CCT GGT TAG TTT CTT TTC CTC CGC T | 56 | Depaquit et al., 2000 |
| | ITS2DR | CGC AGC TAA CTG TGT GAA ATC | 56 | Depaquit et al., 2000 |

769

770 Table 2 – Number and percentage of constant (C), variable (V), parsimony informative (PI)
 771 and total base pairs, for the four different amplified regions. Analysis of the D-loop did not
 772 allow overlap between forward and reverse cycle sequencing (values are presented for each
 773 primer/direction, independently). Percentages are shown between parentheses.

| <i>Mitochondrial</i> | C | V | PI | Total |
|----------------------|-------------|-------------|-------------|-------------|
| COI | 489 (74.43) | 168 (25.57) | 77 (25.57) | 657 |
| COII | 268 (55.95) | 211 (44.05) | 39 (8.14) | 479 |
| D-loop IF | 277 (78.47) | 76 (21.53) | 49 (13.88) | 353 |
| D-loop IIR | 577 (79.80) | 238 (29.02) | 155 (19.02) | 815 |
| <i>Total</i> | | | | <i>2304</i> |
| <i>Nuclear</i> | | | | |
| ITS 2 | 425 (96.37) | 16 (3.63) | 11 (2.49) | 441 |

774

775 Table 3 – Contingency tables of assignments of samples to a) mitochondrial vs. nuclear
 776 clades; b) nuclear vs. typological/morphological species; c) mitochondrial vs.
 777 typological/morphological species. Names correspond to clades following Figure 2. In a), NA
 778 indicates samples that have been only analyzed using the mitochondrial approach.

a)

| | | Nuclear clade | | | | | NA | Total |
|------------------------|----------------|----------------|---------|---------|----------|----|-----|-------|
| | | rotundiventris | trollii | lophota | setifera | nl | | |
| Mitochondrial clade | rotundiventris | 40 | | | | | 49 | 89 |
| | dentifera | | | | | 13 | 25 | 38 |
| | lophota | | | 36 | | | 11 | 47 |
| | clade mtl | | 34 | | 14 | 38 | 52 | 138 |
| | clade mtll | | | 2 | 18 | 40 | 27 | 87 |
| Total | | 40 | 33 | 38 | 32 | 92 | 164 | 399 |

779
 780
 781

b)

| | | Nuclear clade | | | | | Total |
|---------------|--------------------------|----------------|---------|---------|----------|----|-------|
| | | rotundiventris | trollii | lophota | setifera | nl | |
| Morphospecies | <i>C. rotundiventris</i> | 40 | | | | | 40 |
| | <i>C. dentifera</i> | | | | | 14 | 14 |
| | <i>C. lophota</i> | | | 37 | | 1 | 38 |
| | <i>C. inermella</i> | | | | 2 | 38 | 40 |
| | <i>C. trollii</i> | | 30 | | | 2 | 32 |
| | <i>C. macropyga</i> | | | | | 28 | 28 |
| | <i>C. setifera</i> | | 4 | 1 | 30 | 8 | 43 |
| Total | | 40 | 34 | 38 | 32 | 91 | 237 |

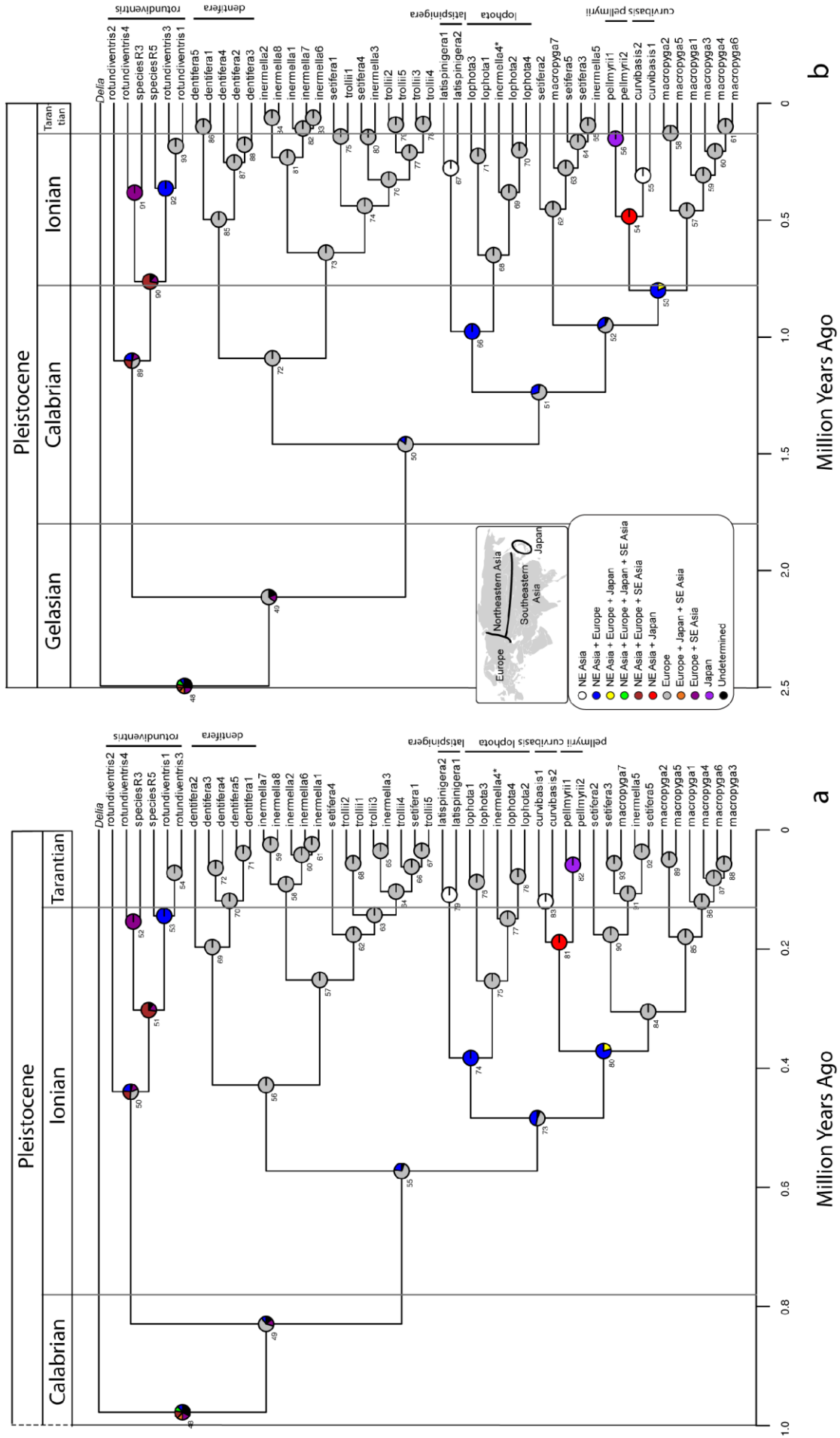
782
 783
 784

c)

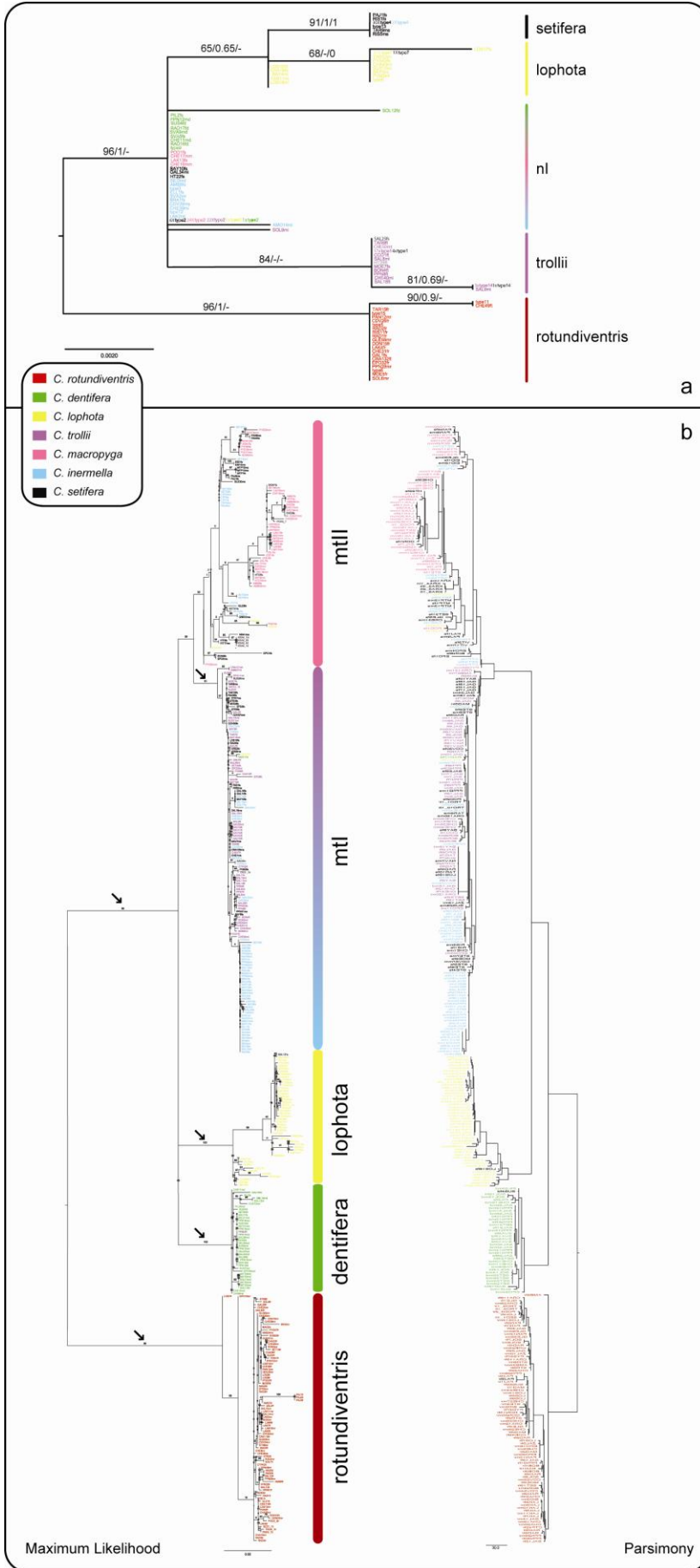
| | | Mitochondrial clade | | | | | Total |
|---------------|--------------------------|---------------------|-----------|---------|-----|------|-------|
| | | rotundiventris | dentifera | lophota | mtl | mtll | |
| Morphospecies | <i>C. rotundiventris</i> | 72 | | | | | 72 |
| | <i>C. dentifera</i> | | 37 | | 1 | | 38 |
| | <i>C. lophota</i> | | | 45 | 1 | 3 | 49 |
| | <i>C. inermella</i> | | | | 53 | 14 | 67 |
| | <i>C. trollii</i> | | 1 | | 53 | | 54 |
| | <i>C. macropyga</i> | | | | | 41 | 41 |
| | <i>C. setifera</i> | | | 1 | 27 | 24 | 52 |
| | larvae | 17 | | 1 | 3 | 5 | 26 |
| Total | | 89 | 38 | 47 | 138 | 87 | 399 |

785

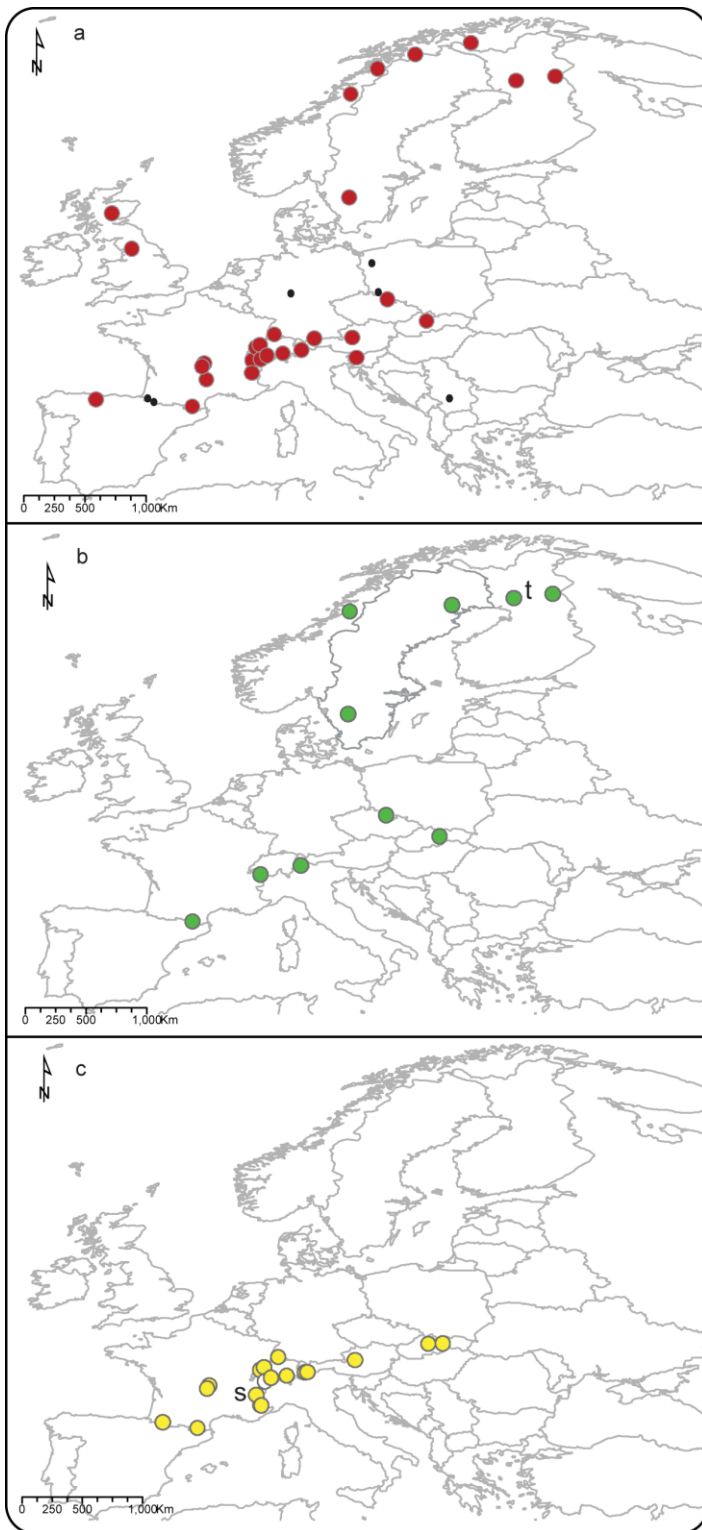
786



788 Figure 1 – Dating and biogeographic reconstruction of twelve species within the genus
789 *Chiastocheta*. Portions in pies indicate ancestral areas assigned by the Dispersion-
790 Extinction-Cladogenesis (DEC) approach. Colors indicate ancestral areas (see figure
791 legend) and map shows biogeographic areas. a – inference considering a substitution rate of
792 2.28%; b - inference considering a substitution rate of 0.9%. Vertical bars delimitate main
793 geological periods. “*” indicates a putative wrongly identified sample in previous studies.
794



796 Figure 2 – Phylogenetic inferences based on (a) nuclear and (b) mitochondrial data and
797 rooted considering relative taxon positions suggested in Figure 1 and Després et al. (2002).
798 Colors of tips indicate morphological species (refer to legend in figure). In a, node supports
799 are shown on branches (ML/Bayesian/MP); in b, main supported clades are shown with an
800 arrow.
801

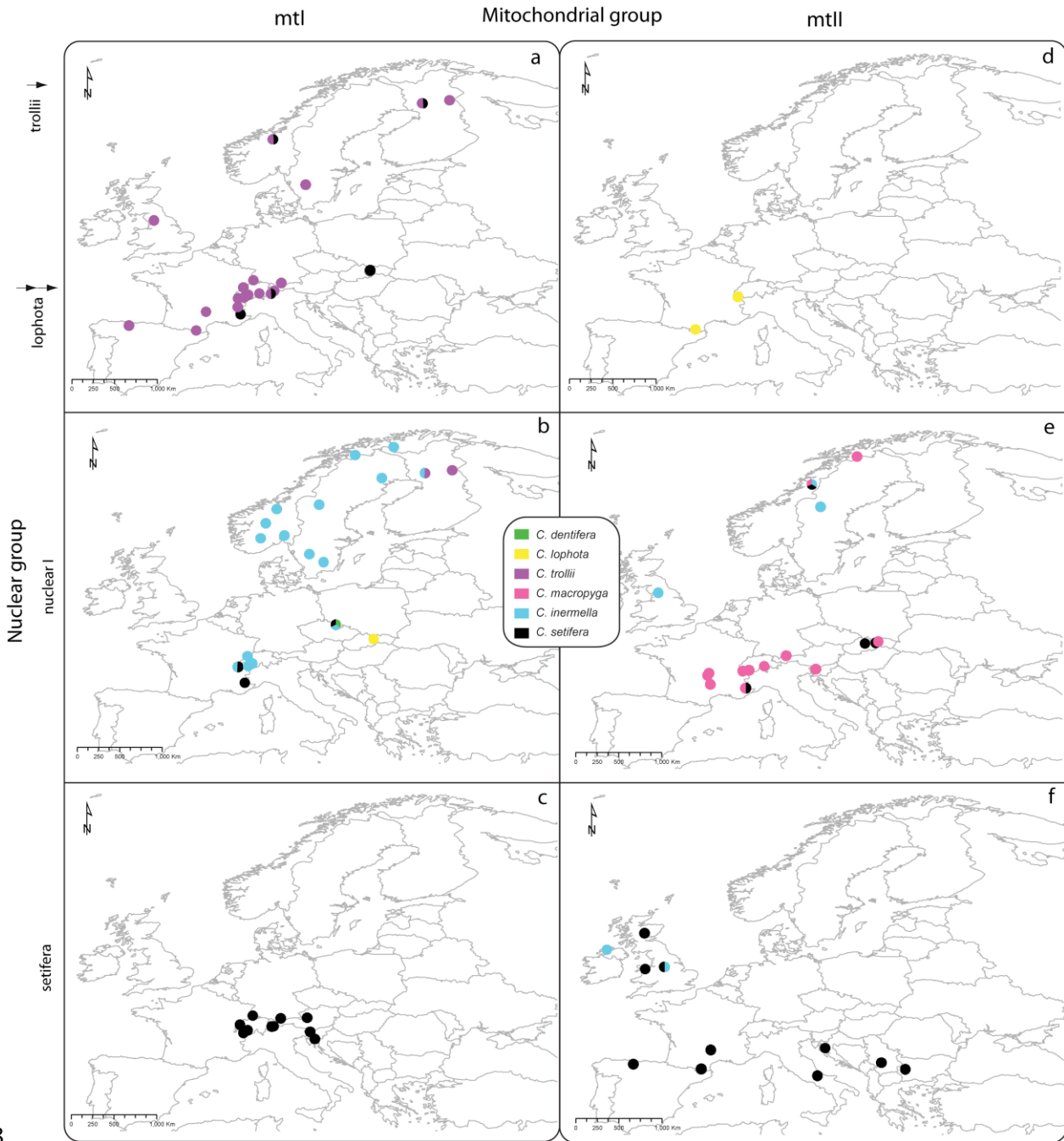


802

803 Figure 3 – Distribution of mitochondrial clades for which congruence between morphological

804 characters and molecular data was observed. a – *C. rotundiventris* , b – *C. dentifera* , c- *C.*805 *lophota*. Letters in b and c represent other exceptional morphospecies identified in the806 clades; t: *C. trollii*; s: *C. setifera*. Black dots in figure (a) indicate sites for which larvae

807 samples fell in the mitochondrial clade.



808

809 Figure 4 – Distribution and morphospecies composition of samples included in mitochondrial

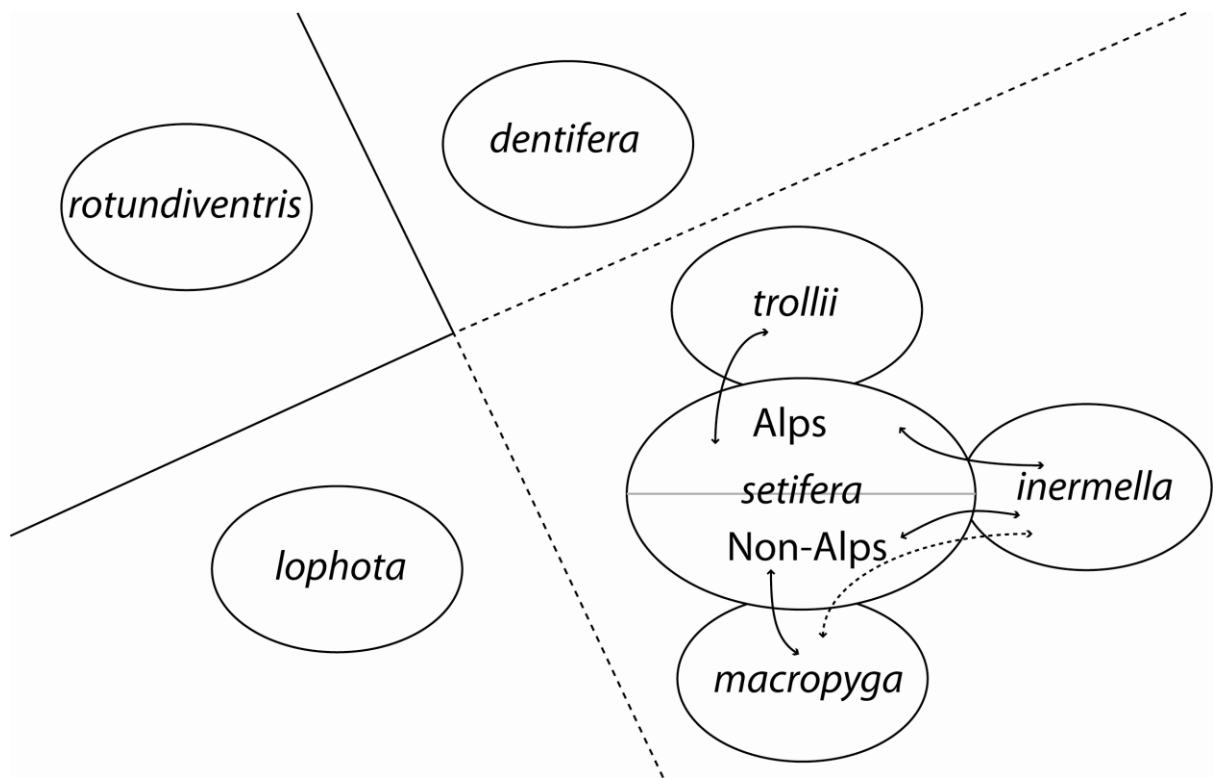
810 groups “mtl” (a-c) and “mtII” (d-f), in respect to nuclear groups in which they were included

811 (a: “trollii”; b and e: “nI”; c and f: “setifera”; d: “lophota”). Colors indicate morphospecies; for

812 explanations refer to the figure legend.

813

814



815

816 Figure 5 – Hybridization scheme in *Chiastocheta* flies pollinating *T. europaeus*. Lines

817 indicate complete isolation; dashed lines indicate important isolation with possible

818 hybridization; arrows indicate hybridizations; dashed arrows show possible indirect genome

819 transfer.

