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Combinatorial chemistry in nematodes: modular assembly of primary metabolism-derived building blocks

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Covering: up to 2015

The nematode *Caenorhabditis elegans* was the first animal to have its genome fully sequenced and has become an important model organism for biomedical research. However, like many other animal model systems, its metabolome remained largely uncharacterized, until recent investigations demonstrated the importance of small molecule-based signalling cascades for virtually every aspect of nematode biology. These studies have revealed that nematodes are amazingly skilled chemists: using simple building blocks from conserved primary metabolism and a strategy of modular assembly, *C. elegans* and other nematode species create complex molecular architectures to regulate their development and behaviour. These nematode-derived modular metabolites (NDMMs) are based on the dideoxysugars ascarylose or paratose, which serve as scaffolds for attachment of moieties from lipid, amino acid, carbohydrate, citrate, and nucleoside metabolism. Mutant screens and comparative metabolomics based on NMR spectroscopy and MS have so far revealed several 100 different ascarylose ("ascarosides") and a few paratose ("paratosides") derivatives, many of which represent potent signalling molecules that can be active at femtomolar levels, regulating development, behaviour, body shape, and many other life history traits. NDMM biosynthesis appears to be carefully regulated as assembly of different modules proceeds with very high specificity. Preliminary biosynthetic studies have confirmed the primary metabolism origin of some NDMM building blocks, whereas the mechanisms that underlie their highly specific assembly are not understood. Considering their functions and biosynthetic origin, NDMMs represent a new class of natural products that cannot easily be classified as "primary" or "secondary". We believe that the identification of new variants of primary metabolism-derived structures that serve important signalling functions in *C. elegans* and other nematodes provides a strong incentive for a comprehensive re-analysis of metabolism in higher animals, including humans.

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1 Introduction: chemical communication in nematodes

The nematodes (roundworms) constitute one of the largest groups of animals, both with respect to the number of different species as well as individuals.¹ Though rarely entering the spotlight, parasitic nematodes are responsible for significant agricultural losses² and are the causative agents of widespread

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human disease in developing countries.^{3,4} In addition, the free-living bacteriovorous nematode *Caenorhabditis elegans* has become one of the most important model organisms for biomedical research. Not least as a result of the early (1998) sequencing of the *C. elegans* genome,⁵ our understanding of nematode biology and genetics has increased dramatically over the past 20 years. It has become apparent that much of *C. elegans* biology is governed by evolutionarily conserved signalling pathways, which enabled exploring ancient features of the regulatory circuitry that controls organismal development, reproduction, longevity, and metabolism.

Surprisingly, nematode biochemistry and metabolism remained largely uncharacterized until recent investigations demonstrated the central importance of small molecule-based signalling cascades for nematode behaviour and development. Interorganismal chemical communication in nematodes was already suspected by Greet in 1964 who reported that males and females of the free living *Panagrolaimus rigidus*, when separated by a cellophane barrier, moved towards members of the opposite sex but not to members of the same sex, suggesting the release of gender-specific attractants, e.g. sex pheromones.⁶ Subsequent research demonstrated the presence of small molecule signals modulating gender-specific attraction in more than 40 representative nematodes including free living, entomopathogenic, plant-parasitic, and zoo-parasitic species.^{7–18} However, despite considerable efforts to isolate and identify nematode pheromones, molecular structures remained elusive for many decades.

Recent research focusing on the free-living *Caenorhabditis elegans*, the necromenic *Pristionchus pacificus*, and a few other species revealed that nematodes are amazingly skilled chemists: using simple building blocks from conserved primary

metabolism and a strategy of modular assembly, they create complex molecular architectures, the nematode-derived modular metabolites (NDMMs) to regulate their development and behaviour. In this review, we will describe the discovery of this modular library, summarize current knowledge regarding the biosynthesis and perception of these components and outline possible implications for small-molecule signalling in higher organisms. Earlier reviews of this field focussed primarily on the biological activity of the initially discovered members of this family of signaling molecules.^{19,20}

2 The discovery of ascaroside-based signalling in the model organism *Caenorhabditis elegans*

The bacteriovorous *C. elegans*, first described by Émile Maupas as *Rhabditis elegans*²¹ was introduced as a model organism for biological research by Sydney Brenner in 1963, given its ease in cultivation, short life cycle, and simple body plan.²² *C. elegans* is an androdioecious nematode species, that is, it is composed of self-fertile hermaphrodites and males. Hermaphrodites constitute more than 99% of worms under laboratory conditions, and the ability to propagate worms from self-fertile hermaphrodites greatly simplifies genetics and mutant creation. Over the past 30 years, *C. elegans* research has contributed to many scientific milestones, including the dissection of programmed cell death,²³ systematic genome cloning,²⁴ first complete genome sequencing of a multicellular organism,⁵ discovery of RNA interference,²⁵ and the use of green fluorescence protein (GFP),²⁶ and *C. elegans* has emerged as an extremely successful model system. However, as is the case for



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many other animal model systems, its metabolome and biochemistry remained largely uncharacterized.