820

821

Supplementary Material 1 – Samples included in the dating and biogeographic analyses

| Name | Sample_ID | Origin | GenBank | Species |
|-----------------|------------|------------------------|------------|--------------------------|
| curvibasis1 | curviba2 | GenBank | AH010127.1 | <i>C. curvibasis</i> |
| curvibasis2 | curvSkr3 | GenBank | AH010133.1 | <i>C. curvibasis</i> |
| Delia | Delia | GenBank | AH010111.1 | <i>Delia brassicae</i> |
| dentifera1 | CHE10md | CH Chemin | This study | <i>C. dentifera</i> |
| dentifera2 | denti3 | GenBank | AH010114.1 | <i>C. dentifera</i> |
| dentifera3 | RAD16fd | Radkow | This study | <i>C. dentifera</i> |
| dentifera4 | SET25fd | Seterasen | This study | <i>C. dentifera</i> |
| dentifera5 | SOL14md | Solberga | This study | <i>C. dentifera</i> |
| inermella1 | AMO8fi | Amot | This study | <i>C. inermella</i> |
| inermella2 | BEI15fi | Beistohlen | This study | <i>C. inermella</i> |
| inermella3 | CHE5fi | CH Chemin | This study | <i>C. inermella</i> |
| inermella4 | inerm3 | GenBank | AH010117.1 | <i>C. inermella</i> |
| inermella5 | inerm4 | GenBank | AH010118.1 | <i>C. inermella</i> |
| inermella6 | inerm5 | GenBank | AH010764.1 | <i>C. inermella</i> |
| inermella7 | inerm6 | GenBank | AH010765.1 | <i>C. inermella</i> |
| inermella8 | SAL23fi | Salla | This study | <i>C. inermella</i> |
| latispinigera1 | latispSKr2 | GenBank | AH010132.1 | <i>C. latispinigera</i> |
| latispinigera2 | latisSKz4 | GenBank | AH010135.1 | <i>C. latispinigera</i> |
| lophota1 | DON3fl | Donovaly | This study | <i>C. lophota</i> |
| lophota2 | HT14fl | Haute Tinee 1 | This study | <i>C. lophota</i> |
| lophota3 | LOS19ml | Loser | This study | <i>C. lophota</i> |
| lophota4 | MOE8ml | Moerlimatt | This study | <i>C. lophota</i> |
| macropyga1 | CHE20mm | CH Chemin | This study | <i>C. macropyga</i> |
| macropyga2 | LAK13fm | Laktatjakka | This study | <i>C. macropyga</i> |
| macropyga3 | macro2 | GenBank | AH010120.1 | <i>C. macropyga</i> |
| macropyga4 | macro4 | GenBank | AH010766.1 | <i>C. macropyga</i> |
| macropyga5 | macrop3 | GenBank | AH010768.1 | <i>C. macropyga</i> |
| macropyga6 | POD1fm | Podlesok | This study | <i>C. macropyga</i> |
| macropyga7 | PYD63fm | Puy de Dome (FR) | This study | <i>C. macropyga</i> |
| pellmyrii1 | pellmy1 | GenBank | AH010769.1 | <i>C. pellmyrii</i> |
| pellmyrii2 | pellmy2 | GenBank | AH010767.1 | <i>C. pellmyrii</i> |
| rotundiventris1 | CHE25mr | CH Chemin | This study | <i>C. rotundiventris</i> |
| rotundiventris2 | LOS7fr | Loser | This study | <i>C. rotundiventris</i> |
| rotundiventris3 | PAN12mr | Puerto de Panderruedas | This study | <i>C. rotundiventris</i> |
| rotundiventris4 | ZAL10mr | Zali Log | This study | <i>C. rotundiventris</i> |
| setifera1 | BAY9fs | Bayasse | This study | <i>C. setifera</i> |
| setifera2 | EID1fs | Eidda Pastures | This study | <i>C. setifera</i> |
| setifera3 | MTP1fs | Monte Pizi | This study | <i>C. setifera</i> |
| setifera4 | setifera | GenBank | AH010123.1 | <i>C. setifera</i> |
| setifera5 | VIT17ms | Vitoshia | This study | <i>C. setifera</i> |
| speciesR3 | spR3 | GenBank | AH010125.1 | species R3 |
| speciesR5 | spR5 | GenBank | AH010131.1 | species R5 |
| trollii1 | EPO11mt | Esposouille | This study | <i>C. trollii</i> |
| trollii2 | GAL36mt | Col du Galibier (FR) | This study | <i>C. trollii</i> |
| trollii3 | MOE7ft | Moerlimatt | This study | <i>C. trollii</i> |
| trollii4 | NAV17ft | Naverdal | This study | <i>C. trollii</i> |
| trollii5 | trollii | GenBank | AH010122.1 | <i>C. trollii</i> |

823 Supplementary Material 2 – Q matrix used for biogeographic reconstructions. A: Central
824 Asia; B: Europe; C: Japan; D: Southeastern Asia. Values indicate probability of dispersion
825 between areas.

| | A | B | C | D |
|----------|----------|----------|----------|----------|
| A | - | 1 | 1 | 1 |
| B | 1 | - | 0.01 | 0.01 |
| C | 1 | 0.01 | - | 1 |
| D | 1 | 0.01 | 1 | - |

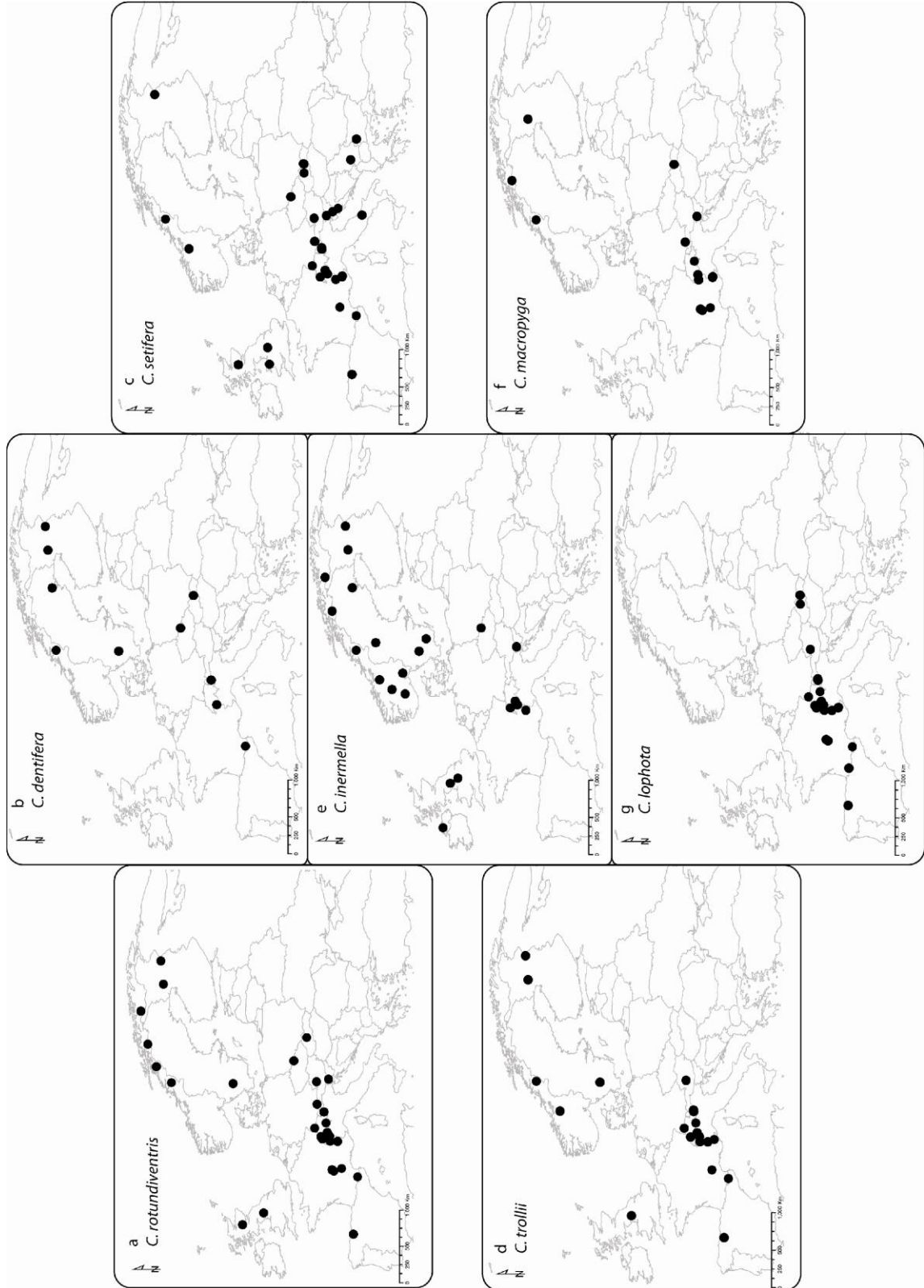
826

827

| Location | Longitude | Latitude | Altitude (m) | Collection date | dentifera | inermella | lophota | macropyga | rotundiventris | setifera | trollii | larvae | Total |
|---------------------------|-----------|-----------|--------------|-----------------|-----------|-----------|---------|-----------|----------------|----------|---------|--------|-------|
| Ambri | 8.702920 | 46.506800 | 1000 | 30.5.08 | 0 | 0 | 2 | 4 | 2 | 0 | 1 | 0 | 9 |
| Amot | 8.42346 | 59.62199 | 481 | 18.06.07 | 0 | 4 | 0 | 0 | 0 | 0 | 0 | 0 | 4 |
| Bayasse | 6.740670 | 44.308140 | N/A | 9.06.07 | 0 | 0 | 2 | 2 | 0 | 2 | 2 | 0 | 8 |
| Beistohlen | 8.95473 | 61.20761 | 729 | 19.06.07 | 0 | 5 | 0 | 0 | 0 | 0 | 0 | 0 | 5 |
| Bidjovagge | 22.478080 | 69.297780 | 613 | 1.8.08 | 0 | 1 | 0 | 0 | 2 | 0 | 0 | 0 | 3 |
| Bobolice | 16.59895 | 53.94660 | 110 | 19.6.08 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 2 | 2 |
| Col de Bonnetcombe | 3.11410 | 44.57557 | 1335 | 21.05.07 | 0 | 0 | 0 | 3 | 1 | 1 | 3 | 0 | 8 |
| Braas | 15.06817 | 57.09309 | 209 | 15.06.07 | 0 | 1 | 0 | 0 | 0 | 0 | 0 | 0 | 1 |
| Col de la Colombière (FR) | 6.469722 | 45.98722 | 1600 | 14.06.06 | 0 | 0 | 1 | 1 | 1 | 0 | 1 | 0 | 4 |
| CH Creux du Van | 6.741193 | 46.93526 | N/A | 29.06.06 | 0 | 1 | 2 | 0 | 2 | 2 | 0 | 0 | 7 |
| CH Chasseral | 7.021299 | 47.12569 | N/A | 29.06.06 | 0 | 0 | 5 | 0 | 5 | 0 | 1 | 0 | 11 |
| CH Chemin | 7.089778 | 46.08993 | N/A | 26.05.06 | 4 | 4 | 1 | 6 | 7 | 3 | 5 | 0 | 30 |
| CH Crans-Montana | 7.538896 | 46.34650 | N/A | 25.05.06 | 0 | 1 | 1 | 0 | 3 | 1 | 1 | 0 | 7 |
| Cressbrook Dale | -1.740410 | 53.267240 | 246 | 10.6.08 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 0 | 2 |
| Colt Park | -2.352470 | 54.193650 | 380 | 10.6.08 | 0 | 4 | 0 | 0 | 1 | 0 | 1 | 0 | 6 |
| Donovaly | 19.230680 | 48.889220 | 1345 | 21.6.08 | 0 | 0 | 3 | 0 | 2 | 1 | 0 | 0 | 6 |
| Eidda Pastures | -3.741900 | 53.037200 | 234 | 9.6.08 | 0 | 0 | 0 | 0 | 0 | 2 | 0 | 0 | 2 |
| Ellingsrudelva | 10.91844 | 59.91771 | 161 | 18.06.07 | 0 | 1 | 0 | 0 | 0 | 0 | 0 | 0 | 1 |
| Esposouille | 2.094500 | 42.623410 | 1521 | 3.6.08 | 1 | 0 | 1 | 0 | 2 | 2 | 3 | 0 | 9 |
| Froson | 14.60268 | 63.18205 | 424 | 20.06.07 | 0 | 2 | 0 | 0 | 0 | 0 | 0 | 0 | 2 |
| Col du Galibier (FR) | 6.438611 | 45.08528 | 2000 | 14.06.06 | 0 | 3 | 3 | 0 | 2 | 3 | 7 | 0 | 18 |
| Glen Fender | -3.794850 | 56.781380 | 345 | 14.6.08 | 0 | 0 | 0 | 0 | 4 | 2 | 0 | 0 | 6 |
| Haute Tinee 1 | 6.818710 | 44.296170 | 2020 | 4.06.07 | 0 | 0 | 3 | 0 | 0 | 0 | 0 | 0 | 3 |
| Haute Tinee 2 | 6.855810 | 44.284260 | 1770 | 4.06.07 | 0 | 0 | 0 | 2 | 0 | 1 | 0 | 0 | 3 |
| Krasno Polje | 14.97271 | 44.808690 | 1487 | 9.7.08 | 0 | 0 | 0 | 0 | 0 | 1 | 0 | 4 | 5 |

| | | | | | | | | | | | | | |
|------------------------|-----------|-----------|------|----------|---|----|---|----|---|---|---|---|----|
| Laktatjakka | 18.40674 | 68.42931 | 453 | 22.06.07 | 0 | 2 | 0 | 10 | 7 | 0 | 0 | 0 | 19 |
| Lough Fern | -7.711300 | 55.065690 | 26 | 13.6.08 | 0 | 1 | 0 | 0 | 0 | 0 | 0 | 0 | 1 |
| Losser | 13.78485 | 47.66052 | 1598 | 04.06.07 | 0 | 0 | 5 | 0 | 4 | 1 | 1 | 0 | 11 |
| Col long de Magnabaigt | -0.43646 | 42.87060 | 1615 | 23.7.08 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 4 | 4 |
| Moerlimatt | 8.07760 | 47.90597 | 939 | 10.06.07 | 0 | 0 | 1 | 0 | 1 | 1 | 1 | 0 | 4 |
| Monte Pizi | 14.167140 | 41.915240 | 1546 | 28.5.08 | 0 | 0 | 0 | 0 | 0 | 3 | 0 | 0 | 3 |
| Naverdal | 10.13002 | 62.70417 | 480 | 19.06.07 | 0 | 10 | 0 | 0 | 0 | 1 | 6 | 0 | 17 |
| Pajino Preslo | 20.819700 | 43.277990 | 1802 | 8.7.08 | 0 | 0 | 0 | 0 | 0 | 2 | 0 | 3 | 5 |
| Puerto de Panderruedas | -4.972230 | 43.127430 | 1227 | 4.6.08 | 0 | 0 | 2 | 0 | 4 | 2 | 1 | 0 | 9 |
| Pila | 20.294490 | 48.900170 | 972 | 21.6.08 | 2 | 0 | 3 | 0 | 0 | 1 | 0 | 0 | 6 |
| Podlesok | 20.351900 | 48.949620 | 579 | 21.6.08 | 0 | 0 | 0 | 1 | 0 | 1 | 0 | 2 | 4 |
| petit papa noel | 25.79386 | 66.51647 | 72 | 23.06.07 | 6 | 4 | 0 | 1 | 4 | 2 | 4 | 0 | 21 |
| Puy de Dome (FR) | 2.963333 | 45.77222 | 1460 | 15.06.06 | 0 | 0 | 2 | 2 | 1 | 0 | 0 | 0 | 5 |
| Puy de Sancy (FR) | 2.809722 | 45.53500 | 1520 | 16.06.06 | 0 | 0 | 6 | 4 | 1 | 0 | 0 | 0 | 11 |
| Radkow | 16.353210 | 50.468660 | 712 | 20.6.08 | 2 | 2 | 0 | 0 | 2 | 2 | 0 | 5 | 13 |
| Risnjak - Snjeznik | 14.584940 | 45.438710 | 1466 | 5.06.08 | 0 | 0 | 0 | 0 | 0 | 2 | 0 | 0 | 2 |
| Salla | 28.65427 | 66.83020 | 194 | 23.06.07 | 7 | 5 | 0 | 0 | 3 | 1 | 6 | 0 | 22 |
| Sede de Pan | -0.486510 | 43.039490 | 1556 | 22.7.08 | 0 | 0 | 2 | 0 | 0 | 0 | 0 | 2 | 4 |
| Seterasen | 13.67744 | 65.53432 | 285 | 21.06.07 | 6 | 7 | 0 | 1 | 1 | 1 | 2 | 0 | 18 |
| Solberga | 13.56116 | 57.95194 | 239 | 15.06.07 | 4 | 4 | 0 | 0 | 3 | 0 | 1 | 0 | 12 |
| Steingaden | 11.01296 | 47.59529 | 1158 | 30.05.07 | 0 | 0 | 0 | 1 | 1 | 6 | 0 | 0 | 8 |
| Straumen | 15.64921 | 67.38440 | 75 | 21.06.07 | 0 | 0 | 0 | 0 | 3 | 0 | 0 | 0 | 3 |
| CH Susch | 10.074725 | 46.747277 | N/A | 05.06.06 | 2 | 0 | 2 | 0 | 1 | 1 | 3 | 0 | 9 |
| Svartla | 21.22062 | 65.99583 | 36 | 22.06.07 | 4 | 3 | 0 | 0 | 0 | 0 | 0 | 0 | 7 |
| CH Tarasp | 10.250556 | 46.777299 | N/A | 05.06.06 | 0 | 0 | 2 | 0 | 0 | 2 | 2 | 0 | 6 |
| Trollblumenwiese | 11.04118 | 51.68314 | 488 | 16.6.08 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 4 | 4 |
| Vitosha | 42.590320 | 23.293420 | 1779 | 3.7.08 | 0 | 0 | 0 | 0 | 0 | 2 | 0 | 0 | 2 |
| Zali Log | 14.110800 | 46.203420 | 533 | 17.5.08 | 0 | 1 | 0 | 3 | 2 | 1 | 0 | 0 | 7 |

829 Supplementary Material 4 –Distribution ranges of morphologically identified (typological)
 830 species. a- *C. rotundiventris*, b- *C. dentifera*, c- *C. setifera*, d- *C. trollii*, e- *C. inermella*, f- *C.*
 831 *macropyga*, g- *C. lophota*.



832

833 *Morphospecies and their distribution ranges*

834 Our study confirms some indications provided by Pellmyr (1992), which proposed
835 distribution ranges for the different *Chiastocheta* species associated with *Trollius europaeus*.
836 While he based his conclusions on egg identifications, we studied adult distributions, and
837 even if some results are compatible with those previously presented, other strongly differ.

838 Different patterns appeared depending on the morphological group analyzed: while some
839 species are widely distributed and cover the whole distribution range (first case), others
840 present a fragmented distribution or are restricted to some regions (second case).

841 In the first case, we found samples belonging to *C. rotundiventris* and to some extent *C.*
842 *dentifera*, *C. trollii*, *C. setifera* and *C. inermella*. The first species (Figure a) presents in our
843 study a similar distribution range than that depicted by Pellmyr in his revision of the *Trollius-*
844 *Chiastocheta* interaction (Pellmyr, 1992). We here also confirm the presence of this taxon in
845 the Eastern Alps and the Tatra region, a zone designed by the last surveys as putative for
846 the presence of flies. This geographic constancy could indicate that this species is also the
847 most reliable for pollination.

848 *C. dentifera* (Fig b) is also present in all regions, but with rarer occurrences than *C.*
849 *rotundiventris* (ten vs. 25 locations, respectively). The distribution range presented by our
850 study shows some congruence with that obtained by Pellmyr (1992), since we did not find
851 any *C. dentifera* in the British Isles, but we did observe them in the Pyrenees. Differences in
852 the presence and absence of *C. dentifera* in the British Isles in Pellmyr's and our study,
853 respectively, could be either due to a recent change in the distribution range of this species
854 in that region –the last records in the zone date back to 1960 (NBN, 2010) and correspond
855 exactly to those presented in Pellmyr's work– or to a random lack of this species in our
856 sampling. This should be however easily tested by new surveys in the area.

857 *C. setifera* (Figure c) also occupies an important part of the sampled area, and is the only
858 present in the Italian Peninsula and Balkan zone. This last point would indicate its ecological
859 importance in these regions, since the lack of other species would design *C. setifera* as the
860 only species available to pollinate flowers. Moreover, our results also extend the distribution

861 range of this species to Scandinavia, a region from which it was completely lacking during
862 the last surveys.

863 Results on the distribution range of *C. trollii* (Figure d) correspond well to distributions
864 obtained based on egg morphology by Pellmyr (1992). However, we additionally show the
865 presence of this species in the Pyrenees and Cantabria.

866 The last species displaying a large distribution is *C. inermella* (Figure e), which appeared
867 in our survey as clearly present in Scandinavia, but also in the Alps, the Tatra and the British
868 Isles. This difference to Pellmyr (1992) is substantial and would tend to reject a previous
869 hypothesis of a vicariant identity between this typological species and *C. lophota* (its
870 supposed Southern vicariant; Pellmyr, 1992).

871 Only *C. macropyga* (Figure f) appeared to present a fragmented distribution range. The
872 species is present in the Alpine chain, the French Massif Central and to some extent the
873 Tatra region. However, there are no records of the species between these zones and
874 Northern Scandinavia, the other region where it has been observed. This distribution
875 confirms the view proposed by Pellmyr (1992).

876 Finally, the only species appearing restricted to only some regions is *C. lophota* (Figure
877 g), which is mentioned by Pellmyr (1992) as the Southern vicariant of *C. inermella* (despite
878 our results contradict this view). Our survey shows that the species is thus only present in
879 the Southern mountainous chains of Europe.

880 *References*

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882 *Trollius* and its seed-parasitic pollinators. *Biological Journal of the Linnean Society*,
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885

Chapter seven

*No fossils, no answers?
Inferring postglacial genetic consequences in cold-
adapted species when neither ancient DNA nor
macroremains are available*

Anahí Espíndola, Loïc Pellissier and Nadir Alvarez

1 Article type: Original article

2 Title: No fossils, no answers? Inferring postglacial genetic
3 consequences in cold-adapted species when neither ancient DNA
4 nor macroremains are available

5 Authors: Espíndola Anahí¹*, Pellissier Loïc² and Alvarez Nadir²

6 ¹Laboratory of Evolutionary Entomology, Institute of Biology, University of Neuchâtel. Emile-Argand
7 11, 2009 Neuchâtel, Switzerland.

8 ²Department of Ecology and Evolution, Biophore Building, University of Lausanne, 1015 Lausanne,
9 Switzerland.

10

11 * Correspondence: Anahí Espíndola, Laboratory of Evolutionary Entomology, Institute of Biology,
12 University of Neuchâtel. Emile-Argand 11, 2009 Neuchâtel, Switzerland. Fax: +41 (0)32 718 3001.

13 Email: maria.espindola@unine.ch

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15 Running head: Postglacial contraction in cold-adapted taxa

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28 **ABSTRACT**

29 Aim: Unlike temperate species, whose distribution ranges have expanded since the end of
30 the Last Glacial Maximum (LGM), cold-adapted species have experienced substantial
31 postglacial range contractions. To understand the dynamics of their contraction process,
32 researchers have proposed to confront current phylogeographic patterns with past genetic
33 and/or distribution data, inferred from ancient material. However, because fossils and/or
34 macroremains are unavailable for most species, direct inferences on postglacial histories are
35 only possible for a restricted number of taxa. In this study we present a novel approach to fill
36 this technical gap and to open-up new possibilities for inferring the postglacial fate of
37 organisms for which no ancient material is available.

38 Location: West-Palearctic.

39 Methods: Using *Trollius europaeus* (Ranunculaceae) as a case study, we sampled the
40 whole European range of the plant and further inferred its current spatial genetic structure
41 based on AFLP data. Simultaneously, we defined the Ecological Niche Model (ENM) of the
42 species and used it to hindcast its LGM distribution. Based on this, we identified past suitable
43 areas (suitability hotspots), which could have potentially hosted large numbers of individuals
44 and specific genetic lineages at the LGM, and compared their distribution with the current
45 spatial genetic structure of the plant. Because isolation of these hotspots might have driven
46 lineage divergence at LGM, this comparative approach provides a probabilistic view of the
47 distribution of past genetic diversities. We thus propose that the level of congruence between
48 the number and spatial distribution of current genetic clusters and hindcasted LGM hotspots
49 is a good proxy for informing on the genetic diversity loss induced by the important range
50 contractions experienced after the LGM.

51 Results: The current genetic structure of the plant is spatially divided in four well-defined
52 clusters, which correspond to four out of the five hindcasted LGM suitability hotspots.
53 Assuming that lineages were not shared between hotspots, this discrepancy informs on a
54 probable post-glacial loss of one genetic lineage in the species.

55 Main conclusions: Our approach provides a valuable probabilistic view on the genetic
56 consequences of the last postglacial contractions in cold-adapted species, especially those
57 for which other inferences are impossible because of the absence of ancient material.

58

59 **Keywords**

60 hindcasting, Ecological Niche Modelling, *Trollius europaeus*, spatial genetic structure,
61 postglacial contraction, phylogeography, arctic-alpine species

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64 **INTRODUCTION**

65 Species distributions are constrained both by physiological limitations related to abiotic
66 factors (e.g., temperature, moisture) and by the effect of biotic interactions in which they are
67 involved (e.g., competition, parasitism; Pulliam, 2000). As a consequence, species are only
68 able to occupy the subset of the available environment where their requirements are fulfilled,
69 a concept referred to as the ecological niche by Hutchinson (1957). The spatial distribution of
70 species is thus both mainly defined by their respective ecological niche, their dispersal
71 characteristics and their colonization histories (Alvarez *et al.*, 2009). Therefore, any
72 environmental change has the potential to induce large variations in the area occupied by a
73 species, and to consequently influence other characteristics of the studied organism, as for
74 instance, demographic and genetic parameters at the population level. If the distribution
75 range of a species becomes fragmented and gene flow among populations is limited or
76 absent, evolutionary processes (e.g., vicariance, drift, selection) are expected to drive
77 differences in adaptive and neutral genetic variation among isolated patches of populations
78 (Hewitt, 1999). Hence, the genetic variation within a species is not only defined by intrinsic
79 biological characteristics such as its reproductive strategy or its generation time, but also by
80 extrinsic factors such as climate, topography and habitat availability (Avisé, 2009).

81 Because of the interaction between orbital and tectonic forces, the Quaternary has been
82 marked by cyclic climatic oscillations, defined by successive glacial and interglacial periods
83 that have modified species' habitat availability through space and time (Hewitt, 1996). During
84 the Last Glacial Maximum (LGM: 23 -18 thousand years ago -kya) most of northern Europe
85 was covered by polar ice caps and glaciers filled internal valleys of southern European
86 mountain ranges. Moreover, because of the large volume of water contained in the ice caps,
87 sea levels lowered and new lands became exposed, especially in the English Channel and
88 the Adriatic Sea (Ehlers & Gibbard, 2007). Consequently, biomes were displaced southwards
89 and cold steppic environments could be found as far South as southern France, the Jura and
90 southern Carpathians (Frenzel, 1968; Hewitt, 1999). While the main part of the continent
91 presented suitable habitats for cold-adapted species (i.e., currently arctic, alpine or arctic-

92 alpine), southern regions harboured conditions more favourable to temperate organisms.
93 This was mainly due to the proximity of the Mediterranean Sea that provided humidity and
94 higher temperatures to surrounding lands, but also to the topography of southern mountains
95 that offered a wide range of climatically protected habitats (Médail & Diadema, 2009; Stewart
96 *et al.*, 2010). Demographic and distributional responses to climatic variations are expected to
97 differ between temperate and cold-adapted species (reviewed in Stewart *et al.*, 2010): while
98 temperate species are expected to be widespread during interglacial and restricted to refugia
99 during glacial periods, cold-adapted species reflect the opposite demographic pattern.

100 For the last two decades, the biogeography of temperate species has been widely studied
101 in the western Palearctic and phylogeographic patterns have been summarized into a few
102 paradigms as the ones represented by the European beech *Fagus sylvatica*, the hedgehog
103 *Erinaceus* and the bear *Ursus arctos* (e.g., Taberlet *et al.*, 1998; Hewitt, 1999; Schmitt,
104 2007). In contrast, probably because of more difficult sampling strategies associated to their
105 arctic-alpine distributions, cold-adapted species received less attention and, compared to
106 temperate species, only a few of their corresponding phylogeographic patterns were
107 unravelled (Brochmann *et al.*, 2003; Schönswetter *et al.*, 2006; Ehrich *et al.*, 2007). Whereas
108 investigating the spatial genetic structure of temperate organisms (currently showing an
109 expanded distribution) provides information on the genetic consequences of postglacial
110 expansion processes, studying the phylogeography of cold-adapted species, which are
111 currently found under refugial conditions (*i.e.* at high altitude and/or latitude areas), allows
112 scenario testing on the genetic consequences of distributional contractions. Understanding
113 consequences of such contractions is not only important for historical reasons, it also
114 provides information for predicting the fate of species expected to be negatively affected by
115 the current and future global climate warming (Hampe & Petit, 2005).

116 Direct inference on the variation and temporal maintenance of the genetic diversity
117 associated with previously expanded cold-adapted lineages is currently only possible when
118 ancient DNA is available (Stewart *et al.*, 2010). For instance, Dalén *et al.* (2007) have shown
119 that in the specific case of the polar fox, several ancient lineages – tracked back by the

120 analysis of ancient DNA – have nowadays become extinct and their respective alleles are
121 absent from the current species' distribution. Even though such an approach brings a
122 substantial contribution to our knowledge on the genetic consequences of climatic
123 oscillations, it is restricted to the study of taxa for which ancient DNA or macroremains are
124 accessible. Because there is a lack of available ancient DNA in some cold-adapted species,
125 retracing the evolution of ancient lineages in organisms with current arctic or alpine
126 distributions is thus limited to indirect inference.

127 In the present work, we propose an indirect technique to infer the fate of Quaternary
128 lineages when ancient DNA or fossils are not available. By combining Ecological Niche
129 Modelling (ENM) and further hindcasted climatic probabilities of occurrence, with an
130 estimation of the current spatial genetic structure of a given organism, our approach allows
131 inferring the probable contraction dynamics of species after the end of the European LGM.
132 Based on a solid phylogeographic paradigm (Avise *et al.*, 1987; Avise, 2009), we suggest
133 that hotspot areas showing high hindcasted probabilities of past suitability are correlated with
134 a high probability of past occurrence of one or several endemic genotype(s) in that region.
135 We thus aim to provide a probabilistic sight on the past genetic diversity of a species. Here
136 we propose first to identify ENM-based *suitability hotspots*, defined as well-circumscribed
137 regions showing high climatic suitability for a given species at the LGM. Second, based on a
138 sampling covering a substantial portion of the current species distribution and applying
139 genome fingerprinting techniques, we evaluate its present genetic structure. Finally, we
140 combine these two approaches and test the following hypotheses: if the number and
141 localization of LGM *suitability hotspots* correspond to the number and distribution of the
142 current genetic lineages identified, it is likely that at least one sample per hotspot was
143 preserved after the end of the last glacial period and contributed to the current genetic
144 variation of the species; alternatively, if this was not the case, we can consider that the range
145 contraction might have induced a possible loss of genetic diversity (Dalén *et al.*, 2007).

146 In order to test these hypotheses, we use the globeflower *Trollius europaeus* L.
147 (Ranunculaceae) as a case study. This Eurasian arctic-alpine species is currently associated

148 with cold and humid habitats and covers the southern mountainous ranges of Europe as well
149 as some northeastern European plains, extending up to northern Scandinavia and the Ural
150 (Meusel *et al.*, 1965). As all arctic-alpine species, *T. europaeus* is considered to be currently
151 in a retracted demographical situation and its distribution during the LGM is expected to have
152 been more extensive than today (Stewart *et al.*, 2010). Based on a thorough sampling
153 covering the whole European range of the plant distribution, we evaluate the amount of past
154 genetic diversity that has been conserved in this species following the distributional
155 contraction experienced at the beginning of the current interglacial period.

156

157 MATERIAL AND METHODS

158 Genetic data and analyses

159 *Trollius europaeus* samples were collected during summers 2006, 2007 and 2008 in 79
160 populations covering the whole European range of the plant (Figure 1, Appendix S1 in
161 Supporting Information). Plant tissue from up to six individuals per population were
162 conserved in silica gel and DNA was extracted using the standard protocol of the QIAGEN
163 DNeasy 96 Plant kit (QIAGEN, Hombrechtikon, Switzerland). amplified fragment length
164 polymorphism (AFLP; Vos *et al.*, 1995) was performed with *EcoRI* and *MseI* endonucleases
165 (Promega, Dübendorf, Switzerland) and further amplified with the two primer pairs E-ATC/M-
166 CAT and E-ACA/M-CTG. For digestion, 5µl of DNA were added to a final volume of 20 µl,
167 containing 1x Buffer 2, 0.1mg/ml BSA and 1 unit/µl of *MseI* and *EcoRI*; the mix was
168 incubated at 37°C for 2h. Adaptors were ligated to the digested products at 37°C for 2h,
169 using 40µl of mix containing 1X Buffer T4, 0.45µM adaptor E, 0.36µM adaptor M, 0.015
170 units/µl T4 ligase (Promega, Dübendorf, Switzerland) and 20µl of digestion product.
171 Preselective PCR was run in a 20µl mix composed of 1X buffer GoTaq (Promega SA,
172 Dübendorf, Switzerland), 2µM MgCl₂ (Promega, Dübendorf, Switzerland), 250 µM dNTPs
173 (Promega, Dübendorf, Switzerland), 0.25µM primers E-A and M-C, 0.025 units/µl of Go-Taq
174 polymerase (Promega, Dübendorf, Switzerland) and 2µl ligated DNA. Thermocycler
175 conditions were 2min at 94°C, followed by 29 cycles of 45 s at 94°C, 45 s at 56°C and 1 min

176 at 72°C, with 10 min of final extension at 72°C. The final selective step was done in 20µl mix
177 composed of 1X buffer GoTaq (Promega, Dübendorf, Switzerland), 2µM MgCl₂ (Promega
178 SA, Dübendorf, Switzerland), 250 µM dNTPs (Promega, Dübendorf, Switzerland), 0.4µM of
179 selective primers E-ACA/M-CTG and E-ATC/M-CAT, 0.025 units/µl of Go-Taq polymerase
180 and 3µl of a solution 1/20 of the preselective PCR product. Thermocycler conditions were 2
181 min at 94°C, followed by 13 cycles of 30 s at 94°C, 30 s at 65°C with temperature lowering
182 0.7°C at each cycle and 1 min at 72°C, followed by 23 cycles of 30 s at 94°C, 30 s at 65°C
183 and 1 min at 72°C, and 5 min of final extension at 72°C. Selective amplification products
184 were run by Macrogen Inc. (Seoul, South Korea) in a denaturing polyacrylamide gel with an
185 internal size standard (ROX 400HD) on an automated DNA sequencer (ABI 377; Applied
186 Biosystems, Foster City, USA). Automated allele-calling was further performed using
187 Genemapper 3.7 (Applied Biosystems, Foster City, USA) with the following parameters: RFU
188 threshold fixed to 50; binning range between 50bp and 270bp for E-ACA/M-CTG and 50bp
189 and 350bp for E-ATC/M-CAT. Because digestion and amplification problems can arise during
190 the procedure and can bias results (Arrigo *et al.*, 2009), we randomly chose 36 individuals
191 per plate to be used as intra-plate replicates, while five samples out of the total were selected
192 as inter-plate replicates. To test for allele reproducibility, we analyzed the percentage of
193 bands shared by all replicates of a given sample.

194 In order to identify the genetic structure of the dataset, ten Monte-Carlo Markov chain
195 (MCMC) runs of 1,000,000 generations with a 200,000 burnin period were run for an *a priori*
196 number of genetic clusters (hereafter referred to as *K*) ranging from one to 20 (200 runs in
197 total) in a model-based Bayesian framework implemented in STRUCTURE 2.2 (Falush *et al.*,
198 2007). Following recommendations of D. Falush (pers. comm.), data was coded as diploid
199 and recessive alleles as present. The most probable *K* was identified using approaches
200 proposed both by Pritchard *et al.* (2000) and Evanno *et al.* (2005).

201 In parallel and in order to confirm the genetic structure, a non-model-based approach was
202 applied using the K-means clustering technique as introduced by Hartigan and Wong (1979)
203 and as recently implemented in an AFLP-based phylogeographic framework by several

204 studies focusing on European plants (Burnier *et al.*, 2009; Arrigo *et al.*, 2010). Following
205 Kergoat and Alvarez (2008), we identified the number of clusters (K) that optimizes the
206 inertia (*i.e.*, the average distance between the centroid of each cluster and the samples
207 assigned to each of them) of the dataset, and that best represents its most parsimonious
208 splitting; the best K is identified by the analysis of the variation of inertias and of their first
209 derivatives for each value of K . Here, inertias and assignments were calculated for K ranging
210 from one to 20. Calculations were repeated starting at different random points 10,000 times,
211 and run in the R 9.2.1 CRAN environment (R Core Development Group, 2009). Scripts are
212 available upon request to the first author.

213 Nei genetic diversities (Nei & Li, 1979) were calculated based on samples having
214 successfully amplified fragments using both AFLP primer-pairs, by applying a geographic
215 moving window (Arrigo *et al.*, 2010) in a R 9.2.1 CRAN environment (R Core Development
216 Group, 2009). The method consists of the following steps: i) the region of study is defined
217 with maximum longitude and latitude values; ii) a grid of fixed cell-size (in our case 75 km)
218 and their associated centroids are calculated; iii) a fixed maximum number of samples (four
219 in our analysis) is randomly selected in a user-defined centroid perimeter (200 km in our
220 case); iv) Nei's diversity indices are calculated for each centroid, v) points iii and iv are
221 repeated 1,000 times, vi) mean values of the 1,000 Nei's indices are calculated for each
222 centroid. This method allows not only to calculate the Nei diversity values at each point, but
223 also to observe the index variation and distribution throughout the area of study. Moreover, it
224 allows for a correction of possible local sampling biases since the analysis is grid-based,
225 instead of sampled-population based. Scripts are available upon request to the first author.

226 Finally, to confirm the genetic structure obtained, a distance-based phylogenetic analysis
227 was applied based on the calculation of modified Nei-Li pairwise distances between
228 genotyped individuals, as implemented in PHYLIP ("restdist" and "neighbor" programs;
229 Felsenstein, 2005). The resulting topology was rooted using the mid-point rooting method, as
230 implemented in FigTree 1.2.2 (<http://tree.bio.ed.ac.uk/software/figtree>).

231

Environmental analyses*Ecological Niche Modelling (ENM) and hindcasting*

Occurrences of *T. europaeus* recorded with a spatial accuracy $\leq 5\text{km}$ were extracted from seven biodiversity databases (GBIF: www.gbif.org; Flora Croatica: hirc.botanic.hr/fcd/Search.aspx; Swedish Artportalen: <http://www.artportalen.se>; database of the Botanical Alpine Conservatory of Gap-Charance, France; Anthos Project: www.anthos.es; NBN gateway: www.searchnbn.net/index_homepage/index.jsp; Centre du Réseau Suisse de Floristique www.crsf.ch) with a total of 49,863 occurrences. The accuracy threshold has been applied taking into account that 5km is also the approximate resolution of the most detailed climatic grids currently available (see below). This dataset was pooled with occurrences collected during fieldwork (79 locations throughout the distribution range, see above). Because the total 49,932 occurrences were highly aggregated in some regions of Europe (mostly western Europe), we avoided oversampling by defining a minimal distance between points (*i.e.*, 20 km) and randomly selected a subset of occurrences spatially distributed across the range of the species using this buffer.

Most modeling techniques require not only information on presences but also on absences or pseudo-absences when no true absences are available (Wisz & Guisan, 2009). Therefore, 10,000 pseudo-absence points were selected based on a random selection of pixels with a resolution equal to that of the climatic layers (2.5 arc-minutes). Because the databases used in the present study provided very good information on occurrences of the plant in the western and central parts of the sampling area, but not in most eastern parts, pseudo-absences were only defined within the limits of the investigated range (Figure 1) and not within the whole distribution of the plant, extending towards the Russian Urals. This was done to avoid biases in the response curves of the models (Phillips *et al.*, 2009).

Because biases in the ENM definition can arise when too many correlated climatic variables are considered (Nogués-Bravo, 2009), we selected the following seven WorldClim bioclimatic layers (Hijmans *et al.*, 2005), which are recognized to be of strong importance for the ecology of alpine and arctic plants (Guisan & Zimmermann, 2000; Körner, 2003):

260 minimum temperature (bio5); maximum temperature (bio6); average temperature of the
261 coldest quarter (bio10); average temperature of the warmest quarter (bio11); precipitation
262 seasonality (bio15); average precipitation of the coldest quarter (bio18); and average
263 precipitation of the warmest quarter (bio19). The distribution of *T. europaeus* was modeled
264 using eight of the modeling techniques proposed by the BIOMOD tool (Thuiller *et al.*, 2009),
265 implemented in the R 9.2.1 CRAN environment (R Core Development Group, 2009): (1)
266 generalized linear model (GLM), (2) generalized additive model (GAM), (3) classification tree
267 analysis (CTA), (4) artificial neural networks (ANN), (5) multivariate adaptive regression
268 splines (MARS), (6) mixture discriminant analysis (MDA) (7) generalized boosting model
269 (GBM), and (8) random forest (RF).

270 In order to evaluate the predictive performance of the species distribution model, we used
271 a random subset of 70% of the data to calibrate each model, while the remaining 30% was
272 used for its assessment (following, Thuiller *et al.*, 2009). Models were evaluated using a
273 Relative Operating Characteristic (ROC) curve and the Area Under the Curve (AUC)
274 (Fielding & Bell, 1997). We replicated 50 times the data splitting and calculated the average
275 AUC of the repeated split-sample, which gives a more robust estimate of the predictive
276 performance of each model (Thuiller *et al.*, 2009).

277 Finally, each model was projected into the past with the WorldClim data of the Community
278 Climate System Model (CCSM) circulation model for the last glacial maximum (LGM; -21,000
279 years) downscaled at a resolution of 2.5 arc-minute. Because ensemble forecasting
280 approaches significantly improve the accuracy of species distribution models (Marmion *et al.*,
281 2009), we applied a weighted average of the eight modeling techniques based on their
282 predictive power (AUC). We thus obtained the main tendency of these distributions,
283 accounting also for possible variation among modeling techniques.

284

285 *Circumscription of regions of high hindcasted probabilities*

286 ENM predicts habitat suitability (in our case based on climatic factors) of the study area
287 for the modeled organism by attributing a probability value to each pixel. Since we aim at

288 delimitating regions of high suitability, it is necessary to define a threshold to retain only
289 those regions in which the probability of suitability for long-term persistence is high. In order
290 to do so, several methods have been previously proposed (Liu *et al.*, 2005; Jiménez-
291 Valverde & Lobos, 2007). Some of these are based on solid statistical backgrounds and are
292 useful when applied to pure theoretical modeling approaches, while others present a deeper
293 biological relevance, more compatible with an evolutionary framework. This is why in the
294 present study we applied two different approaches. First, the one proposed by Manel *et al*
295 (2001), which considers both presences and absences and defines a threshold optimizing
296 sensitivity/specificity ratios (referred to as “SS”). Second, taking into account only the
297 variation in presence data, we extracted the current probability values predicted by the model
298 for these points, and calculated their mean and standard error (SE). In this method (referred
299 to as “M”), the threshold value corresponded to “mean - SE”. With both “SS” and “M”
300 methods, regions falling below thresholds were considered as marginal habitats.

301 Once cut-offs were applied, areas with high hindcasted probabilities, hereafter referred to
302 as *suitability hotspots*, were defined according to the following rules: i) their surface is large
303 and continuous (*i.e.*, we did not consider punctual regions of high probability if they occupied
304 areas smaller than 1,000 km²); ii) their distribution range is isolated from others hotspots (*i.e.*,
305 by boundaries composed of large regions of low-hindcasted probabilities and/or by the
306 presence of ice-sheets).

307

308 **Inference of the post-glacial history**

309 In order to identify an eventual post-glacial loss of diversity, we applied a strategy similar
310 to those proposed by Magri *et al.* (2007) and Cheddadi *et al.* (2006), in terms of a
311 complementary use of non-molecular data. In our case, the inference method replaced
312 inferences on fossil data by an ENM approach, which allows extending the application range
313 of the technique to virtually any cold-adapted species. We compared the number and
314 distributions of current genetic clusters with those of *suitability hotspots* defined from the
315 ENM approach, using the following inference key: i) if their numbers and respective

316 distributions were similar, the probability is high that at least one genetic lineage per
317 *suitability hotspot* was preserved after the last postglacial contraction; ii) if the number of
318 *suitability hotspots* was higher than the number of genetic groups, this would indicate,
319 assuming that lineages were not shared between hotspots, a probable genetic loss due to
320 postglacial contraction; iii) if the number of genetic groups was higher than the number of
321 *suitability hotspots*, this would indicate that lineages from each ENM *suitability hotspots*
322 participated to the current diversity and that at least one of them harbored more than one
323 single lineage.

324

325 **RESULTS**

326 **Genetic analyses**

327 Among the 354 extracted samples of *T. europaeus*, 270 samples were successfully
328 genotyped with the two primer pairs (374 AFLP fragments in total). In addition, 15 and 64
329 samples amplified correctly only for a single primer pair. Mismatch error rate (*sensu* Bonin *et*
330 *al.* 2004) was 0.90% for primer-pair E-ACA/M-CTG and 0.91% for primer-pair E-ATC/M-CAT.

331 The spatial genetic structure inferred using STRUCTURE showed that for values of $K \geq 4$,
332 the number of “realized” clusters (*i.e.*, comprising maximum probabilities higher than 0.005)
333 was always equal to four and the assignment of samples to each group was identical in all
334 cases. According to Pritchard *et al.* (2000), there is thus a strong structuring of the data into
335 four clusters. Following Evanno *et al.* (2005), the most likely K value was nine, whose best
336 run yielded four “realized” clusters and five “ghost” clusters (*i.e.*, comprising maximum
337 probabilities lower than 0.005). The assignment of samples to the four clusters was strongly
338 structured geographically (Figure 1a): the first cluster was exclusive to south-eastern Europe
339 (blue); the second was only present in northern Scandinavia (green); the third was mainly
340 restricted to south-western Europe (red); and the fourth was more widespread with a
341 central/northwestern European distribution (yellow). At the edges of each cluster distribution,
342 some admixture was observed.

343 The non-hierarchical, non model-based, K-means analysis also indicated that the *K* value
344 that optimized the distribution of inertias was four (Appendix S2 in Supporting Information).
345 When the assignment of samples to each genetic group was analyzed, results were almost
346 identical to those obtained in the model-based STRUCTURE analysis: significant positive
347 correlations between both methods for the assignment of samples to different clusters were
348 illustrated by R^2 values and slopes ranging from 0.84 to 0.96, and from 0.71 to 0.90, for each
349 *K* cluster (Appendix S3 in Supporting Information).

350 The geographic structure of the Nei diversity indices showed a main centre of diversity in
351 the southernmost regions (Alps), with additional highly diverse areas observed in northern
352 Carpathians, central Scandinavia and the North of the British Islands (Figure 1b). In contrast,
353 there seemed to be a loss of genetic diversity in northern continental Europe and southern
354 Scandinavia.

355 The phylogeny inferred from pairwise AFLP-based Nei-Li distances confirmed the
356 existence of the four genetic groups previously identified by the clustering approaches
357 (Figure 2), although their monophyly was not strictly retrieved. While south-western samples
358 were monophyletic and segregated basally, samples from central/north-western and south-
359 eastern Europe were paraphyletic, the later including a nested cluster comprising
360 monophyletic northern Scandinavian samples.

361

362 **ENM and hindcasting**

363 AUC values ranged between 0.772 and 0.847 (Table 1) indicating that the models were
364 performing well (i.e., >0.75 ; Swets, 1988). Current and past climatic suitability showed
365 substantial differences in their geographic extents (Figure 1c-d). The projection of the model
366 for the current environment (Figure 1d) indicated high probability of occurrence in southern
367 and central European mountain ranges (Pyrenees, Apennines, Balkans, Carpathians, Alps),
368 as well as at lower altitudes in northern Europe (Scandinavia, North of the British Islands). It
369 also predicted high probabilities in the north-eastern European wetlands (Baltic region) and
370 extended further East. The suitable area highlighted by the model fitted well with the actual

371 distribution of the plant (Meusel *et al.*, 1965). Hindcasted probabilities (Figure 1c) indicated
372 that a large extent of the land exposed during the LGM showed high suitability for *T.*
373 *europaeus*.

374 The two cut-off methods (“SS” and “M”) applied provided similar results, even though “M”
375 appeared as more restrictive than “SS”, with threshold values of 0.578 and 0.494,
376 respectively. After the application of the cut-off and identification of centres of high
377 probabilities at LGM, five *suitability hotspots* were observed (Figure 1c): i) western Pyrenees;
378 ii) north-eastern Europe, iii) western Europe, with a particularly high probability area in the
379 West (method “SS”); iv) central Carpathian lowlands; v) Northern Balkans with a probable
380 connection with the Apennines. In contrast, regions showing low occurrence probabilities
381 were mainly present in the northern Carpathians and surrounding areas, in central France
382 and across the southern tips of the Mediterranean peninsulas.

383

384 **Congruence between LGM *suitability hotspots* and current spatial genetic structure**

385 The combined evaluation of hindcasted probabilistic hotspots and current spatial genetic
386 structure showed that the number and localization of current genetic groups roughly
387 corresponded to regions of high hindcasted probabilities. We observed however, that there
388 was one additional hindcasted hotspot (five), compared to the number of genetic clusters
389 (four): the genetic cluster covering southeastern Europe (blue, Figure 1a) covered the areas
390 of the two hotspots *iv* and *v* (Figure 1c). In contrast, each of the three other clusters
391 corresponded to one single hotspot: the northern Scandinavian genetic group (green, Figure
392 1a) occupied a region covered with ice-sheets during the LGM, but placed in the
393 neighbourhood of the hindcasted hotspot *ii*, located in the direct southeastern vicinity of the
394 glacial ice-sheet (Figure 1c); the southwestern clade (red, Figure 1a) largely overlapped with
395 the location of hotspot *i* (Figure 1c); finally, the central/northwestern European clade (yellow,
396 in Figure 1a) showed a position similar to that of hotspot *iii* (Figure 1c).

397 At least one of the two hindcasted *suitable hotspots* *iv* or *v* (Figure 1c) seemed to have not
398 participated to the composition of the current genetic diversity of the species in the sampled

399 area, unless these two neighbouring areas shared identical lineages during LGM (see
400 Discussion).