The first hint that small molecule signalling may play an important role in *C. elegans* life history emerged from studies of a peculiar state of developmental diapause called dauer, from the German word for “enduring”. Under unfavourable conditions such as lack of food, high temperature, or high population density, *C. elegans* larvae abort reproductive development and instead enter a specialized, non-feeding and highly stress resistant larval stage that is optimized for dispersal and long-term survival (Fig. 1a). When environmental conditions improve, these dauer larvae resume normal development to reproductive adults.²⁷ Formation of dauer larvae is regulated by conserved signalling pathways, including insulin and TGF- β signalling as well as steroid hormone biosynthesis converging on the nuclear hormone receptor DAF-12, a liver-X and vitamin D receptor homologue.^{28–31} Given that dauer formation results in greatly extended lifespan, the mechanisms that regulate entry and exit from the dauer stage have become an important focus of aging research.²⁸

In 1982 Golden and Riddle reported that dauer formation is modulated by a worm-derived small molecule signal that is constitutively excreted by the worms and “may be a hydroxylated, short-chain fatty acid or a mixture of closely related compounds”.^{32,33} The nematode origin of this pheromone was proven unambiguously by showing that its production depends on a gene (of then unknown identity) named *daf-22*.³⁴ However, the molecular structure of the dauer pheromone remained elusive for more than 20 years until several laboratories reported the identification of a family of dauer inducing glycosides

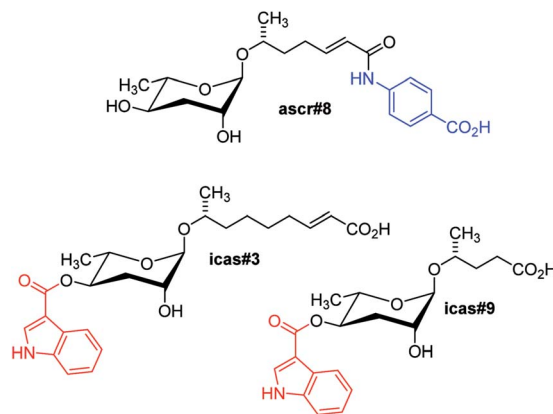


Fig. 2 Modular ascarosides regulating development and mediating behavioural phenotypes in *C. elegans*, including building blocks derived from tryptophan (red) and folate (blue) metabolism.

derived from 1- α -3,6-dideoxymannose, a sugar known as ascarrylose, and a variety of hydroxylated short-chain fatty acids.^{35–38}

Ascarrylose-based glycosides were first described as an “unsaponifiable matter” that accounted for about 25% of the total lipid content of the human intestinal parasitic nematode *Ascaris lumbricoides*.³⁹ At the time of this initial report (1912!), the chemical structure of the *A. lumbricoides*-derived lipids could not be determined, and it was not until 1957 that the exact structures of the sugar and fatty acid parts of these ascarosides were determined (Fig. 1c).⁴⁰ The *A. lumbricoides*-derived ascarosides feature very long aliphatic side chains (predominantly 29 and more carbons in length), are produced in copious amounts and are presumed to form part of a protective coating of *Ascaris* eggs.

In contrast, the dauer-inducing ascarosides identified from *C. elegans* incorporate short fatty acid-derived side chains of only 3–9 carbons (Fig. 1b) and are produced at low (nano- or micromolar) concentrations. Activity-guided fractionation of worm culture media with dauer-inducing activity resulted in the isolation of *ascr#1* (daumone#1, C7),^{35†} the methyl ketone *ascr#2* (daumone#2, C6), *ascr#3* (daumone#3, C9),³⁶ and *ascr#5* (C3),³⁷ which were shown to induce dauer formation at near-physiological concentrations. However, even mixtures of synthetic samples of these compounds failed to fully reproduce the activity of unfractionated worm media, suggesting that there must be additional dauer-inducing components that had been missed by activity-guided fractionation. As described in the following section, the use of comparative metabolomics enabled the identification of important additional dauer pheromone components, including the so far most-potent dauer-inducing ascaroside, *ascr#8* (Fig. 2).³⁸ *Ascr#8* represents the first example for an ascaroside that incorporates additional moieties beyond the ascarrylose sugar and the fatty acid-derived side chain, and thus provided a first glimpse at the vast diversity of multi-modular ascarosides – the NDMMs – later identified from *C. elegans*, *P. pacificus*, and other nematodes.