401

402 **DISCUSSION**

403 **Spatial genetic structure**

404 The spatial genetic structure of *T. europaeus* identified in our study gives a much more
405 complete insight into its European phylogeographic pattern when compared to a previous
406 study on this plant (Després *et al.*, 2002), which comprised a much more scattered sampling
407 (18 sampling locations vs. 79 in the present survey). The major novelties and corrections
408 addressed here are (i) the presence of a cluster specific to southeastern Europe (blue,
409 Figure 1a), (ii) the admixed genetic identity of several locations in the southern Alps and
410 eastern Pyrenees, and (iii) the existence of an exclusive and independent northern
411 Scandinavian cluster (green, Figure 1a), different from the cluster found in southern
412 Scandinavia, the Carpathians and northern Poland (yellow, Figure 1a). Our results represent
413 the first evidence in arctic-alpine plants that Scandinavia was colonized twice by two distinct
414 lineages that established a contact zone at mid latitudes similar to the phylogeographic
415 pattern demonstrated for the bear *Ursus arctos* in this region (Hewitt, 1999), but contrasting
416 with results obtained in other cold-adapted species (in contrast to Schönswetter *et al.*, 2006;
417 Skrede *et al.*, 2006; Schmitt, 2009).

418 Overall, the spatial genetic structure inferred in this study shows a well circumscribed
419 geographic organization of the genetic variation whatever approach is considered (Figures
420 1a and 2). The phylogenetic analysis (Figure 2) provides results highly congruent with those
421 obtained in the clustering analysis (Figure 1a). Although the Nei-Li distance-based analysis
422 lacks sensitivity, notably when inferring branch lengths (Felsenstein, 2004), the mid-point
423 rooted topology is compatible with an organization of the genetic diversity into four genetic
424 groups. Nevertheless, only samples from the south-western region (red) were retrieved as
425 monophyletic and basally segregated from the other samples. Monophyletic North
426 Scandinavian samples (green) are nested within samples from the Carpathians/Balkans

427 region (blue). Similarly, samples from this latter area are nested within samples from central
428 Europe and southern Scandinavia (yellow). The paraphyletic and nested nature of these
429 groups might, however, be due to methodological biases related to distance calculations
430 based on AFLP data.

431 Extrapolated grid-based Nei genetic diversities (Figure 1b) indicate that regions of high
432 diversity are mainly distributed in the southern part of the distribution, especially in the Alpine
433 arc (Figure 1b). The high diversity retrieved in southern massifs might be explained by the
434 potential of these massifs to play a role of long-term refugia during interglacial periods (*sensu*
435 Stewart *et al.*, 2010). The high level of diversity found all over the Alpine arc might also result
436 from the presence of ancient (*i.e.*, pre-LGM) glacial contact zones that could have been
437 “preserved” today in a refugial state (*in situ* survival, *sensu* Brochmann *et al.*, 2003).
438 Alternatively, it could also reflect a colonization of this central mountain range by several
439 extra-Alpine contracting genetic groups after the retreat of the ice-sheets which covered the
440 Alps during the LGM, and thus created a current postglacial contact zone (*i.e.*, habitat
441 tracking hypothesis, *sensu* Brochmann *et al.*, 2003). The latter scenario is favoured by the
442 spatial genetic structure retrieved here (Figure 1a), which points out the existence of three
443 lineages (red, yellow and blue; Figure 1a) in this mountain range. The other three regions
444 that also harbour high levels of genetic diversity are the Sudetes, central Scandinavia and
445 northern Scotland. While the first two regions correspond to contact zones between
446 neighbouring genetic lineages (yellow and blue clusters for the Sudetes, green and yellow
447 clusters for central Scandinavia; Figure 1a), the third region represents a more unexpected
448 case of admixture, which could be due to long-distance dispersal events of individuals from
449 three different clusters that reached the north of the British Isles independently [*i.e.*, three
450 different lineages (red, yellow and blue; Figure 1a) are admixed in one single population in
451 the Grampians Mounts].

452

453 **General considerations on ENM and hindcasting for *T. europaeus***

454 Because (i) the current ENM-predicted suitable area for *T. europaeus* fits well the known
455 distribution of the species (Meusel *et al.*, 1965) (Figure 1d) and (ii) the hindcasted LGM
456 suitable area predicted (Figure 1c) is congruent with past distributions hindcasted for boreal
457 trees such as *Betula pubescens* or *Salix caprea* (Svenning *et al.*, 2008) that currently occupy
458 habitats similar to that of *T. europaeus* (European Commission DG Environment, 2007), it is
459 highly probable that the hindcasted model accurately reflects the past distribution of the
460 species. Once thresholds are applied, our results indicate that, as expected, the distribution
461 range of *T. europaeus* was more extended during the LGM than it is today and covered a
462 large part of the lands free of ice (Figure 1c). Several patches of the current distribution,
463 particularly in southern Europe, were thus probably interconnected during the LGM (see
464 below).

465 It is worth noting that the two threshold methods, “M” and “SS”, yielded very similar
466 results, the first being slightly more restrictive. With the latter, several hindcasted *suitability*
467 *hotspots* could have been interconnected, such as those corresponding to western Europe
468 (*iii*) and the Pyrenees (*i*) or to the Carpathians (*iv*) and the Balkans (*v*). Finally, one should
469 take into consideration that because during glacial cycles lowered sea levels rendered
470 possible several dry land connections, a continuous distribution might have occurred
471 between the Italian peninsula and both the Balkan (Mussi, 1990) and the West-Pyrenean
472 (Rigaud & Simek, 1990) *suitable hotspots*, which would also coincide with our molecular
473 results (Figure 1a).

474

475 **Relationships between genetic clusters and hindcasted *suitability hotspots***

476 Our novel approach combines inferences on the current intra-specific genetic structure
477 with information on hindcasted distributions and provides a new framework to understand the
478 dynamics of cold-adapted distributional contractions. Using *T. europaeus* as a case study,
479 we identified four genetic clusters (Figure 1a) and five *suitability hotspots* (Figure 1d), which
480 roughly match spatially.

481 The south-western cluster (red, Figure 1a) is spread in the Pyrenees as well as in
482 surrounding mountain ranges such as the Cantabrics and the French Massif Central, and
483 extends further East towards the western Alps and the Apennines, where it is admixed with
484 other neighbouring lineages. The eastward spread of the cluster into this latter massif could
485 attest of the existence of a corridor that, at this time, might have linked the Italian peninsula
486 with southern France (Rigaud & Simek, 1990). The dominance of this cluster in several
487 southern massifs, which can be considered as contemporaneous refugia for cold-adapted
488 species (Stewart *et al.*, 2010), shows that this lineage, despite being currently highly spatially
489 fragmented, is not yet subdivided into more restricted lineages. This result gives clues on the
490 timing of differentiation: the 18,000 years since the LGM were not sufficient to allow detecting
491 for genetic differentiation. Because the distribution of this cluster fits well with the position
492 and extent of the *suitability hotspot i* (Figure 1c), it is likely that the latter harboured the
493 ancestors of lineages currently found in the Pyrenees, Massif Central, Cantabria and to some
494 extent the Apennines.

495 The central/northwestern cluster (yellow, Figure 1a) covers the central Alpine region as
496 well as southern and central Scandinavia, central Europe and the British Isles. Some
497 admixture is observed on the edges of its distribution, represented mainly by contact zones
498 with other neighbouring genetic groups, as in the Sudetes mountain range, the Massif
499 Central and the Southern edges of the Alps. This cluster is the currently most widespread
500 across Europe and the most frequent in the Alpine arc. Explanations for this pattern could be
501 as follows: (i) this genotype was the most widely distributed across the European steppe
502 during the LGM; (ii) it was the most central and thus able to quickly colonize the newly
503 deglaciated lands (Scandinavia and the Alps) (Hewitt, 1996); (iii) a combination of (i) and (ii).
504 The distribution of this cluster fits well the *suitability hotspot iii* (Fig. 1c), located close to the
505 southern edge of the northern ice-sheet, which could represent the LGM origin of this
506 lineage. Its position and extent could largely explain the presence of the yellow cluster in the
507 British Isles, as well as in Southern Scandinavia. Accordingly, when the ice-sheets started to

508 melt down, invasion of new free lands in northern Europe would have happened more easily
509 from this hotspot than from any other.

510 The blue cluster (Figure 1a) is restricted to the Carpathians, the Sudetes, the Balkans and
511 to a lesser extent the south-eastern Alps and the Apennines. The presence of this cluster in
512 the latter massif (although admixed with the red cluster) is probably a consequence of land
513 exposure of part of the Adriatic sea due to lowered sea levels during the LGM, which allowed
514 the communication between the Italian peninsula and the Balkans (Mussi, 1990). This cluster
515 does not extend to the North further than the Sudetes, as shown in other Eurasian
516 organisms, in which the Carpathians harbour specific lineages, as in the cases of the plant
517 *Arabis alpina* (Ehrich *et al.*, 2007) or the butterfly *Erebia euryale* (Schmitt & Haubrich, 2008).
518 This pattern is expected since the northern Carpathians show a longitudinal extension along
519 a West/East axis, which is known to have provided shelter against the cold and dry winds
520 coming from the northern plains during glacial periods (Feurdean, 2004), but which also
521 rendered dispersal towards the North more difficult. Accordingly, current Poland and western
522 Ukraine were characterized by LGM climatic conditions too extreme to allow the survival of *T.*
523 *europaeus*, a species adapted to humid habitats. As indicated by the hindcasted ENM, the
524 Carpathian and Balkan regions appear to have provided suitable climatic conditions for *T.*
525 *europaeus* (Figure 1c), harbouring two *suitability hotspots* (*iv* and *v*) strongly isolated towards
526 the North. However, because only one genetic cluster was found in this region, we cannot
527 identify which of these two regions was the most probable source for the blue cluster. It is
528 also possible that the absence of a second genetic cluster in this area is a consequence of
529 the geographical proximity of these two hotspots, which might have experienced continuous
530 gene flow during glacial periods. Such a homogenising force would thus have limited *in situ*
531 diversification in each hotspot.

532 Finally, the location of the North-Scandinavian cluster (green, Figure 1a) suggests that
533 this lineage invaded the region from a zone outside the sampled range. Good candidates of
534 origin for this group are the Russian plains, an area known to have acted as a colonization
535 source of the whole Scandinavian peninsula in other organisms (e.g. Brochmann *et al.*, 2003;

536 Alsos *et al.*, 2009). Including plant samples from the Russian edge of the distribution range of
537 the plant would allow testing this hypothesis. The hypothesis of an eastern colonization of
538 northern Scandinavia is also compatible with results obtained by the hindcasting approach,
539 which identified the *suitability hotspot ii* in the south-eastern edge of the northern ice-sheet
540 (Figure 1d) that could have extended into the Russian plains. This hotspot area thus appears
541 as the most likely origin of the green cluster, which would have invaded northern Scandinavia
542 once the northern glacial sheets retracted.

543

544 **Preservation or loss of genetic diversity?**

545 In contrast to temperate species, cold-adapted species show refugial contracted
546 distributions during interglacial periods (Schmitt, 2009). In Europe, long-term and cryptic
547 refugia have been identified for cold-adapted arctic-alpine organisms, for instance in
548 Scandinavia and in the southern mountain ranges, respectively (Stewart *et al.*, 2010). The
549 distribution of *T. europaeus* currently covers most areas harbouring one or the other type of
550 these refugia.

551 Based on the combination of the two independent analytical approaches presented here,
552 we can propose a post-glacial contraction scenario for *T. europaeus* (Figure 3), in which four
553 contraction/colonization events, mostly in a centrifugal manner (*i.e.*, from central lowlands to
554 surrounding highlands), explain the current spatial genetic structure found in the species.

555 As shown in Figures 1 and 3, the number of genetic clusters is lower than that of *suitability*
556 *hotspots* during the LGM. Based on the parsimony criterion it seems plausible that all but one
557 *suitability hotspots* participated to the current genetic diversity since in the area comprising
558 the Balkans (*suitability hotspot v*) and the Carpathians (*suitability hotspot iv*) only one genetic
559 cluster was retrieved (see grey crossed zone in Figure 3). Past genotypes associated with
560 one of these two hindcasted hotspots could have been lost during post-glacial contraction or
561 are currently restricted to highly localized areas, not covered by the present study (*e.g.*,
562 current Montenegro and Bosnia-Herzegovina). It is also likely that the distance between the
563 Carpathian and Balkan *suitable hotspots* was small enough to allow gene flow and that these

564 two regions finally shared the same lineage. As a consequence, the loss of diversity in *T.*
565 *europaeus* during the interglacial contraction period seems to be reasonably low, especially
566 when compared to results obtained for organisms such as the arctic fox, which lost a
567 substantial proportion of its Pleistocene genetic diversity after the end of the LGM (i.e., all
568 European and some Scandinavian lineages, Dalén *et al.*, 2007).

569 In the present work, we demonstrate that the inference of postglacial dynamics of cold-
570 adapted species, for which no ancient DNA is available, is possible by the simultaneous use
571 of hindcasting and current spatial genetic structure analyses. In the future, and in order to
572 test more accurately for the post-LGM contraction addressed in this study, additional
573 coalescent-based approaches should be performed and the likelihood of different scenarios
574 should be tested. This, however, would require an accurate evaluation of parameters related
575 to population demography, a challenge that has not been overcome yet when using AFLP
576 data (e.g., Beerli & Felsenstein, 1999; Excoffier *et al.*, 2005).

577

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585

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741

742 **BIOSKETCH**

743 Anahí Espíndola is a PhD student at the University of Neuchâtel (Switzerland). She is
744 interested on the evolutionary history of interspecies interactions. Her research integrates
745 spatial, historical and ecological approaches to answer general phylogeographic and
746 biological questions. Loïc Pellissier is a PhD student at the University of Lausanne
747 (Switzerland) and currently works on ecological niche modelling approaches applied on
748 conservation of Alpine butterfly communities. Nadir Alvarez is a research associate at the
749 University of Lausanne, interested in the ecology and evolution of biological interactions.

750

751 Table 1 - Area under the curve (AUC) values for the different BIOMOD modeling techniques.

752

| Modeling technique | AUC |
|---|------------|
| Artificial Neural Networks (ANN) | 0.797 |
| Classification Tree Analysis (CTA) | 0.815 |
| Generalized Additive Model (GAM) | 0.829 |
| Generalized Boosting Model (GBM) | 0.841 |
| Generalized Linear Model (GLM) | 0.826 |
| Multivariate Adaptive Regression Splines (MARS) | 0.826 |
| Mixture Discriminant Analysis (MDA) | 0.772 |
| Random Forest (RF) | 0.847 |

753

754

755 **Figure captions**

756

757 Figure 1 – Genetic structure (a) and estimated Nei genetic diversities (b) of *Trollius*
758 *europaeus* in Europe. Projected climatic probabilities for past (c) and current (d) conditions,
759 as inferred from the Ecological Niche Model (ENM) and after the application of the two
760 thresholds (“M” and “SS”; see text). a – proportions of the pies indicate the probability to
761 belong to one of four genetic clusters (colours), as estimated by STRUCTURE 2.2 (Pritchard
762 *et al.*, 2000). b –dot sizes are proportional to the Nei genetic diversities extrapolated using a
763 moving-window approach; scales are referred in the figure legend. c – d – probability values
764 are indicated on each figure. White: values below both thresholds; grey: values below “M”
765 threshold; black: values above both thresholds; in c, dotted areas correspond to glaciated
766 regions; dashed lines define LGM coasts, as delimited by Ehlers and Gibbard (2007); the
767 five defined *suitable hotspots* are illustrated by roman numbers.

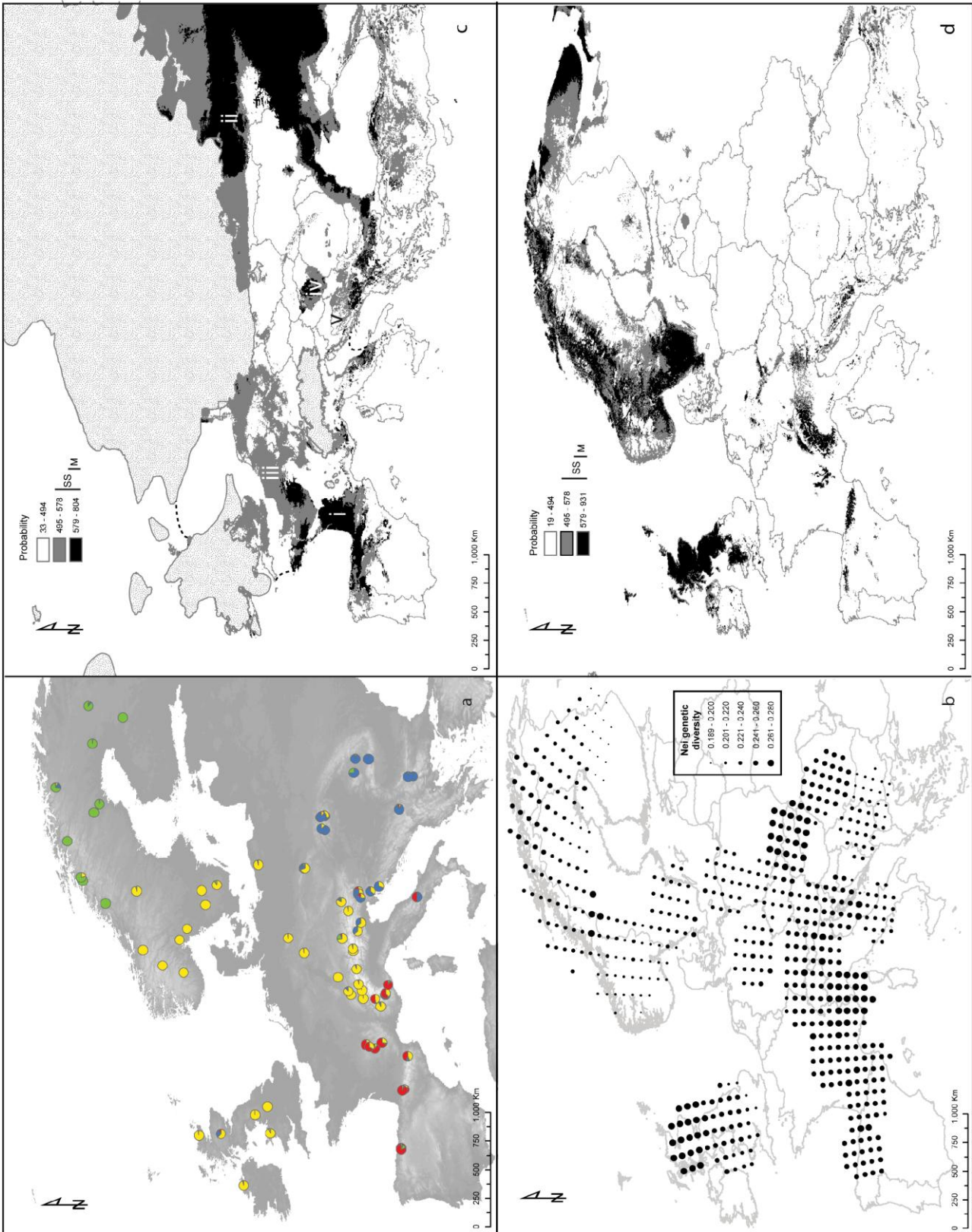
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769 Figure 2 – Midpoint-rooting tree reconstructed with AFLP-based Nei-Li distances and
770 Neighbour-Joining reconstruction for *Trollius europaeus*. Colours indicate regional clusters
771 as retrieved in Figure 1a.

772

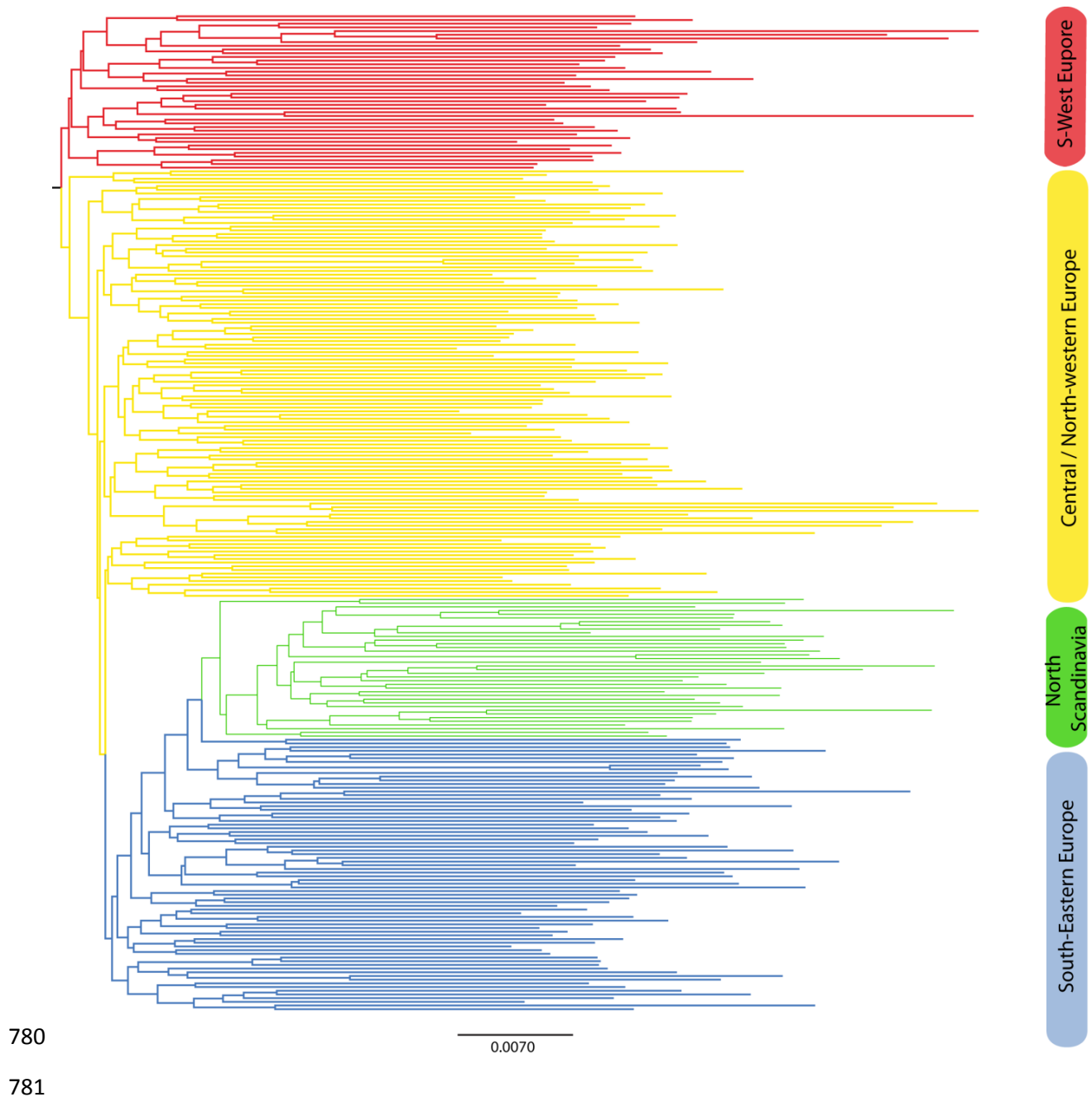
773 Figure 3 – Post-glacial contraction route scenarios for *Trollius europaeus*. Colours
774 correspond to different genetic groups as in Figures 1 and 2; arrows indicate the direction of
775 movement; thick dashed lines show main contact zones. The grey zone delimited by thin
776 dashed lines indicates the putative region for which no genetic group was identified.

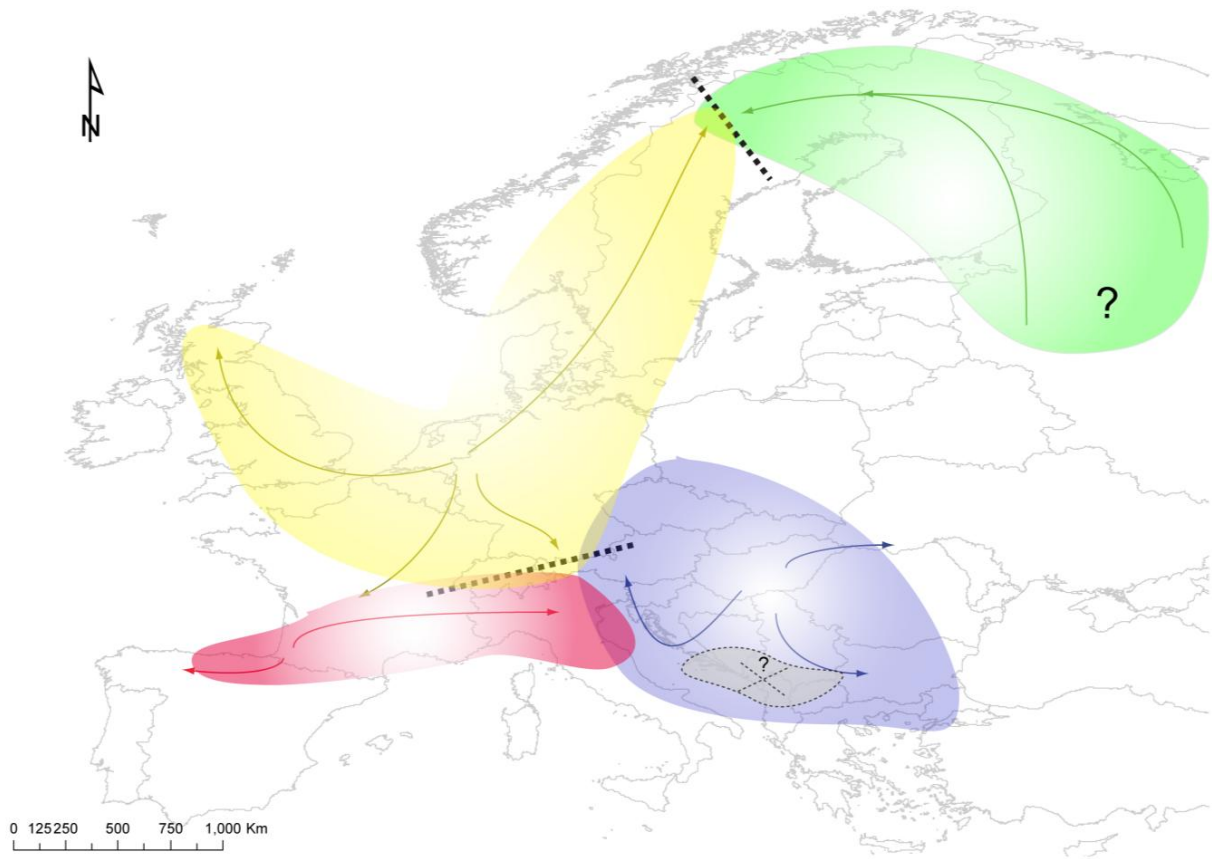
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784 Appendix S1 – Locations of sampled sites

| Location | Country | Longitude | Latitude (m) | Altitude |
|------------------------|-------------|-----------|--------------|----------|
| Ambri | Switzerland | 8.70292 | 46.50680 | 1000 |
| Amot | Norway | 8.42346 | 59.62199 | 481 |
| Badgastein | Austria | 13.09639 | 47.12056 | 1963 |
| Bayasse | France | 6.74067 | 44.30814 | N/A |
| Beistohlen | Norway | 8.95473 | 61.20761 | 729 |
| Bidjovagge | Norway | 22.47808 | 69.29778 | 613 |
| Bobolice | Poland | 16.59895 | 53.94660 | 110 |
| Braas | Sweden | 15.06817 | 57.09309 | 209 |
| Chasseral | Switzerland | 7.02130 | 47.12569 | N/A |
| Chemin | Switzerland | 7.08978 | 46.08993 | N/A |
| Col de Bonnecombe | France | 3.11410 | 44.57557 | 1335 |
| Col de la Colombiere | France | 6.46972 | 45.98722 | 1600 |
| Col de Roanza | Italy | 12.20989 | 46.17638 | 816 |
| Col du Festre | France | 5.86278 | 44.66639 | 1430 |
| Col du Galibier | France | 6.43861 | 45.08528 | 2000 |
| Col long de Magnabaigt | France | -0.43646 | 42.87060 | 1615 |
| Colt Park | England | -2.35247 | 54.19365 | 380 |
| Crans-Montana | Switzerland | 7.53890 | 46.34650 | N/A |
| Cressbrook Dale | England | -1.74041 | 53.26724 | 246 |
| Creux du Van | Switzerland | 6.74119 | 46.93526 | N/A |
| Donovaly | Slovakia | 19.23068 | 48.88922 | 1345 |
| Eidda Pastures | Wales | -3.74190 | 53.03720 | 234 |
| Ellingsrudelva | Norway | 10.91844 | 59.91771 | 161 |
| Esposouille | France | 2.09450 | 42.62341 | 1521 |
| Fanske | Norway | 15.40131 | 67.26452 | 46 |
| Feleacu | Romania | 23.59028 | 46.70118 | 670 |
| Froson | Sweden | 14.60268 | 63.18205 | 424 |
| Glen Fender | Scotland | -3.79485 | 56.78138 | 345 |
| Haiduta Mt | Bulgaria | 23.32314 | 42.19968 | 2430 |
| Haute Tinee 1 | France | 6.81871 | 44.29617 | 1770 |
| Haute Tinee 2 | France | 6.85581 | 44.28426 | 1770 |
| Hlupy | Slovakia | 20.21572 | 49.24040 | 1775 |
| Kajaani | Finland | 27.77275 | 64.22848 | 141 |
| Karersee | Italy | 11.57341 | 46.40885 | N/A |
| Karersee | Italy | 11.57341 | 46.40885 | N/A |
| Karlow | Poland | 16.34070 | 50.47810 | 759 |
| Krasno Polje | Croatia | 14.97021 | 44.80746 | 1444 |
| Lac Balea | Romania | 24.62192 | 45.60239 | 2224 |
| Laktatjakka | Sweden | 18.40674 | 68.42931 | 453 |
| Logarska Dolina | Slovenia | 14.58630 | 46.36956 | 1401 |
| Loser | Austria | 13.78485 | 47.66052 | 1598 |

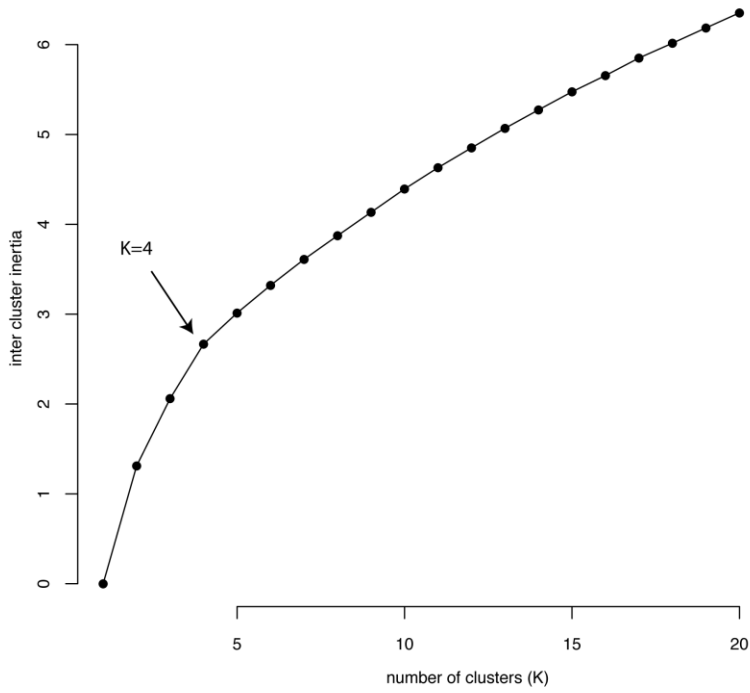
| Location | Country | Longitude | Latitude (m) | Altitude |
|------------------------|---------------|-----------|--------------|----------|
| Lough Fern | North Ireland | -7.71130 | 55.06569 | 26 |
| Moerlimatt | Germany | 8.07760 | 47.90597 | 939 |
| Monte Pizi | Italy | 14.16714 | 41.91524 | 1546 |
| Muntii Fagarasului | Romania | 24.63611 | 46.59972 | 2205 |
| Naverdal | Norway | 10.13002 | 62.70417 | 480 |
| O. Olteid | Norway | 11.74560 | 59.34895 | 104 |
| Pajino Preslo | Serbia | 20.81970 | 43.27799 | 1802 |
| petit papa noel | Finland | 25.79386 | 66.51647 | 72 |
| Pila | Slovakia | 20.29449 | 48.90017 | 972 |
| Podlesok | Slovakia | 20.35190 | 48.94962 | 579 |
| Puerto de Panderruedas | Spain | -4.97223 | 43.12743 | 1227 |
| Puy de Dome | France | 2.96333 | 45.77222 | 1460 |
| Puy de Sancy | France | 2.80972 | 45.53500 | 1520 |
| Puy Mary | France | 2.68083 | 45.11139 | 1550 |
| Radkow | Poland | 16.35321 | 50.46866 | 712 |
| Risnjak - Snjeznik | Croatia | 14.58494 | 45.43871 | 1466 |
| Salla | Finland | 28.65427 | 66.83020 | 194 |
| Sede de Pan | France | -0.48651 | 43.03949 | 1556 |
| Seterasen | Norway | 13.67744 | 65.53432 | 285 |
| Sjostorp | Sweden | 14.65743 | 58.24696 | 100 |
| Solberga | Norway | 13.56116 | 57.95194 | 239 |
| Sommerberg | Germany | 9.92068 | 50.47913 | 770 |
| Sta Marina de Valdeon | Spain | -4.87745 | 43.12790 | 1294 |
| Steingaden | Germany | 11.01296 | 47.59529 | 1158 |
| Storzic | Slovenia | 14.39488 | 46.34390 | N/A |
| Strath Hallerdale | Scotland | -3.89403 | 58.45140 | 36 |
| Straumen | Norway | 15.64921 | 67.38440 | 75 |
| Susch | Switzerland | 10.07473 | 46.74728 | N/A |
| Svartla | Sweden | 21.22062 | 65.99583 | 36 |
| Tarasp | Switzerland | 10.25056 | 46.77730 | N/A |
| Tatranska Lomnica | Slovakia | 20.27944 | 49.16582 | 839 |
| Tende | France | 7.49970 | 44.10770 | 1622 |
| Trollblumenwiese | Germany | 11.04118 | 51.68314 | 488 |
| Val de Choc | Slovakia | 19.34333 | 49.15083 | 1550 |
| Vitoshka | Bulgaria | 23.29342 | 42.59032 | 1779 |
| Vuollerim | Sweden | 20.57896 | 66.42037 | 166 |
| Zali Log | Slovenia | 14.11080 | 46.20342 | 533 |
| Zielowe Ludowe | Serbia | 16.34920 | 50.40210 | 695 |

785

786

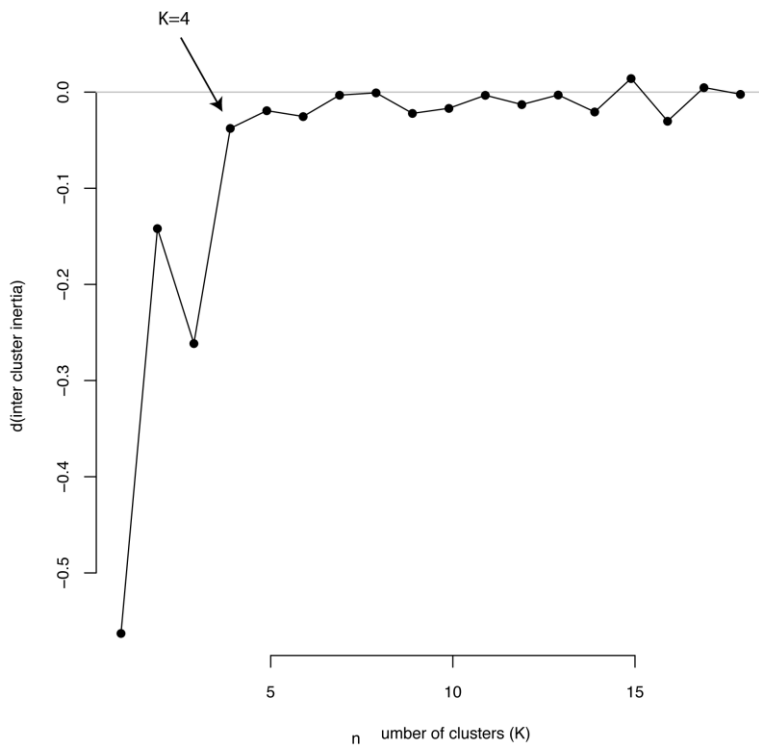
787 Appendix S2 – First (a) and second (b) derivatives for inertias, for all values of K calculated
 788 using the K-means approach.

789 a)



790

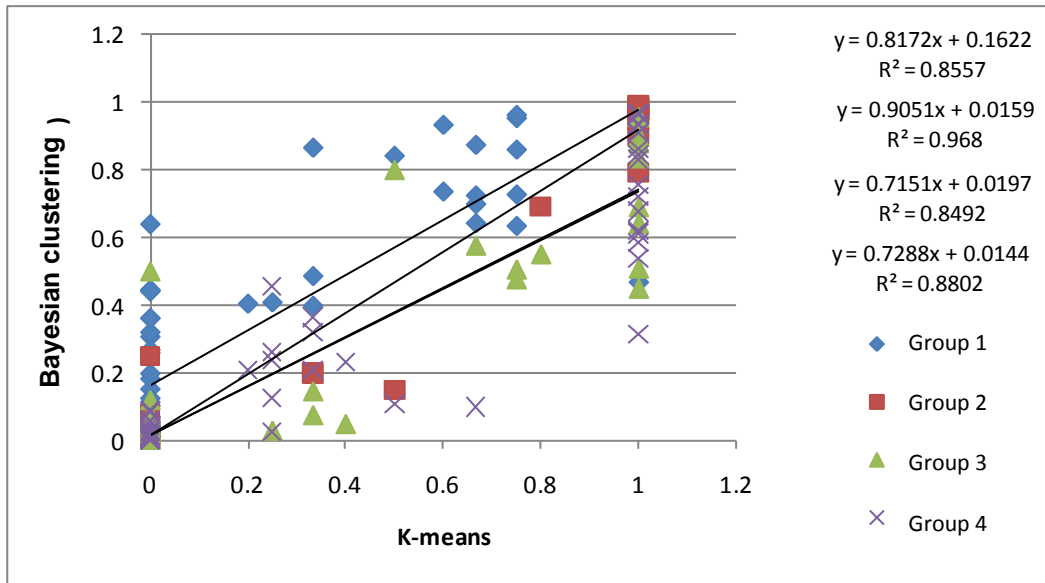
791 b)



792

793 Appendix S3 –Correlation between sample assignments considering the two genetic
 794 clustering methods (Bayesian and K-means) used in this study.

795



796

797

Chapter eight

Phylogeography of mutualists and the effect of the host on the genetic structure of its partners

Anahí Espíndola, Bryan C. Carstens and Nadir Alvarez

1 **Phylogeography of mutualists and the effect of the host on the**
2 **genetic structure of its partners**

3 Anahí Espíndola¹, Bryan C. Carstens² and Nadir Alvarez³

4 ¹Laboratory of Evolutionary Entomology, Institute of Biology, University of Neuchâtel.
5 Emile-Argand 11, 2000 Neuchâtel, Switzerland.

6 ²Life Sciences Building, Louisiana State University, Baton Rouge, LA 70803 USA.

7 ³Department of Ecology and Evolution, University of Lausanne, Biophore Building, 1015
8 Lausanne, Switzerland

9

10 **Abstract**

11 The field of phylogeography has seen an important development during the last decades,
12 especially in Europe and North America. In spite of this, little is known about the effect of
13 ecological interactions on the phylogeographic patterns of interacting organisms. However,
14 the coevolutionary theory predicts that interacting species should present similar histories,
15 especially if interactions are specific and obligate.

16 To investigate this idea, we use here the mutualistic nursery pollination system formed by
17 the Eurasian arctic-alpine species *Trollius europaeus* L. and its specific pollinators of the
18 genus *Chiastocheta* Pokorny as a case study. In this system, the plant is visited and
19 pollinated exclusively by the flies, whose larvae feed specifically on the plant seeds.
20 Because the system concerns a low number of interacting species, it is highly appropriate to
21 test the effect of interactions in a comparative framework. Based on a large-scale sampling
22 of pollinating flies, we first infer the phylogeographic histories of each fly species from
23 mitochondrial data. Because the large-scale genetic structure of the plant has been recently
24 identified, we exploit this information and ask whether or not the phylogeographic patterns of
25 the flies are significantly different of what would be expected under a scenario of plant-insect
26 congruence.

27 Our results demonstrate that while all fly species occupy an extremely close ecological
28 niche, their phylogeographic histories highly differ. While *C. rotundiventris* presents no
29 geographic structure, *C. lophota* has a genetic structure largely explained by geography,
30 while the phylogeographic pattern of *C. dentifera* does not statistically differ of what would be
31 expected if it had evolved under a scenario guided by the plant. These results indicate that
32 forces shaping the genetic structure of the three species are highly different. Differential
33 dispersion capabilities could explain these patterns.

34

35 Introduction

36 The high number of species and enhanced rate of diversification in insects and
37 angiosperms is often explained by reciprocal adaptive radiation of these two groups in a
38 coevolutionary context (Simpson 1953; Schluter 2000; Lunau 2004; Futuyma and Agrawal
39 2009). In the coevolutionary model of Ehrlich & Raven (1964), the “escape and radiate”
40 process enhances codiversification of plants and insects ecologically related by herbivory.
41 Adjustments against exploitation from the plant side can arise in two opposite directions:
42 either by defending chemically or physically against insects, or by evolving towards a
43 cooperative interaction in which the cost of insect exploitation is balanced by an ecological
44 service imposed by the plant (Dufaÿ and Anstett 2003). The latter case is notably
45 encountered in mutualistic pollination systems, in which costs and benefits of plants and
46 insects tend to be equilibrated. Ecological interactions thus include a strong coevolutionary
47 component (Thompson 2009) that in cases of strict coevolution, leads to an increased
48 specificity of the plant-insect relationship, or in cases of diffuse coevolution, maintains a
49 generalist relationship (Lunau 2004).

50 Specific and obligate interactions, though much rarer than generalist relationships
51 (Ollerton, Killick et al. 2007), represent simple cases for hypothesis testing and have
52 therefore been frequently used as models by evolutionary biologists. Investigating biological
53 systems with a reduced number of specifically interacting organisms allows understanding
54 processes that would be much more difficult to disentangle when working with more relaxed
55 interactions, as those happening in generalist relationships. Most studies emphasizing the
56 fate of organisms in a context of strict coevolution and interested in unraveling evolutionary
57 histories in pollination mutualisms have focused on large spatio-temporal scales, showing
58 long-term processes putatively leading to cocladogenesis and codiversification (Agosta
59 2006), as for instance in the case of the fig and fig-wasps (Jousselin, van Noort et al. 2008).
60 However, investigation at smaller evolutionary scales holds the potential to inform us on the
61 origin and maintenance of specific mutualistic pollination. A few biological systems have

62 served for this purpose, as for instance, the interaction between *Trollius europaeus* L.
63 (Ranunculaceae) and the pollinating flies within the genus *Chiastocheta* Pokorny (Diptera:
64 Anthomyiidae), which represents a unique widespread European example of specific
65 mutualism between plants and insects (Pellmyr 1989).

66 *Trollius europaeus*, the European globeflower, is a geophyte inhabiting cold meadows in
67 the west-Palearctic, occupying southern European mountainous environments and being
68 largely present in northern arctic-alpine areas of the continent (Pellmyr 1992). This species
69 is characterized by a special flower morphology, adapted to the nursery pollination
70 interaction it maintains with the *Chiastocheta* species complex (Pellmyr 1992). In this
71 system, the plant is visited and specifically pollinated by the small Anthomyiids, which visit
72 the flowers to lay eggs on carpels, since larvae feed specifically on *T. europaeus* seeds.
73 There is strong evidence to consider that the mutualistic behaviour of *Chiastocheta* flies
74 derives from phytophagy on different plant organs, since closely-related fly genera such as
75 *Delia* Robineau-Desvoidy are generalist herbivores (Soós and Papp 1986). Considering this
76 biological system in a coevolutionary framework, it is likely that herbivore pressure was
77 released by evolving towards a specific pollination mechanism. While eight *Chiastocheta*
78 species have been described based on taxonomic grounds (Michelsen 1985; Pellmyr 1992;
79 Jaeger and Després 1998), a recent phylogenetic study by Espíndola et al. (unpublished
80 data) has shown that the genetic identity of these species is far from being well
81 circumscribed, with hybridization happening among several taxa. In fact, only three species
82 are consistent from both genetic and morphological points of view: *C. rotundiventris*, *C.*
83 *lophota* and *C. dentifera*; the remaining taxa appear either impossible to be identified by
84 means other than morphology (*C. abruptiventris*), or extremely admixed with other
85 morphotypes (*C. macropyga*, *C. trollii*, *C. inermella*, *C. setifera*). In that study, authors have
86 also shown that diversification within the genus *Chiastocheta* arose during the Quaternary
87 (up to 2.6 Mya). This specific interaction is thus much more recent than other systems such
88 as those involving figs and fig wasps, which originated around 75 million years ago (Cruaud,
89 A., University of Montpellier II, unpublished data).

90 Here, we exploit the unique attributes of this biological system to investigate the fate of
91 specific mutualists at short-time evolutionary scales. We study the synchrony of
92 diversification and more generally the congruence in phylogeographic histories of interacting
93 species. This is done in order to address whether or not patterns similar to those found in
94 more ancient specific mutualisms such as the case of figs and fig wasps (i.e., high level of
95 congruence and large-scale co-cladogenesis) can be retrieved at smaller temporal scales.
96 Because mutualistic species are strongly dependent on each other, we predict that *T.*
97 *europaeus* and *Chiastocheta* flies should experience similar environmental and geographical
98 effects on their dispersal and demographic components, and that their phylogeographic
99 histories should thus be highly congruent. The alternative hypothesis is that the genetic
100 structures of mutualistic species are different, what would indicate no effect of the interaction
101 on the observed phylogeographic patterns.

102 The pollination mutualism involving *T. europaeus* and its pollinating flies is well-suited to
103 test such a hypothesis. First, the phylogeography of European organisms is well understood.
104 Two decades of phylogeographic studies in this continent have allowed unraveling general
105 phylogeographic patterns and identifying spatial (Avice 2009) and ecological (Alvarez, Thiel-
106 Egenter et al. 2009) factors responsible for the current spatial genetic structure of species.
107 For instance, it is currently largely accepted that during glacial periods temperate species
108 found refugia in southern peninsulas (Taberlet, Fumagalli et al. 1998; Hewitt 1999), and
109 sometimes in northern refugia (Stewart and Lister 2001; Bhagwat and Willis 2008; Provan
110 and Bennett 2008). Recently, Stewart, Lister et al. (2010) have summarized the effect of
111 glacial and interglacial periods for cold-related species, which are found today under refugial
112 situations occupying the southern European mountain ranges as well as northern areas.
113 Second, the interaction between the globeflower and its pollinators has been widely studied
114 at the European scale, especially in the Alps (Jaeger and Després 1998) and southern
115 Scandinavia (Pellmyr 1989; Johannesen and Loeschcke 1996). In these studies, authors
116 have identified a sound ecological background relative to the strength and characteristics of
117 the interaction maintained by the different *Chiastocheta* species and to the dynamics of the

118 system. Third, new analytical approaches have been recently proposed in the field of
119 phylogeography, to go further than simply qualitatively describing the distribution of lineages
120 in space (Hickerson, Carstens et al. 2010). Probably the most ground-breaking of these
121 recent approaches is the application of statistical phylogeography, which is derived from the
122 theory of allele coalescence (Kingman 1982) and which proposes defining and statistically
123 testing different phylogeographic scenarios (Knowles and Maddison 2002). This technique
124 considers that the variation of coalescent times of trees contained in species (or population)
125 trees can be used to create an expected distribution of a given statistic under a given
126 scenario, which allows testing the probability of the empirical data to have been produced
127 under that given scenario. The technique has been used for a little less than ten years, and it
128 is currently acquiring importance, especially after the demonstration of its versatility when
129 used in combination with ecological niche models and hindcasted spatial distributions
130 (Carstens and Richards 2007), or with other geographical external data as glaciation and
131 deglaciation times (Knowles 2001).

132 While one other study has focused on similar questions at small evolutionary scales using
133 coalescent approaches (DeChaine and Martin 2006), our study is the first using a pollination
134 mutualism to test for congruence in recent evolutionary histories and directly exploiting the
135 known genetic structure of one of the organisms as a model into which the phylogeographic
136 history of its partner should fit. Because the plant genetic structure has been recently shown
137 to have been influenced by the Last Glacial Maximum (LGM, 21-18 thousand years ago -
138 Kya; Espíndola, Pellissier et al. in preparation), we propose here to define and test
139 hypotheses asking whether or not the phylogeographic patterns of the insects are driven by
140 that of the plant. Based on three sequenced regions from the mitochondrial genome, we first
141 infer the phylogeographies of each well-defined insect species (*i.e.*, *C. rotundiventris*, *C.*
142 *lophota* and *C. dentifera*) and further test if the phylogeographic patterns of the insects are
143 more similar to a model fitting the plant's phylogeographic pattern than what would be
144 expected by chance.

145

146 **Material and Methods**147 ***Sampling and DNA amplification***

148 Insect samples covering the whole European distribution range of the plant were
149 collected during springs 2006-2008 and preserved in 70% ethanol. Samples were identified
150 at the species level (Sup. Mat. 1) following Hennig (1976) and further confirmed by the
151 European specialist of Anthomyiids, V. Michelsen (Natural History Museum of Denmark). In
152 several cases, only expert identification allowed identifying morphotypes that were not
153 described by Hennig (1976) (e.g., *C. lophota*). Only the three taxa presenting a consistent
154 phylogenetic identity (*i.e.*, *C. rotundiventris*, *C. dentifera*, and *C. lophota*; Espíndola, Buerki
155 et al. in preparation) were further selected to infer phylogeographic patterns and
156 demographic parameters.

157 DNA of 87 *C. rotundiventris*, 38 *C. dentifera* and 47 *C. lophota* samples was extracted
158 using the QIAGEN DNeasy Animal Tissue extraction kit (QIAGEN, Hombrechtikon,
159 Switzerland), following the manufacturer's indications. Three mitochondrial regions (*COI*,
160 *COII* and *D-loop*) were amplified using primers shown in Table 1. PCRs were done in a 20µl
161 mix consisting of 0.5X buffer, between 1 and 2.5 mM MgCl₂, 10mM dNTPs, 1 unit of GoTaq
162 DNA polymerase (Promega, Dübendorf, Switzerland), 0.5 µM primers and 3 µl DNA and run
163 in a TGradient thermocycler (Biometra, Goettingen, Germany). Two types of thermocycler
164 programs were used, depending on the region amplified: for *COI* and *COII*, we used 1:30
165 min at 95°C, followed by 40 cycles of 35 secs at 95°C, 1 min at 52°C, 45 secs at 72°C, and a
166 final elongation of 8 min at 72°C; for the *D-loop*, which is a highly A-T rich region, we used 5
167 min at 95°C, followed by 35 cycles of 1 min at 95°C, 1 min at 55°C, 2 min at 60°C and a final
168 elongation of 5 min at 60°C. Amplified fragments were purified and sequenced at MacroGen
169 Inc. (South Korea) and FASTERIS S.A. (Switzerland). Chromatograms were corrected on
170 ChromasPro 1.41 (Technelysium Pty. Ltd.), aligned either using a Clustal-Wallis algorithm

171 for *COI* and *COII* (Thompson, Higgins et al. 1994) as implemented in BioEdit 7.0.4.1 (Hall
172 1999) or applying the moderately accurate option proposed by the MAFFT v6 online
173 alignment service for the *D-loop* (Kato and Toh 2008). All alignments were further visually
174 checked and corrected if necessary. Gaps were coded applying the simple gap-coding
175 method of Simmons and Ochoterena (2000), as implemented in FastGap1.2 (Borchsenius
176 2009). Total, variable, constant and parsimony informative sites were calculated per species
177 and mtDNA region using MEGA 4.0 (Tamura, Dudley et al. 2007).

178 ***Phylogenetic inferences and haplotype networks***

179 MrBayes 3.1.2 (Ronquist and Huelsenbeck 2003) was used to infer Bayesian inference
180 phylogenies for *C. dentifera* and *C. lophota* (no convergence was reached for *C.*
181 *rotundiventris*, see Results). For each single species alignment, we parameterized two
182 Markov-chain Monte Carlo (MCMC) runs with two chains, consisting of 50'000'000
183 generations, a temperature of 0.5 and one sampled tree every 1'000 generations. Data was
184 partitioned for each mtDNA region using specific models of evolution following results
185 obtained with MrAIC (Nylander 2004). A restriction model of evolution was assigned to the
186 partition corresponding to gaps (coded as binary data). Convergence was assumed when
187 standard variation between chains fell below 0.01, when the Potential Scale Reduction
188 Factor index (Gelman and Rubin 1992) reached at most 1.002, when a unimodal distribution
189 of sampled parameters was retrieved and when sampling for all parameters presented
190 values of ESS (Effective Sampling Size) higher than 200 (checked on Tracer v1.4, Rambaut
191 and Drummond 2004). Half-compatible consensus trees were calculated applying burnins of
192 10'000'000 (*C. dentifera*) and 20'000'000 (*C. lophota*) generations.

193 Maximum Likelihood (ML) searches were done using RaxML 7.2.6 (Stamatakis 2006)
194 with a 10'000 rapid bootstrap analyses followed by the search of the best-scoring ML tree in
195 one single run. Here, we considered the three mtDNA regions as one single partition and did
196 not account for gap information. A majority-rule consensus tree was further inferred from the

197 node information. This analysis was done using the facilities offered by the CIPRES portal
198 (San Diego, CA, USA).

199 Maximum Parsimony (MP) searches were performed using parsimony ratchet (Nixon
200 1999) as implemented in PAUPrat (Sikes and Lewis 2001). Based on recommendations by
201 Nixon (1999), ten independent searches were performed with 200 iterations and 15% of the
202 parsimony informative characters perturbed. The shortest equally most parsimonious trees
203 were combined to produce a majority rule consensus tree. To assess the support at each
204 node, Bremer supports (Bremer 1988) were calculated using TreeRot 3.0 (Sorenson and
205 Franzosa 2007).

206 All topologies were rooted based on the general relationships previously inferred and
207 proposed by Després, Pettex et al. (2002) and Espindola et al. (unpublished data), which
208 considered the genus *Delia* Robineau-Desvoidy as the closest outgroup.

209 Finally, haplotype-networks were constructed using TCS 1.21 (Clement, Posada et al.
210 2000), applying a connectivity threshold of 95% (*i.e.*, networks were unlinked if they differed
211 by more than 95% of their sequences). While gaps were considered as missing data for *C.*
212 *lophota* and *C. rotundiventris*, they were considered as a fifth state in *C. dentifera*, because
213 of the lower level of polymorphism in the latter species.

214 In order to identify the spatial genetic structure of each of the three species, supported
215 clusters were plotted on maps using ArcMap (ESRI, Redlands CA, USA).

216 ***Phylogeographic hypothesis testing***

217 To test whether the spatial genetic structures of the insects are congruent with that of the
218 plant, we applied a statistical phylogeography approach, as proposed by Knowles and
219 Maddison (2002). This approach consists in defining different alternative phylogeographic
220 scenarios –represented by coalescent population trees– on which trees and data are
221 simulated. Based on these simulations, a null distribution of a chosen statistic is produced
222 and is used to test a given phylogeographic scenario based on empirical data. This is done

223 by evaluating empirical values in a percentage range (generally 2.5%-97.5%) of the null
224 model distribution (Knowles and Maddison 2002). Because demographic parameters can
225 have a non negligible influence on the time of coalescence (Maddison 1997), one of the key
226 points when applying these methods is the use of values and parameters that specifically
227 represent the empirical data. In order to apply these methods to our study we first defined
228 different phylogeographic hypotheses. For this, we considered the spatial genetic structure
229 unraveled in a previous study on the plant (Espíndola et al, submitted), which allowed us to
230 rely on an external and independent source of clustering, a principle similar to the one
231 Carstens and Richards (2007) applied when exploiting information from ecological-niche
232 models. Specifically, population trees were defined by assigning insect samples based on
233 the genetic cluster to which the plant populations had previously been assigned (see Figure
234 1). This assignment was straightforward since samples from insects and plants had been
235 collected simultaneously in all localities visited. Because actors of this mutualism are cold-
236 adapted species (*sensu* Stewart, Lister et al. 2010) and since the current distribution of the
237 plant has been driven by the last range contraction following the LGM (in our case related to
238 the termination of the last glacial period, Raymo 1997), we used splitting times of lineages as
239 follows (Figure 1): in the working hypothesis H_1 , the genetic clustering of the insect was
240 hypothesized to have followed contraction of plant lineages after complete deglaciation of
241 the main European lowlands (-13Kya, Raymo 1997), while in the alternative hypothesis H_2 ,
242 the genetic clustering was hypothesized to have followed contraction of plant lineages
243 immediately after the previous Glacial Maximum (-23 Kya, Boulton, Dongelmans et al. 2004).
244 The null hypotheses for each of these scenarios were that the splitting of insects' genetic
245 clusters was not defined by the distribution of plant lineages (*i.e.*, phylogeographies of the
246 insects and the plant were not related).

247 The demographic parameter Θ takes into account the effective population size (N_e) and
248 the mutation rates (μ) ($\Theta=N_e*\mu$, in mitochondrial maternally inherited data; Beerli 2008) and
249 was calculated for the two species in which a spatial genetic structure was retrieved (*i.e.*, *C.*
250 *lophota* and *C. dentifera*, see Results) using the MCMC approach applied in a Maximum-

251 Likelihood (ML) framework as implemented in Migrate-n (Beerli and Felsenstein 1999; Beerli
252 and Felsenstein 2001) and run on the HPC-LSU cluster (Baton Rouge, LA, USA). Two runs
253 of ten short and three long chains were used for 50,000,000 generations, sampling one
254 value every 1,000 generations and applying a burnin period of 10'000'000 generations.
255 Convergence between runs was verified using the Gelman criterion (Gelman and Rubin
256 1992) (option *gelman-convergence=YES:PAIRS*). Once this parameter was inferred, we
257 applied it in the software ms (Hudson 2002) to simulate 1,000 tree topologies fitting the
258 coalescent models represented by our hypotheses. Based on these topologies, sequences
259 carrying the same characteristics as the empirical data (*i.e.*, sequence length, model of
260 evolution, number of variable sites) were simulated for each simulated tree using seq-gen
261 v1.3.2 (Rambaut and Grassly 1997). Following the approach proposed by Carstens et al.
262 (2005), we considered two rates of mutation (μ): either 1×10^{-7} or 1×10^{-8} . Using these
263 sequences, parsimony heuristic searches were performed with PAUP*4b10 (Swofford 2003),
264 with TBR branch swapping, maxtrees = 100 and 10 random-addition sequence replications.
265 A strict consensus topology of the most parsimonious trees was computed for each dataset
266 and the resulting trees (1'000) were used to create a null distribution of the test statistic.

267 To estimate the fit or discordance between gene (*i.e.*, simulated and empirical data) and
268 species (*i.e.*, phylogeographic scenarios) trees (Knowles and Maddison 2002), we used the
269 statistic S of Slatkin & Maddison (1989), which indicates the minimum number of migration
270 events (parsimony steps) required to fit the phylogeny into the population tree. This statistic
271 is optimal in our case, because we aim at testing the congruence between the plant and the
272 insects' spatial genetic structures. Because our hypotheses consider that dispersal events
273 should have happened in parallel among the different actors involved, using a measure of
274 migration appears appropriate. Calculations of S for simulated trees and empirical data (*i.e.*,
275 the Bayesian phylogenetic tree inferred from real data) were performed using MESQUITE
276 2.72 (Maddison and Maddison 2009). We considered rejection of hypotheses H_1 or H_2 if the
277 empirical value fell outside the 2.5%-97.5% range of the simulated data (*i.e.*, $\alpha = 5\%$).

278

279 **Results**

280 The geographic distribution of the sampled species covered their entire known European
281 distribution ranges (Figures 2, 3 and 4) (Pellmyr 1992). Total length, number of constant,
282 variable, parsimony informative sites and number of gaps per species are shown for each
283 mtDNA region in Table 2. The *D-loop* region was by far the most variable and its
284 corresponding alignment was the only one comprising gaps (gaps were absent in both *COI*
285 and *COII* alignments).

286 ***Phylogeographic inferences***287 *C. rotundiventris*

288 No convincing topology could be retrieved using the Bayesian inference approach:
289 independent runs did not converge, even though trials with different MCMC parameters,
290 models of evolution, and among-partition linkage were performed. As a consequence,
291 Bayesian topologies were not taken into consideration for *C. rotundiventris*. In contrast, ML
292 and MP approaches provided partially congruent results, indicating that resolution is very low
293 in this taxon (Figure 2a), with extremely short branch lengths (ML) or polytomized tips (MP),
294 though about 5% of the nucleotides among the three regions were variable. When analyzing
295 patterns of the haplotype network inferred for *C. rotundiventris*, some spatial structure could,
296 however, be recovered among the 44 haplotypes (of which eight were shared by several
297 samples and 36 were private to single samples) (Figure 2b): the most abundant haplotype
298 (red) was absent from Scandinavia, while the purple and yellow haplotypes were restricted
299 to this region. Other haplotypes were private to some populations, as the grey (south
300 Scandinavia), the blue (northern Poland) and the black (Balkans).

301 *C. lophota*

302 Unlike *C. rotundiventris*, Bayesian inference runs converged well, and all methods
303 (Bayesian, ML and MP) were fully congruent in terms of both topology and support of the
304 three main clades (Figure 3a). The same three groups were also retrieved in the haplotype
305 network analysis (Figure 3b), and relationships between clades were also congruent with the
306 topological analysis (i.e., haplotypes from blue and red groups presented closer relationships
307 -a maximum of three steps- than those relating them with the yellow clade). The distribution
308 of clades indicated a clear, geographically structured genetic distribution (Figure 3c). While
309 the yellow clade was restricted to eastern populations, the red clade was exclusive of the
310 southwestern Alps and the blue clade was present mainly in the western distribution and on
311 the northern edge of the Alps.

312 *C. dentifera*

313 The three phylogenetic approaches yielded similar topologies and node supports, except
314 in the MP approach, which harbored low Bremer supports (Figure 4a). Analyses retrieved
315 four main clades (blue, red, yellow and purple) supported by at least one method, and two
316 additional clans (*sensu* Wilkinson, McInerney et al. 2007) placed in polytomies (green and
317 orange). The haplotype network approach indicated the presence of three shared and 15
318 unique haplotypes (Figure 4b). Haplotypes were tightly interrelated, with a maximum of five
319 steps between the two more distant haplotypes. Some of the shared haplotypes matched
320 with clades defined by the phylogenetic approach (i.e., red clade), while others contained
321 only some part of the samples included in the clades (i.e., green and yellow). The distribution
322 of clades indicated a trend towards spatial genetic structure, with samples from several
323 clades restricted to particular regions (Figure 4c): the yellow clade spread over southern
324 Scandinavian, as well as several locations in the Alps; the blue and red clades were
325 restricted to Scandinavia, while the orange clan and purple clade were mainly present in
326 Alpine locations. The remaining green clan appeared present in all regions, but was the only
327 one observed in the Pyrenees and in the northeastern most populations.

328 ***Phylogeographic hypothesis testing***

329 Testing phylogeographic scenarios in fly species using the canvas of the plant genetic
330 structure could be done only in species for which Bayesian inference analyses converged
331 and yielded supported clades (*i.e.*, *C. dentifera* and *C. lophota*). Maximum Likelihood
332 Estimates (MLE) of theta values for *C. dentifera* and *C. lophota* were 0.000229 and
333 0.001486, respectively, providing N_e equal to 26'220 ($\mu = 1 \times 10^{-7}$) and 262'200 ($\mu = 1 \times 10^{-8}$) for
334 *C. dentifera* and to 14'860 ($\mu = 1 \times 10^{-7}$) and 148'600 ($\mu = 1 \times 10^{-8}$) for *C. lophota*. Calculated S
335 values for the empirical data were 24, for *C. dentifera* and 27 for *C. lophota*.

336 Phylogeographic scenario-testing indicated that while we could not statistically reject the
337 null hypothesis for *C. lophota*, at least when considering a mutation rate equal to 1×10^{-7} (p_{H0} -
338 $p_{H1} = 0.39$ and $p_{H0-H2} = 0.55$; Figure 5), this was not the case for *C. dentifera*, for which the
339 calculated S value fell completely within the simulated distributions under the assumption of
340 a phylogeographic structure based on the plant genetic structure, whatever mutation rate
341 was considered [$p_{H1} = 0.76$ ($\mu = 1 \times 10^{-7}$) or $p_{H1} = 0.39$ ($\mu = 1 \times 10^{-8}$) and $p_{H2} = 0.77$ ($\mu = 1 \times 10^{-7}$) or
342 $p_{H2} = 0.52$ ($\mu = 1 \times 10^{-8}$); Figure 5].

343

344 Discussion

345 Though the phylogeography of European organisms has been revisited several times in
346 the last two decades (e.g., Taberlet, Fumagalli et al. 1998; Hewitt 1999; Schmitt 2009), no
347 studies have treated the fate of organisms involved in specific and obligate interactions in a
348 framework of comparative phylogeography. In contrast to our hypothesis of congruence
349 among spatial genetic structures of the different actors of a plant-pollinator mutualism, the
350 three pollinator species studied here show different phylogeographic patterns, which are only
351 in part congruent with patterns found in the plant. While *Chiastocheta rotundiventris* had no
352 clear, geographically consistent genetic structure (haplotypes and topologies were not
353 congruent; Figure 2), *C. lophota* (Figure 3) and *C. dentifera* (Figure 4) presented well-
354 defined spatial genetic structures whatever method was used. This result is a first evidence

355 of the overall incongruence in the spatial genetic structure of the partners of this interaction,
356 despite their closely related ecologies.

357 *Chiastocheta rotundiventris* is a species that has long been considered as the “most
358 mutualistic” of all those interacting with *Trollius europaeus* (Pellmyr 1992). First, it has been
359 shown to visit flowers at the very beginning of the flowering period, thus being present when
360 the flower is extremely fresh and receptive (Pellmyr 1989) and laying only one egg per flower
361 head, meaning a lower price to be paid by the flower in terms of number of larvae to host
362 (Pompanon, Pettex et al. 2006). Among the three studied pollinators, *C. rotundiventris*
363 appears as the species with the lower spatial genetic structure, indicating high levels of
364 admixture between populations (Figure 2). This could be explained by long-distance
365 dispersal abilities of this species, or by a scenario of incomplete lineage sorting in which
366 colonization of the current distribution of this fly originated from one or a few neighboring
367 regions during glacial periods. Concerning the first idea, it is known that small insects can
368 sometimes exploit air currents to travel extremely long distances (Compton 2001); the
369 likelihood of this hypothesis could even be enhanced by the mountainous habitat of these
370 flies (in the southern part of their distribution at least), more prone to experience strong
371 winds. In Europe, an absence of spatial genetic structure has been also unraveled in other
372 cold-adapted organisms such as *Ligusticum mutellinoides* (Alvarez, Thiel-Egenter et al.
373 2009) or *Ranunculus pygmaeus* (Schönswetter, Popp et al. 2006).

374 However, if we consider the haplotypic approach applied in this study some indications
375 show that even though high admixture (*i.e.*, haplotypes are extremely interconnected) is
376 present in *C. rotundiventris*, some geographic grouping is possible (Figure 2b). The
377 distribution of these haplotypes sometimes presents some geographic structure, as it is the
378 case of the central European/South-Scandinavian yellow haplotype, or the isolated Balkan
379 black type. Probably this last case is the most clearly structured, since it is the only clade
380 which is both supported in the phylogenetic and haplotypic approaches. Finally, despite

381 being somehow geographically structured, all the remaining haplotypes are separated by
382 very small numbers of steps.

383 The incongruence between the haplotypic and topological approaches applied for *C.*
384 *rotundiventris* indicates that it is possible that some past geographic structure was present,
385 but that it is currently being blurred by interpopulational exchanges. It is also interesting to
386 note that even though it is currently impossible to test phylogeographic hypotheses based on
387 the topology obtained in this study, the distributions of some of these haplotypes (*i.e.*, yellow,
388 purple, black, red) are similar distribution to that of the host plant (Espíndola et al,
389 unpublished data). In the future, it would be worth testing more variable molecular markers
390 to more deeply investigate the genetic structure of this species. An approach based on next-
391 generation sequencing technologies might be a good option, as soon as this technology will
392 become available at a reasonable cost to phylogeographers.

393 Unlike *C. rotundiventris*, *C. lophota* shows a phylogeographic pattern with a clear and
394 supported geographic structure (Figure 3c), with all methods yielding congruent results. It is
395 thus probable that the different clades have been isolated for a long time, and that their
396 fragmented distributions started to come into contact only recently. This hypothesis is
397 suggested by the long branches separating eastern (yellow) and western (blue and red)
398 genetic groups (Figure 3a). This genetic structure has also been previously observed in
399 other arctic-alpine European organisms, as the butterfly *Erebia epiphron* (Schmitt, Hewitt et
400 al. 2006) or the gentian *Comastoma tenellum* (Schönswetter, Tribsch et al. 2004), which also
401 appeared structured with east, central and western genetic lineages, coming into contact in
402 the Alpine range. It is possible that this genetic characteristic is a consequence of the
403 presence of this species in mountainous environments (*C. lophota* presents a distribution
404 range which occupies only the southern European mountains), which in southern Europe
405 represent massifs isolated by large areas of low elevation plains, which could have
406 enhanced their isolation during long periods of time.

407 Finally, the spatial genetic structure of *C. dentifera* indicates some geographic structuring
408 (Figure 4c), especially in Scandinavia (red and blue groups), and part of the southeastern
409 (Tatra) distribution (purple). However, similarly to *C. rotundiventris*, some lineages are
410 widespread, as the green clan, which is present in all regions. Like *C. rotundiventris*, but to a
411 lesser extent, haplotypes appear interconnected, probably indicating current or recent
412 genetic exchange between populations.

413 If similarities had to be identified between the three species, we could mention that *C.*
414 *rotundiventris* and *C. dentifera* present –despite the reduced genetic structure of the first-
415 specific lineages which appear restricted to Northern Scandinavia (yellow in *C.*
416 *rotundiventris*, red in *C. dentifera*, Figures 2 and 4). Moreover, both harbor one genetic
417 group absent of northern Scandinavia (red in *C. rotundiventris*, yellow in *C. dentifera*;
418 Figures 2 and 4). Because *C. lophota* presented a clear, geographically structured genetic
419 pattern, it is difficult to identify any parallel between this species and the other two, which did
420 not present such a structure. It appears, nonetheless, that in all cases, the Alps acted as a
421 center of current genetic diversity, which can be explained either by *in situ* glacial survival
422 (Brochmann, Gabrielsen et al. 2003) or by postglacial colonization during demographical
423 contractions, as proposed by the paradigm of the multiple recolonization of the Alps
424 (Schönswetter, Stehlik et al. 2005).

425 Because the three species present very different spatial genetic structures, despite
426 sharing medium to high levels of mtDNA genetic diversity, it seems that they have endured
427 distinct demographic and dispersal processes. As a consequence, we argue that the
428 phylogeography of the plant does not seem to present congruencies with those of all fly
429 species with which it is ecologically associated.

430 ***The effect of the plant on the genetic structure of the mutualistic flies***

431 Statistical phylogeography is a relatively new field that provides the possibility to
432 quantitatively test phylogeographic hypotheses (Knowles and Maddison 2002). It is thus an

433 ideal tool to identify drivers of phylogeographic patterns, and in our case, it fits well our
434 questions related to congruence in the spatial genetic structures of interacting species.
435 Despite the fact that the different insect species are tightly related to the plant from an
436 ecological point of view, our results show that there is a high heterogeneity in the level of
437 congruence between each insect species and the plant spatial genetic structures.
438 Hypothesis testing revealed that while one species (*C. lophota*) does not present a genetic
439 structure likely to be driven by that of the plant (H_0 not rejected, Figure 5), this is not the case
440 of *C. dentifera*. This last species presents a genetic pattern that does not significantly differ
441 of what would be expected in a case of plant-insect phylogeographic congruence (both H_1
442 and H_2 could not be rejected, while the two null hypotheses were rejected; Figure 5).

443 In contrast to *C. lophota*, which presents a genetic structure that is largely explained by
444 geographic factors (Figure 2) and that is not explained by the genetic structure of the plant,
445 *C. dentifera* seems more tightly related to the demographic and dispersal dynamics of *T.*
446 *europaeus*. Considering this, it is interesting to note that our test could not differentially reject
447 any of the non null-hypotheses for *C. dentifera*, indicating either that the plant has probably
448 been shaping the genetic structure of the insect for longer than one glacial period. If this is
449 true, it is possible that the insect and the plant depend strongly on each other to assure
450 survival even under environmental changes, and that demographic events and variations
451 occur in a parallel manner for these species. This should however be tested in the future,
452 considering that because the plant interacts with several fly species, it is possible that it is
453 able to move more freely than the insects.

454 Coupling phylogeographic patterns of organisms that are ecologically associated is a
455 recent practice in the field of phylogeography. In this context, the case of *Fagus* and
456 *Epifagus* parasites is one of the unique examples in which a relationship between interacting
457 species has been demonstrated at a large scale (Tsai and Manos 2010). The present study
458 provides additional support to the idea that recent histories of interacting organisms are

459 partly related, confirming that some processes occurring at large spatio-temporal scales
460 (e.g., synchronous co-cladogenesis) are also relevant at smaller evolutionary scales.

461 However, the pollination system studied here informs on the heterogeneity of scenario
462 matching in species involved in tightly-related mutualistic interactions, demonstrating that it
463 is important that each interacting organism studied is considered independently of the
464 others. The example of *T. europaeus* and its specific pollinators is thus highly instructive
465 since in this case, despite closely related ecologies, flies present different demographic and
466 dispersal histories. It is however possible that because the niches of these species strongly
467 overlap, interspecific competition is high, and differentiation in terms of population dynamics,
468 dispersion capabilities and population sizes could thus be enhanced. This idea of escaping
469 from interspecific competition in the *Chiastocheta* species complex has also been
470 demonstrated in other ecologically focused studies, which have shown that different taxa
471 display differential oviposition patterns (Després and Jaeger 1999) and seed-feeding
472 behaviors (Pompanon, Pettex et al. 2006). It is thus possible that as one of the most basal
473 species in the group (Després, Pettex et al. 2002), *C. dentifera* still harbors demographic
474 and dispersal patterns similar to that of the plant, whereas derived taxa such as *C. lophota*
475 have experienced niche switches towards leading to different phylogeographic patterns.

476 **Conclusion**

477 In the present study we show that interacting species may not only present different
478 phylogeographic patterns, but also that forces shaping phylogeographic structures may differ
479 among species: the presence of the plant during distributional changes for *C. dentifera*,
480 geography for *C. lophota*, and eventually migration capabilities and geography for *C.*
481 *rotundiventris*.

482 In the future it would be worth testing these hypotheses on the plant genetic material,
483 based on the insects' genetic structures. Indeed, it is plausible that in the same way that we
484 could not reject the hypothesis of a genetic structure driven by the plant for *C. dentifera*, the

485 genetic structure of the plant could also be influenced by one or another fly species.
486 However, in order to do this, analytical bases should be first firmly established before
487 allowing us to test hypotheses based on dominant data issued of genomic fingerprinting (i.e.,
488 the spatial genetic structure of *T. europaeus* was retrieved using AFLP).

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- 661
- 662

663 Table 1 – Names, sequences, annealing temperatures and references of primers used for
 664 sequencing mitochondrial region in *Chiastocheta* spp.

| Region | Primer | Sequence | Annealing | Reference |
|--------|---------------------------|---------------------------------|-----------|--------------------------|
| COI | COI-2171 | TTG ATT TTT TGG TCA YCC NGA AGT | 52 | Després and Jaeger, 1999 |
| | tRNA ^{Leu} -3048 | TGG AGC TTA AAT CCA TTG CAC | 52 | Després and Jaeger, 1999 |
| COII | tRNA ^{Leu} -3023 | GAT TAG TGC AAT GGA TTT AGC TC | 52 | Després and Jaeger, 1999 |
| | COII-3683 | CCR CAA ATT TCT GAA CAT TGA CC | 52 | Després and Jaeger, 1999 |
| D-loop | TM-N-193 | TGG GGT ATG AAC CCA GTA GC | 55 | Simon et al. 1994 |
| | SR-J-14612 | AGG GTA TCT AAT CCT AGT TT | 55 | Simon et al. 1994 |

665

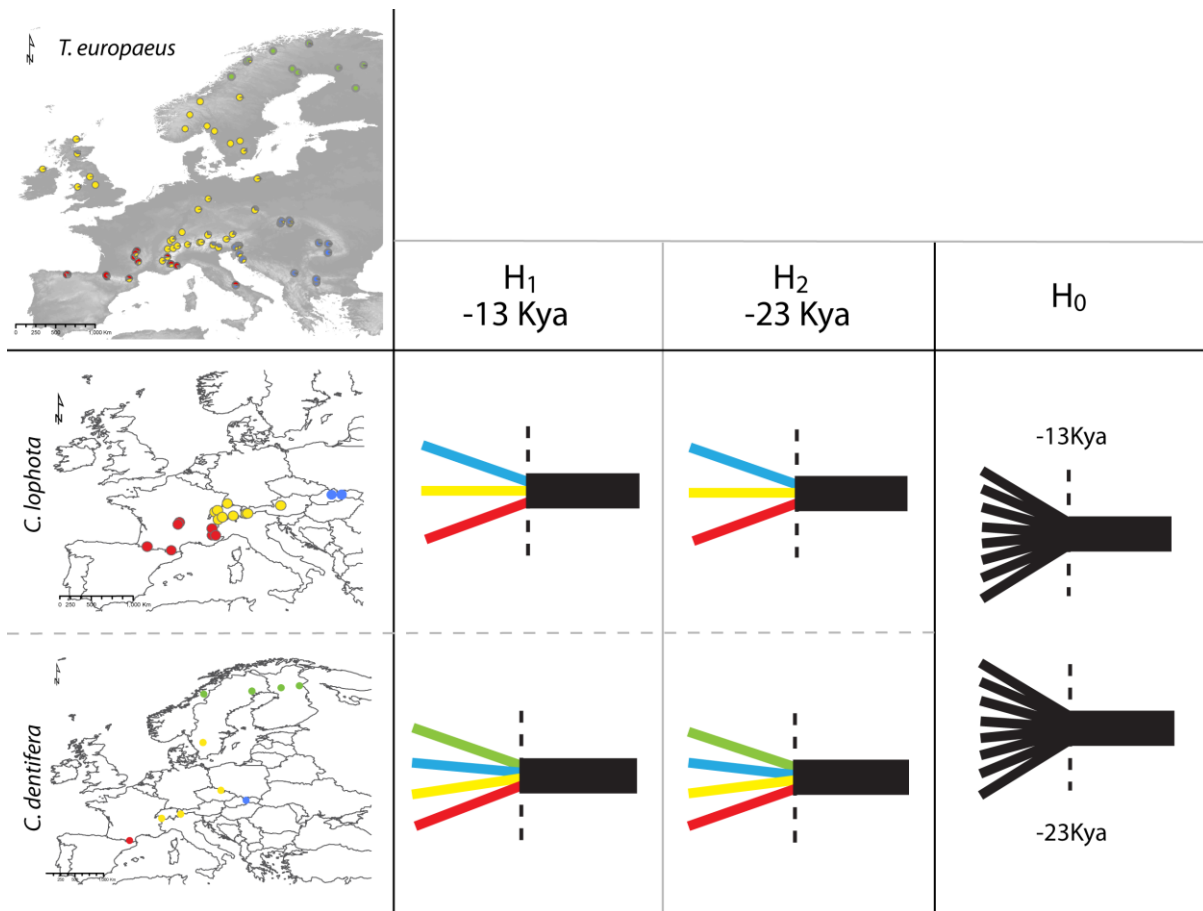
666

667 Table 2 – Number of total, constant, variable and parsimony informative sites (PI), as well
 668 as number of identified gaps per amplified region and species in *Chiastocheta*.

| Species | Region | Sites (base-pairs) | | | | Gaps |
|--------------------------|--------------|--------------------|-----------|-----------|-------------|-----------|
| | | Constant | Variable | PI | Total | |
| <i>C. dentifera</i> | COI | 653 | 4 | 1 | 657 | 0 |
| | COII | 475 | 4 | 0 | 479 | 0 |
| | D-loop | 883 | 32 | 15 | 1168 | 75 |
| | Total | 2011 | 40 | 16 | 2304 | 75 |
| <i>C. lophota</i> | COI | 644 | 13 | 6 | 657 | 0 |
| | COII | 470 | 9 | 6 | 479 | 0 |
| | D-loop | 768 | 60 | 29 | 855 | 62 |
| | Total | 1882 | 82 | 41 | 1991 | 62 |
| <i>C. rotundiventris</i> | COI | 645 | 12 | 4 | 657 | 0 |
| | COII | 457 | 22 | 5 | 479 | 0 |
| | D-loop | 865 | 56 | 27 | 1168 | 87 |
| | Total | 1967 | 90 | 36 | 2304 | 87 |

669

670



671

672

Figure 1 –Phylogeographic scenarios to be tested by coalescent-based phylogeographic

673

statistics, for *Chiastocheta lophota* and *C. dentifera*, considering the inferred genetic identity

674

of populations for *T. europaeus* based on Espindola et al. submitted. Maps for *C. lophota*

675

and *C. dentifera* show the position of sampled populations and the expected genetic identity

676

they should present if the genetic structure of the fly was congruent to that of the plant (map

677

on top). Fly populations included in each population tree branch are identified by the same

678

colors on the map and the phylogeographic scenarios. H₁= Phylogeographic scenario 1:

679

splitting occurred at the termination of the LGM (-13 Kya). H₂= Phylogeographic scenario 2:

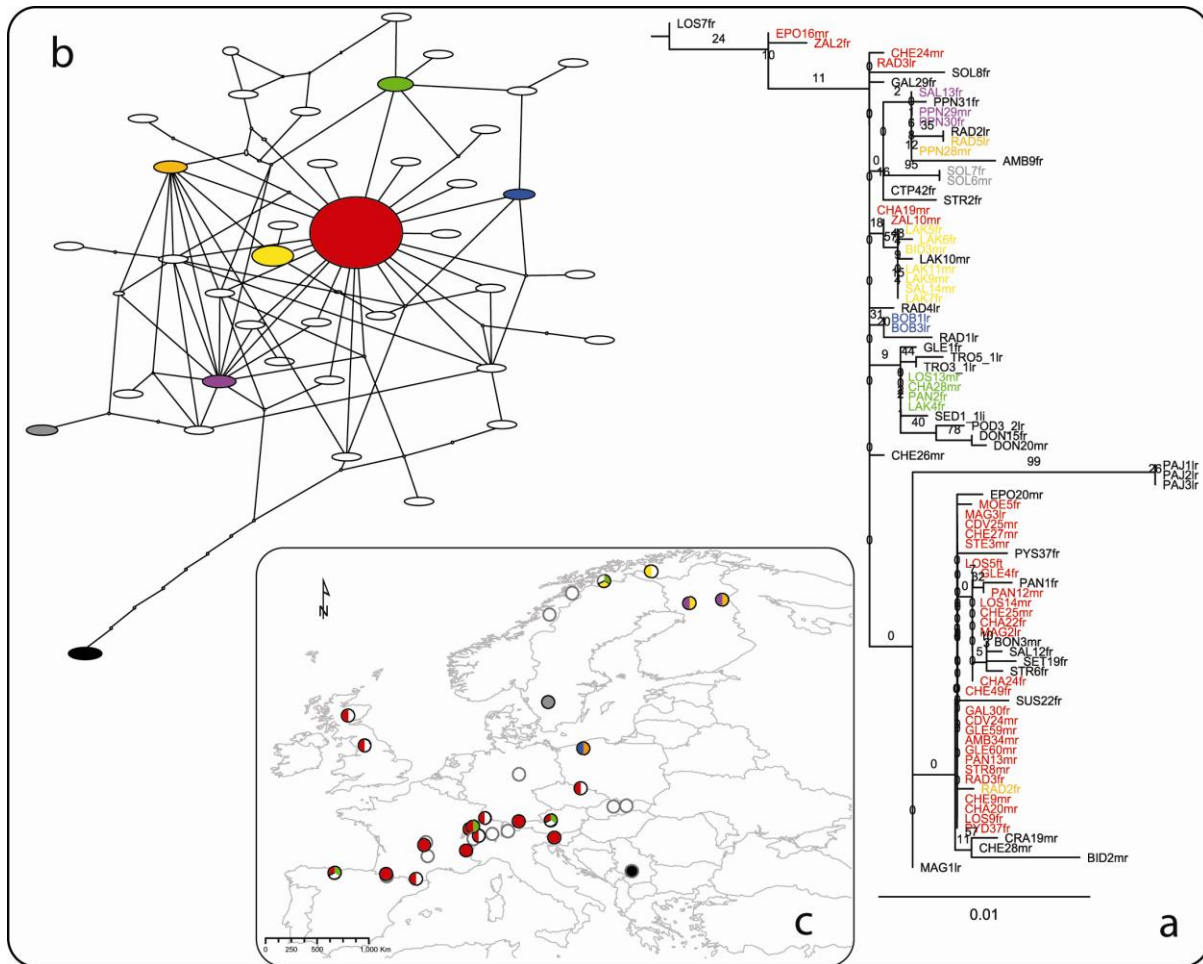
680

splitting occurred at the previous termination (-23 Kya). H₀= Null hypotheses for each of the

681

two first phylogeographic scenarios.

682

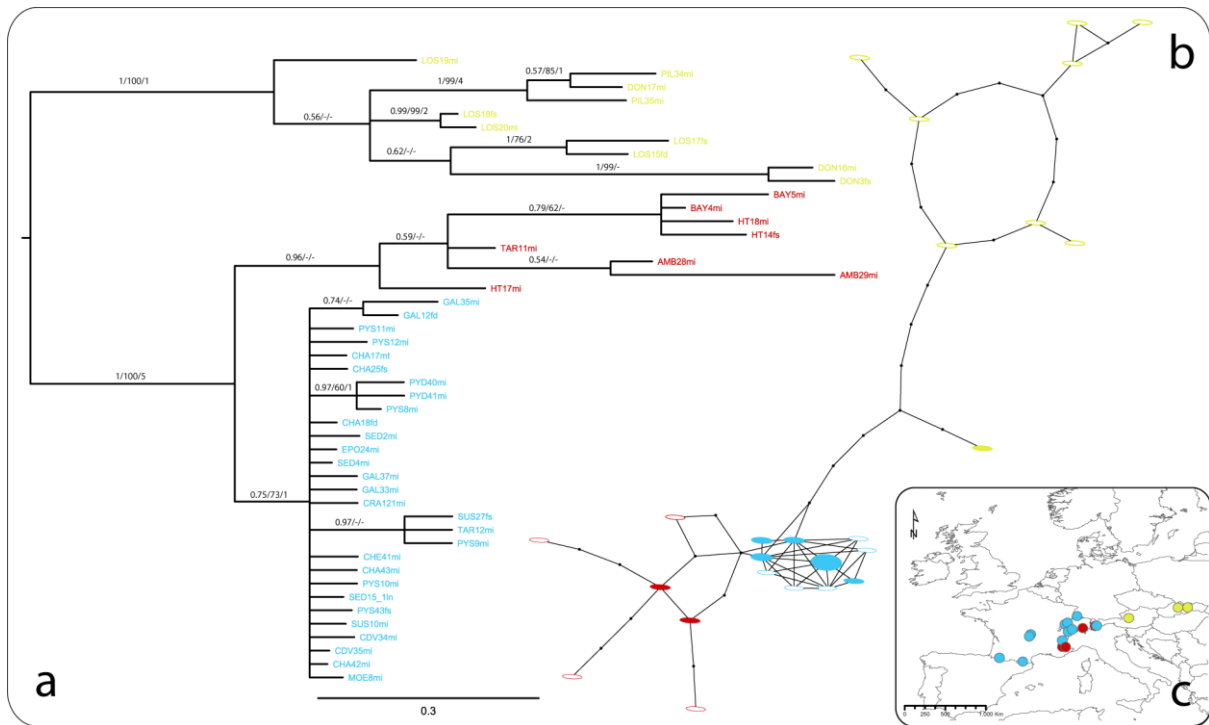


683

684 Figure 2 – Molecular inferences (a and b) and geographic distribution of haplotypes (c) in
 685 *Chiastocheta rotundiventris*. a- Maximum Likelihood (ML) phylogeny. Values on branches
 686 indicate ML supports considering 10,000 bootstraps. Colors indicate haplotypes (see b). b-
 687 Haplotype network. Colors show shared haplotypes; empty circles indicate unique
 688 haplotypes. c- Geographic distribution of haplotypes. Colors correspond to b. Sites where
 689 several colors appear indicate locations comprising different haplotypes.

690

691



692

693 Figure 3 – Molecular inferences (a and b) and geographic distribution of haplotypes (c) in

694 *Chiastocheta lophota*. a- Bayesian topological inference. Values on branches indicate

695 Bayesian supports, Maximum Likelihood supports and Maximum Parsimony Bremer

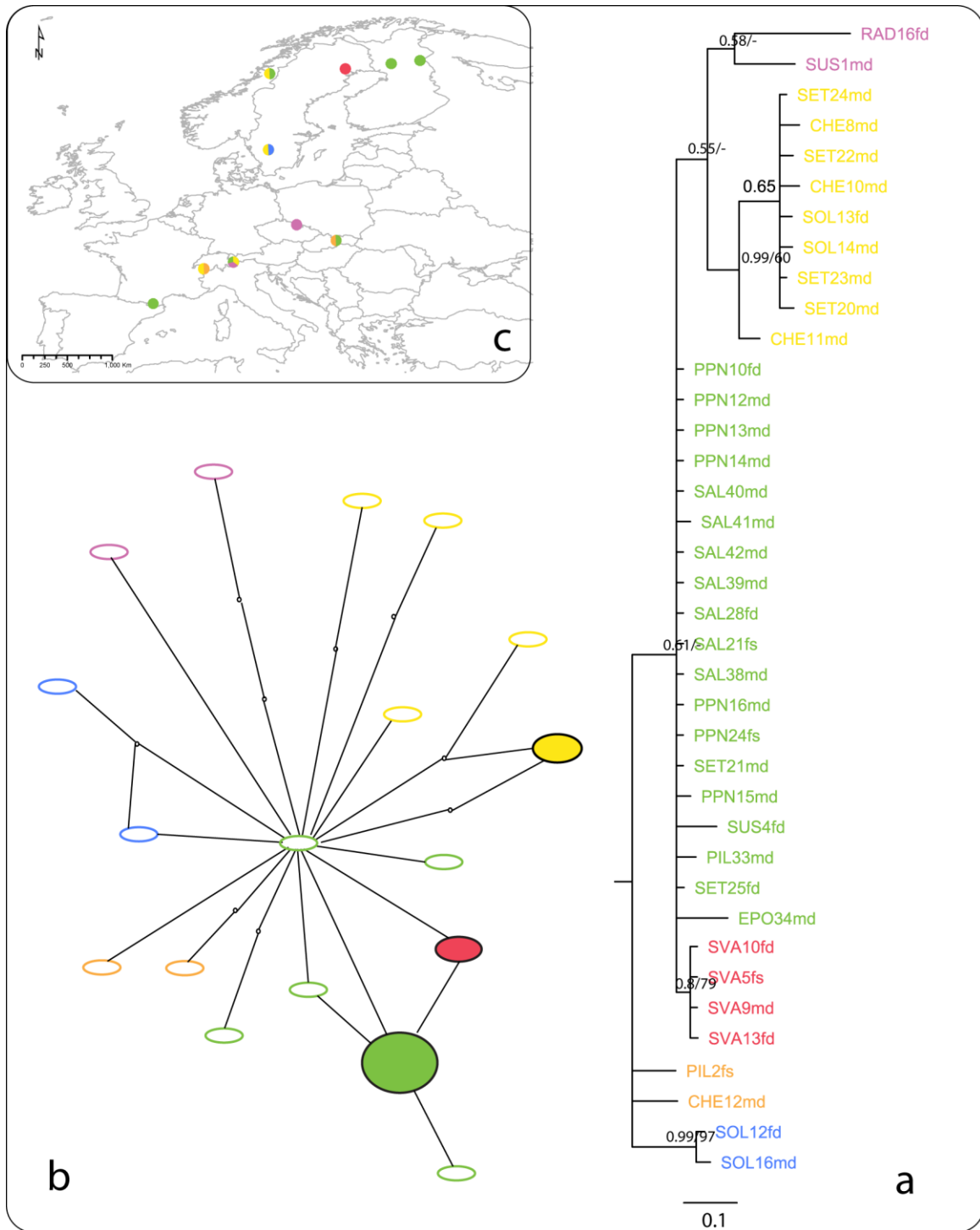
696 supports. Colors indicate main clades. b- Haplotype network. Colors indicate clades

697 identified in a. Empty circles indicate unique haplotypes; filled circles indicate shared

698 haplotypes. c- Geographic distribution of main clades. Colors correspond to a.

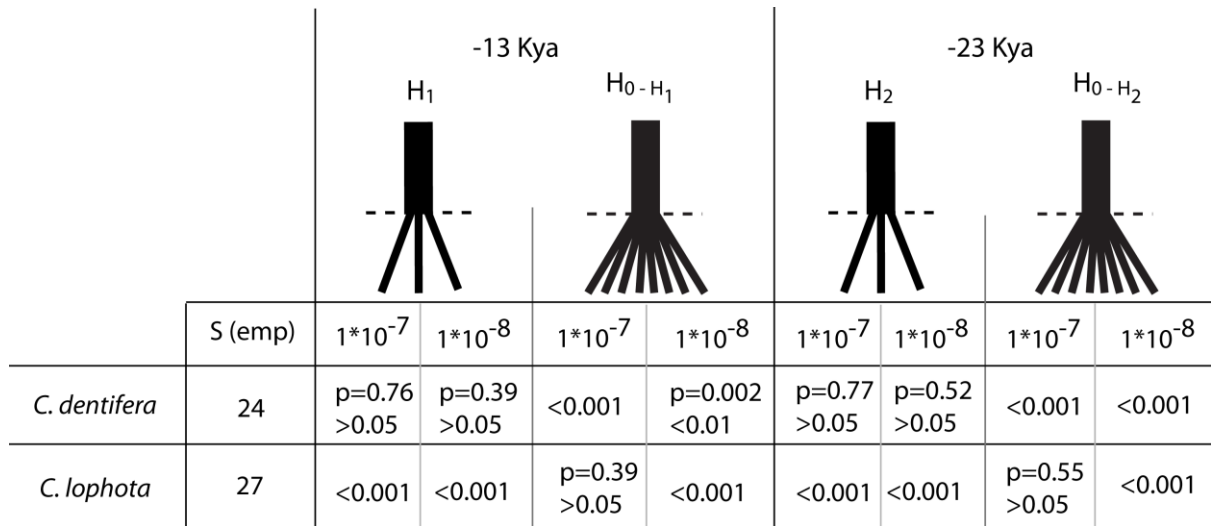
699

700



701

702 Figure 4 – Molecular inferences (a and b) and geographic distribution of haplotypes (c) in
 703 *Chiastocheta dentifera*. a- Bayesian topological inference. Values on branches indicate
 704 Bayesian supports and Maximum Likelihood supports. Colors indicate main groups. b-
 705 Haplotype network. Colors indicate clades identified in a. Empty circles indicate unique
 706 haplotypes; filled circles indicate shared haplotypes. c- Geographic distribution of clades.
 707 Colors correspond to a.



708

709 Figure 5 – Hypothesis testing for *Chiastocheta dentifera* and *C. lophota*. p-values (p) and

710 levels of significance are indicated for each species, scenario and mutation rate (μ)

711 considered. S(emp) indicates the calculated empirical S value for each dataset.

712

713 Supplementary Material 1 – Locations, identity and number of sampled flies.

| Location | Longitude | Latitude | Altitude (m) | Date | C. dentifera | C. lophota | C. rotundiventris |
|------------------------|-----------|-----------|--------------|----------|--------------|------------|-------------------|
| Ambri | 8.702920 | 46.506800 | 1000 | 30.5.08 | - | 2 | 2 |
| Amot | 8.42346 | 59.62199 | 481 | 18.06.07 | - | - | - |
| Bayasse | 6.740670 | 44.308140 | N/A | 9.06.07 | - | 2 | - |
| Beistohlen | 8.95473 | 61.20761 | 729 | 19.06.07 | - | - | - |
| Bidjovagge | 22.478080 | 69.297780 | 613 | 1.8.08 | - | - | 2 |
| Bobolice | 16.59895 | 53.94660 | 110 | 19.6.08 | - | - | 2 |
| Col de Bonnecombe | 3.11410 | 44.57557 | 1335 | 21.05.07 | - | - | 1 |
| Braas | 15.06817 | 57.09309 | 209 | 15.06.07 | - | - | - |
| Col de la Colombière | 6.469722 | 45.98722 | 1600 | 14.06.06 | - | - | - |
| Creux du Van | 6.741193 | 46.93526 | N/A | 29.06.06 | - | 2 | 2 |
| Chasseral | 7.021299 | 47.12569 | N/A | 29.06.06 | - | 5 | 5 |
| Chemin | 7.089778 | 46.08993 | N/A | 26.05.06 | 4 | 1 | 7 |
| Crans-Montana | 7.538896 | 46.34650 | N/A | 25.05.06 | - | 1 | 1 |
| Cressbrook Dale | -1.740410 | 53.267240 | 246 | 10.6.08 | - | - | - |
| Colt Park | -2.352470 | 54.193650 | 380 | 10.6.08 | - | - | 1 |
| Donovaly | 19.230680 | 48.889220 | 1345 | 21.6.08 | - | 3 | 2 |
| Eidda Pastures | -3.741900 | 53.037200 | 234 | 9.6.08 | - | - | - |
| Ellingsrudelva | 10.91844 | 59.91771 | 161 | 18.06.07 | - | - | - |
| Esposouille | 2.094500 | 42.623410 | 1521 | 3.6.08 | 1 | 1 | 2 |
| Froson | 14.60268 | 63.18205 | 424 | 20.06.07 | - | - | - |
| Col du Galibier | 6.438611 | 45.08528 | 2000 | 14.06.06 | - | 4 | 2 |
| Glen Fender | -3.794850 | 56.781380 | 345 | 14.6.08 | - | - | 4 |
| Haute Tinee 1 | 6.818710 | 44.296170 | 2020 | 4.06.07 | - | 3 | - |
| Haute Tinee 2 | 6.855810 | 44.284260 | 1770 | 4.06.07 | - | - | - |
| Krasno Polje | 14.97271 | 44.808690 | 1487 | 9.7.08 | - | - | - |
| Laktatj akka | 18.40674 | 68.42931 | 453 | 22.06.07 | - | - | 7 |
| Lough Fern | -7.711300 | 55.065690 | 26 | 13.6.08 | - | - | - |
| Loser | 13.78485 | 47.66052 | 1598 | 04.06.07 | - | 5 | 5 |
| Col long de Magnabaigt | -0.43646 | 42.87060 | 1615 | 23.7.08 | - | - | 3 |
| Moerlimatt | 8.07760 | 47.90597 | 939 | 10.06.07 | - | 1 | 1 |
| Monte Pizi | 14.167140 | 41.915240 | 1546 | 28.5.08 | - | - | - |
| Naverdal | 10.13002 | 62.70417 | 480 | 19.06.07 | - | - | - |
| Pajino Preslo | 20.819700 | 43.277990 | 1802 | 8.7.08 | - | - | 3 |
| Puerto de Panderruedas | -4.972230 | 43.127430 | 1227 | 4.6.08 | - | - | 4 |
| Pila | 20.294490 | 48.900170 | 972 | 21.6.08 | 2 | 2 | - |
| Podlesok | 20.351900 | 48.949620 | 579 | 21.6.08 | - | - | 1 |
| petit papa noel | 25.79386 | 66.51647 | 72 | 23.06.07 | 7 | - | 4 |
| Puy de Dome | 2.963333 | 45.77222 | 1460 | 15.06.06 | - | 2 | 1 |
| Puy de Sancy | 2.809722 | 45.53500 | 1520 | 16.06.06 | - | 6 | 1 |
| Radkow | 16.353210 | 50.468660 | 712 | 20.6.08 | 1 | - | 7 |
| Risnjak - Snjeznik | 14.584940 | 45.438710 | 1466 | 5.06.08 | - | - | - |

| | | | | | | | |
|------------------|-----------|-----------|------|----------|---|---|---|
| Salla | 28.65427 | 66.83020 | 194 | 23.06.07 | 7 | - | 3 |
| Sede de Pan | -0.486510 | 43.039490 | 1556 | 22.7.08 | - | 3 | 1 |
| Seterasen | 13.67744 | 65.53432 | 285 | 21.06.07 | 6 | - | 1 |
| Solberga | 13.56116 | 57.95194 | 239 | 15.06.07 | 4 | - | 3 |
| Steingaden | 11.01296 | 47.59529 | 1158 | 30.05.07 | - | - | 1 |
| Straumen | 15.64921 | 67.38440 | 75 | 21.06.07 | - | - | 3 |
| Susch | 10.074725 | 46.747277 | N/A | 05.06.06 | 2 | 2 | 1 |
| Svartla | 21.22062 | 65.99583 | 36 | 22.06.07 | 4 | - | - |
| Tarasp | 10.250556 | 46.777299 | N/A | 05.06.06 | - | 2 | - |
| Trollblumenwiese | 11.04118 | 51.68314 | 488 | 16.6.08 | - | - | 2 |
| Vitosha | 23.293420 | 42.590320 | 1779 | 3.7.08 | - | - | - |
| Zali Log | 14.110800 | 46.203420 | 533 | 17.5.08 | - | - | 2 |

714

General Discussion

Because of the diverse nature of topics, approaches and conclusions addressed in the eight chapters of my thesis, I have chosen to group milestones of my research into the three following themes: i) the species concept in a (co)evolutionary framework; ii) the biology of interactions; iii) the phylogeography of interactions.

Finally, in order to have an insight on the advances and questions opened by this work, I present a small review on the main topics achieved considering each biological system separately. I hope that this approach will not only help resume the work done, but also present some of the new paths that research should take if future work on each system is to be performed.

The species concept in a (co)evolutionary framework

When studying coevolutionary processes it is important to identify which components are particularly driven by selective forces. This aspect has been widely discussed and is intimately related to the species concepts (Wheeler and Meier 2000). Several chapters of this thesis focus on the use of comparative approaches to corroborate and clarify the delimitation of taxonomic groups currently under discussion. Even though involving unrelated groups, the cases of *Arum* (chapter one), *Chiastocheta* (chapter six) and Psychodinae (chapter two) are similar in the approaches used to delimit taxonomic units. In the three chapters, we have demonstrated that the complementary use of different techniques (morphology, molecular phylogeny and spatio-temporal analyses) allows a better understanding and definition of the circumscription and evolutionary history of taxonomic units.

For instance, we have shown that the definition of species in *Arum* is not entirely supported by molecular analyses, indicating that some revision should be undertaken in this genus. Our study has also demonstrated that there are important geographic and historical components in the diversification and relationships among lineages within this genus, a

scenario that was formerly proposed by some authors on the base of distributional evaluations (Hruby 1910).

In the case of species within *Chiastocheta* found to interact with *Trollius europaeus*, limits of morphological species are not always confirmed using molecular markers. Despite the existence of morphological variation between types, some of them frequently hybridize or share considerable ancestral polymorphism. It is thus highly likely that what we currently observe is the result of an ongoing speciation process, in which barriers to hybridization are still not completely defined. As mentioned by de Queiroz (2005), our study thus points out the fact that the main problem in the discussion of the species concept is related to the fact that different techniques evaluate entities formed at different timescales. In this study, we demonstrate that depending of the criterion applied (morphological, mating isolation, molecular-based phylogenetic monophyly), different supported entities are identified. Our study provides a first large-scale survey of the relationships between lineages within this fly group, considering not only their genetic identity, but also their distribution and possible gene flow among species.

Finally, the case of Psychodinae occurs at a higher taxonomic level. Here, we demonstrated that despite their relatively similar morphologies, the subfamily Psychodinae is extremely old (up to 85 My). We also showed that only some aspects of their current systematics (*i.e.*, some subtribal entities, some genera) are confirmed by molecular analyses. Moreover, it appears that in this group most species lineages originated in the Paleogene, shortly after the K-T massive extinction event (Erwin 2001; Jablonski 2001).

These three case studies have in common the fact that taxonomic circumscriptions of entities treated in them were problematic. In all of them, the diverse approaches applied helped open up the possibility to discuss concepts in a different framework than is generally employed (e.g. morphological). The more comprehensive approach we used presents thus new options to explain these complex relationships.

In the framework of the species concept discussion, I hope that the work done here will at least partially help provide arguments to demonstrate that the debate over correctness of one concept over another is not necessarily constructive. Each concept has problems and

advantages, and probably the biggest potential lies in exploring and combining them thoughtfully, using each in ways appropriate to the organisms under study. A good demonstration of this may be the fact that within these chapters, any approach by itself (e.g., molecular, morphological, biogeographic) was able to independently provide convincing propositions in regard to the systematics of the discussed groups, whereas a combination of these approaches brought new light to the boundaries of taxonomic units. Moreover, one of the most exciting points about complementary analyses is that they allow us to position ourselves at different evolutionary and temporal standpoints, to have a deeper understanding of the speciation process.

Ecology and biology of interactions

Coevolutionary processes are characterized by their dynamic nature: species are continuously adapting to each other in order to increase their respective fitness gains (Bronstein, Dieckmann et al. 2004). Despite the fact that coevolutionary processes involve a defined number of species (Ollerton, Killick et al. 2007), organisms participating to the interaction are influenced both by their abiotic environment and by pressures exerted by the other interacting organisms. Investigating the effects of abiotic and biotic aspects on the dynamics of biological and ecological adaptations was the aim of chapters three and four.

First, the Theory of the Geographic Mosaic of Coevolution (TGMC) predicts that in a variable environment, biotic interactions are susceptible to change (Thompson 2005). Taking this idea into account, I demonstrated in chapter three that there is an effect of climatic variation on the identity of potential pollinators in the antagonistic interaction represented by *A. maculatum* and its Psychodid pollinators. As predicted by the theory, it appears that the plant switches pollinators according to clinal changes in precipitation and temperature. Besides confirming the TGMC, this result demonstrates the importance of large-scale studies when working with specific interactions and warns against assuming general laws for processes which occur locally and are punctually confirmed.

In the same way that the environment can influence the nature of a given interaction at a large spatial scale, theory predicts that interactions can force associated species to maximize their benefits from the ecological exchange (Dufaÿ and Anstett 2003). For instance, plants maximize fitness by adapting their reproductive strategy based on the type and efficiency of their pollinators. In chapter four, we showed that the identity and diversity of pollinators appears to shape the reproductive strategy in a series of closely related plants of the genus *Arum*, which all present entomophilous lure-and-trap pollination systems. In this chapter, we test a simple model to help clarify our understanding of the relationship of the reproductive strategy represented by the Size-Advantage Model (Ghiselin 1969) and the type of pollinators a plant is associated.

Probably the most important demonstration of these chapters is that coevolutionary interactions should not be seen as static, even if they appear to be extremely specialized. Variations in the environment or in the respective adaptation of partners can modify selective forces and thus redirect selective pressures, either locally or along the whole geographical range of an organism.

Phylogeography of interactions: understanding interactions in time and space

The field of phylogeography is an integrative field *per se*, since it combines molecular phylogenetic inferences with spatial information (e.g., geographical information systems, spatio-temporal distribution of species; Hickerson, Carstens et al. 2010).

In this context, the comparative phylogeography of interactions goes further by integrating hypotheses that explain phylogeographic patterns of species interacting ecologically and thus integrating ecological factors into the analysis of the evolutionary history of species.

In the same way that understanding the history of species is important for recognizing the effect of external events (e.g., climatic fluctuations, orogenic processes) on their survival (Hewitt 1996), comparing phylogeographic patterns in ecologically related species informs us about the influence that the interaction itself had on the history of species (Tsai and

Manos in press). Knowing the effect one species could have on another thus allows the quantification of the interdependency of species in a temporal framework.

In such a context, in chapters five, seven and eight, I studied the comparative phylogeographies of species involved in two European, obligate and specific interactions. Considering the specific and obligate nature of the relationships, I initially expected that while mutualistic interactions would be prone to share similar (congruent) phylogeographic histories, this would not be true for antagonistic interactions.

Our results partially confirmed these working hypotheses, since antagonistic organisms presented incongruent phylogeographic patterns (chapter five), while some of the interacting species involved in the mutualistic interaction presented congruent phylogeographies (chapter eight).

In chapter five, the finding of incongruence between antagonistic species could be related to differences in generation times and dispersion capabilities between the two partners [as has been proposed by Borer *et al.* (submitted)]. It is interesting to note that the phylogeographic structure demonstrated by the plant is extremely similar to that presented by the composition of flower visitors (chapter three). If the current insect composition has been brought about by the last glacial period, it is possible that the plant genetic structure has been shaped by adaptation to different fly species, which would have been hosted in different glacial refugia.

In chapter eight, our results on the mutualistic interaction between *T. europaeus* and its *Chiastocheta* pollinators showed that while species are similar from an ecological point of view, their evolutionary histories appear to be different. This could be a consequence of interspecific competition between species exploiting similar resources. This struggle could have induced different taxa to develop diverse dispersal capabilities and developmental features to displace their respective ecological niches and avoid competition. Our results thus demonstrated that while the genetic pattern of one species (*i.e.*, *C. lophota*) appeared to be strongly related to geographic features corresponding to phylogeographic patterns already identified for other Alpine species (e.g., Schönswetter, Stehlik *et al.* 2005), the other

two presented almost no spatially defined genetic structure (*C. rotundiventris*), or had a structure which appears to have been driven by the host plant (*C. dentifera*).

While a wide range of causes appeared to explain the phylogeographic patterns of *Chiastocheta* flies, the plant's spatial genetic structure is more likely explained by its distribution at the Last Glacial Maximum (LGM): there is hence a strong effect of the abiotic component in the definition of the plant structure. In the future, it would be interesting to test the plant structure using phylogeographic statistical techniques similar to those used for the insects. This is, however, currently technically impossible considering the type of genetic data (dominant) we used.

These three chapters demonstrate that complex phylogeographic questions, such as those involving congruence testing in the context of interacting species, might require going further than just visually quantifying similarities between distribution maps of genetic diversities. Today, an array of new techniques and technologies is available to biologists, and the potential for understanding the history of evolutionary processes in space is enormous. In this thesis for example, the integration of several areas of expertise has allowed the development of a new technique to infer past demographic contractions in cold-adapted species (chapter seven), and the direct testing of phylogeographic scenarios applied in chapter eight. Without such interdisciplinary work, none of the conclusions obtained in those surveys would have been possible.

State of the knowledge on each biological system

***Arum maculatum* L. – Psychodid flies**

Achievements attained in this thesis allowed a new understanding of evolutionary and biological topics of this antagonistic pollination relationship.

From the plant evolutionary point of view, we could demonstrate that the taxon previously considered as *A. maculatum* should be redefined in the future. Our study (chapter one) indicated that not one but two phylogenetic entities are currently being considered as belonging to this species. While one of them appears restricted to the European part of the

distribution of the plant, individuals from Anatolia seem to belong to another taxon. It appears necessary that these two entities are further investigated to clearly define their taxonomic boundaries. Moreover, whereas the Anatolian entity appears as monophyletic, this is not the case of the European taxon, whose specimens are included into a wide polytomy comprising other *Arum* such as *A. cylindraceum*. In the future, investigating species definition using more variable markers should allow addressing the monophyly of *A. maculatum*.

From the biological point of view, we demonstrated that the number and identities of pollinating flies appear to be geographically structured throughout the European part of the distribution range of the plant (chapter three). While *Psychoda phalaenoides* L. is the dominant visitor in central and northern Europe, populations in southeastern regions are dominated by *Psycha grisescens* Tonnoir. This result is of main importance because it demonstrates that even in highly specialized biological systems as the one studied here, variation in terms of abundances and identities of visitors is also expected when the distributions of interacting species cover large spatial ranges. This study also points out the importance of having a clear view on the natural history of the system studied before further investigating other more complex evolutionary questions.

The main advances done at the phylogeographic level are that even though the plant has a clear geographic genetic structure, this is not the case of the pollinators (chapter five). Despite this incongruence at the level of intraspecific lineages, the spatial structuring of the pollinator compositions is similar to the plant genetic structure, what would indicate some correlation between these two components. It is possible that differences in insect compositions are currently shaping the genetic structure of the plant through differential gene flow defined by the two pollinator groups. Another option could be that plants have been adapting to the two types of visiting insects through glacial times and that what we currently see is the result of selective and adaptive processes. Because plants attract insects by odour and heat production, it would be interesting in the future to investigate the volatile compositions of plants belonging to different regions. If adaptive processes were

happening, we would expect that plants from the two regions would produce different volatiles, more adapted to the attraction of the corresponding insect species.

At the same time, and in order to be sure that the flower chambers compositions do not reflect only the species availabilities in the environment, general insect trapping surveys should be done at the local level. We could thus understand with more accuracy the variation and compositions of insect availability in the two different environments that these plants occupy.

***Trollius europaeus* L. – *Chiastocheta* species complex**

Probably one of the main advances on this mutualism was that thanks to our large-scale sampling strategy, we could provide a sight onto the insect visitation rates throughout a large part of the distribution range of the plant (chapter six). This approach allowed us to observe that while some species currently occupy the whole European distribution of the plant, this is not the case of all of them. However, by using molecular techniques to identify lineages and congruence between molecular and morphological approaches, we showed that even though some species are clearly genetically circumscribed, this is not the case of all of them, with especially four species presenting relatively high levels of genetic admixture. Besides demonstrating that the species boundaries in these closely related set of species are difficult to define, this study warns researchers on conclusions that could be obtained if the species are considered to be clearly isolated entities. From this point of view, it should be important in the future to take these results into account, especially if working on the evolutionary and ecological dynamics of those admixed entities.

From a phylogeographic point of view, we demonstrated that the genetic structure of the plant appears to have been affected by the climatic variations related to the end of the last glacial period (chapter seven). From the fly side, only one species (*C. dentifera*) presented a phylogeographic structure similar to that of the plant (chapter eight), what could indicate that this species is more dependent of the interaction than the others, probably because it is the one visiting plant patches at the end of the flowering period.

The next steps in the understanding of this system would need a deeper survey on the detailed life-cycles and dispersion capabilities of the different interacting species. It would be moreover also interesting to investigate the interspecific gene flow among the different fly morphospecies.

From the fly community point of view, it would be important to study the behavior of those species present at locations in which most of the other taxa are lacking. It could be expected that because of differences in fly interspecific competition pressures at each location, taxa might behave differently when their relative diversities vary.

Finally, from a taxonomical point of view, it appears necessary to redefine the boundaries of the two species appearing to present the highest rates of interbreeding (*C. setifera* and *C. inermella*). For this, collaborations with taxonomists are highly recommended, since their knowledge could largely help find informative morphological characters.

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General Conclusion

The development of different approaches in the biological sciences has made this area extremely dynamic. The diversification of studies has dramatically increased the complexity of topics investigated, but has opened exciting new doors towards the establishment of interdisciplinary studies.

This thesis has exploited such integration of approaches, by basing hypotheses and analyses on different ecological, evolutionary, phylogeographic and historical aspects of interactions, while trying to incorporate the knowledge accumulated in different areas of study in biology. As demonstrated in this work, this is useful not only for the development of new analytical approaches, but also for cases of taxonomic delimitations, especially when incongruence between different types of data is present.

Thanks to such approaches, we are able to answer to the main question of this thesis: do interacting partners present similar histories? The answers were yes for some mutualistic and no for antagonistic relationships. However, in both cases, we demonstrated that relationships between interacting species are not as simple as previously thought, and that the simultaneous and complementary analysis of different aspects can help enhance our view of interactions.

I am convinced that the future of biological research relies on collaboration, exchange and communication between researchers working on different topics. In the same way that this thesis has benefited of interdisciplinary work and scientific flow among researchers and institutions, numerous are the areas of evolution and ecology which should take advantage of this to investigate and understand the complexity of life.

Appendix 1
Curriculum vitae

CURRICULUM VITAE

Espíndola, María Anahí
Chemin des Liserons 9
2000 Neuchâtel
Switzerland
Phone: +41 (0)32 724 4779
maria.espindola@unine.ch

Biologist
28 years-old
married
Argentine

Studies

- 2006 - 2010 PhD thesis at the E-vol Laboratory of the University of Neuchâtel, Switzerland: "Inferring reciprocal evolutionary histories in associated species of plants and insects in two European pollination systems", under the direction of Prof. Martine Rahier and Dr. Nadir Alvarez.
- 2002 - 2006 Biology, diploma at the E-vol Laboratory of the University of Neuchâtel, Switzerland. Diploma: "Interactions between *Horismenus butcheri* Hansson & Aebi (Hymenoptera: Eulophidae) and two *Phaseolus* (Fabaceae) bean species in Central Mexico. Biology, population genetics and geospatial studies", under the direction of Dr. Betty Benrey.
- 2000 - 2001 Biology, at the National University of Córdoba, Argentina.

Courses

- January-March 2010 "Computational Phylogenetics and Phylogeography". Prof. Bryan C. Carstens. Louisiana State University, USA.
- 13-28
May, 2009 "An introduction to the Practice of Statistics using R". Dr. Thomas Gsponer. NCCR Doctoral Program. University of Neuchâtel, Switzerland.
- 10-14
September, 2007 "Phylogénie et évolution moléculaire". Dr. Juan Montoya and Dr. Jan Pawlowski. Departement of Zoology and Animal Biology. University of Geneva, Switzerland.
- 9-11 October, 2006 "Population genetic data analysis". Dr. Felix Gugerli and Dr. Rolf Holderegger. Plant Science Center Zürich. ETHZ, Switzerland.

Conferences

- 13-16
October 2009 Poster. DIVERSITAS Open Science Conference 2. "Biodiversity and Society: Understanding connections, adapting to change", Cape Town, South-Africa.
- 8-10
July, 2009 Oral communication. "Biogeography of *Arum maculatum* pollinators". Xth International Aroid Society Conference . Nancy, France.
- 8-10
July, 2009 Poster. "Pollination biology of *Arum cylindraceum*". Xth International Aroid Society Conference . Nancy, France.
- January 30 –
February 1, 2008 Participation at the "Plant Species Concepts and Evolution": NCCR Plant Survival Conference. University of Neuchâtel, Neuchâtel, Switzerland.
- 17-21
September, 2007 Poster. "Population and landscape genetics of a Mexican parasitoid". Xth European Workshop on Insect Parasitoids, Erice, Italy.

Publications

Espíndola A., Buerki S., Bedalov M., Küpfer P. and Alvarez N. 2010. New insights into the phylogenetics and biogeography of *Arum* (Araceae): unravelling its evolutionary history. *Botanical Journal of the Linnean Society*. 163, 14–32.

Espíndola A., Pellissier L. and Alvarez N. *in press*. Variation in the proportion of flower visitors of *Arum maculatum* along its distributional range in relation with community-based climatic niche analyses. *OIKOS*. doi: 10.1111/j.1600-0706.2010.18937.x

Revel N., Alvarez N., Gibernau M. and **Espíndola, A.** *In preparation*. Size-advantage model and pollination strategies in plants. To be resubmitted to *Evolutionary Ecology*.

Espíndola A., Pellissier L. and Alvarez N. *In preparation*. No fossils, no answers? Inferring postglacial genetic consequences in cold-adapted species when neither ancient DNA nor macroremains are available.

Espíndola A. and Alvarez N. *In preparation*. Comparative phylogeography in a specific and obligate pollination antagonism.

Espíndola A., Buerki S., Jacquier A., Ježek J. and Alvarez N. *In preparation*. Molecular relationships in the sub-family Psychodinae (Diptera: Psychodidae). To be submitted to *Molecular Phylogeny and Evolution*.

Espíndola A., Buerki S. and Alvarez N. *In preparation*. Evolutionary history of *Chiastocheta* species interacting with *Trollius europaeus*.

Espíndola A., Carstens B. C. and Alvarez N. *In preparation*. Phylogeography of mutualists and the effect of the host on the genetic structure of its partners.

Espíndola A., Revel N. and Alvarez N. *In preparation*. Phylogeography of *Arum cylindraceum*.

Triponez Y., **Espíndola A.**, Bassin L., Pellissier L. Alvarez N. *In preparation*. Large biogeography of an oil collecting mutualism: tight interactions involve shared refugia but independent dispersal of partner species.

Internships

15.08.05-09.09.05 Study of gene flow between different species of the genus *Aegilops* in Europe, using RAPD markers: "*La génétique de populations d'Aegilops neglecta Req. et Bertol., A. cylindrica Host, A. triuncialis L. et A. geniculata Roth dans quatre pays d'Europe*". Laboratoire de Botanique Evolutive. University of Neuchâtel. Under the direction of Dr. Roberto Guadagnuolo.

26.07.04-10.09.04 Primer creation and sequencing of genes involved on species isolation in *Drosophila montana*: "Mating song evolution of *Drosophila montana* (Diptera: Drosophilidae). A molecular analysis of the evolution of the cacophony (*cac*) gene". Laboratory of Biology and Evolution, the University of Leeds (UK). Under the direction of Prof. R. K. Butlin.

Teaching

2007 - 2009 Bachelor classes at the University of Neuchâtel: "Ecologie Evolutive", "Méthodes Quantitatives en Ecologie" and "Biologie des Invertébrés".

Master classes at the University of Neuchâtel, Master of Physiology and Ecology of Plants (PEP): "Classics in Biology".

Field experience

April – August, 2007

April – August, 2008 Field trips covering the European continent (Great Britain, Scandinavia, Balkans, Central Europe, Italy and Spain).

26.01.2005-26.02.2005

31.12.2005-8.02.2006 Field trips in Central Mexico.

Prizes

October, 2006

Jean Landry Prize, for a mean of 5.73 at the end of the studies.

Grants

- 2009-2010 *Fonds des Donations* of the University of Neuchâtel, to get training on methods in phylogeographical statistics at the Laboratory of Bryan C. Carstens, Department of Biological Sciences, Louisiana State University, USA.
- April – June
2008 SCNAT+ funds for field work, to carry out the sampling of *Arum cylindraceum* and its pollinators throughout its South-East European distribution
- July – September
2004 Funds Wüthrich - Matthey-Dupraz; for an internship at the Laboratory of Prof. R. K. Butlin, Leeds, UK.

Informatics

- Wide experience with Geographic Information Systems: ArcGIS and ArcView
- Good knowledge in exploration of Biodiversity and geographic databases: GBif, EUNIS, several European local databases, European soils database.
- Good experience in the use of population genetics and phylogenetics softwares: e.g., STRUCTURE, Geneland, MrBayes, F-stats, Arlequin, TreeFinder, MrAIC, PAUP*, Mesquite.
- Good knowledge in the use of graphic treatment of images (Photoshop and Illustrator).

Languages

- Spanish: native speaker.
- English: very good written and spoken knowledge (Upper Intermediate Level).
- French: excellent written and spoken knowledge ("Accès au DALF" certificate + 8 years of studies at the University of Neuchâtel, Switzerland)
- German: good spoken (Swiss-German) and some written knowledge.