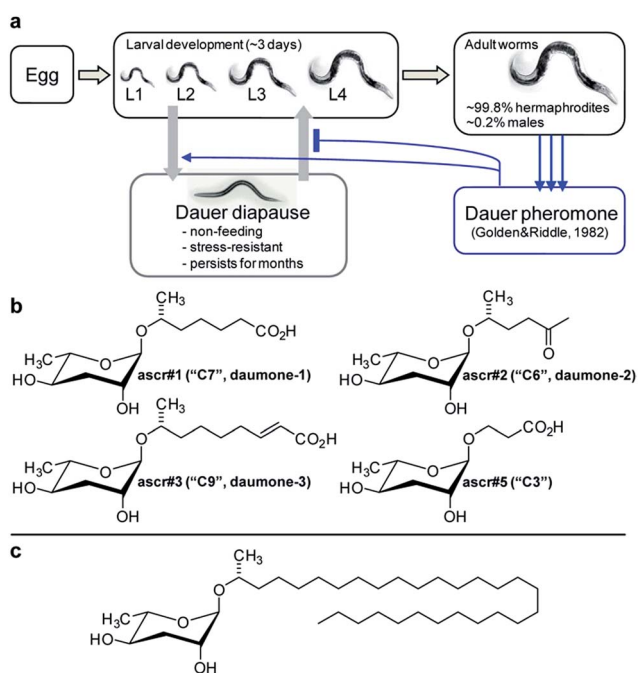


Fig. 1 Initial identification of ascarosides in nematodes. (a) *C. elegans* life cycle. (b) Ascarosides with potent dauer inducing activity. (c) Lipid-like ascaroside from *Ascaris* spp.

† For ascaroside nomenclature see www.smid-db.org



Soon after identification of the first dauer-inducing ascarosides, it became clear that ascarosides regulate many other aspects of nematode life history, including social behaviours and adult lifespan. For example, mixtures of *ascr#2*, its 2-glucoside *ascr#4*, *ascr#3*, and *ascr#8* (Fig. 1b & 2) were found to act as potent male-specific attractants.^{38,41} These hermaphrodite-produced ascarosides strongly synergize in male attraction, that is, mixtures of these compounds were found to be much more attractive than when tested individually, even at elevated concentrations. Behavioural responses to these ascarosides were found to be highly sex-specific: whereas males are attracted by *ascr#2* and *ascr#3*, these compounds act as deterrents for hermaphrodites. Generally, ascaroside concentrations required for behavioural responses (pico- to nanomolar) are much lower than those required for dauer induction.³⁸

In addition, *ascr#2* and *ascr#3* (Fig. 1b) have been shown to regulate adult lifespan in a manner that is distinct from dauer induction. When exposed to nanomolar concentrations of these ascarosides after reaching adulthood, hermaphrodite worms live up to 25% longer than untreated controls.⁴² Other, yet unidentified ascarosides that are produced by male worms appear to shorten hermaphrodite lifespan.⁴³

2.1 The role of comparative metabolomics for NDMM discovery

As described in the preceding paragraphs, the initial discovery of ascarosides as components of the *C. elegans* dauer and mating pheromone relied on activity-guided fractionation of worm culture media extracts.^{35–37,41} The ascarosides' synergistic activities, however, suggested that alternative methods for their detection may be required because compounds that act synergistically are likely to be missed by fractionation-based approaches since their activity will be greatly attenuated or abolished during purification. In fact, it was found that the initially identified ascarosides (*ascr#1–5*, Fig. 1b) did not fully reconstitute the activity of the natural pheromone blend. Therefore, identification of additional pheromone components was pursued using comparative metabolomics-based strategies that do not primarily rely on bioassays of fractionated material. Essential for the use of comparative metabolomics for pheromone identification was the earlier characterization of viable *C. elegans* mutants, e.g. *daf-22*, that do not produce dauer or sex pheromone.^{18,34} Comparative analysis of spectroscopic data derived from pheromone-containing wild-type *exo*-metabolome samples with data sets obtained from pheromone-deficient *daf-22* *exo*-metabolome samples could thus reveal spectroscopic features that are *daf-22* dependent and represent the sought-after additional pheromone components.

For the purpose of comparing wild-type and *daf-22* metabolome samples, Differential analysis of 2D NMR spectra (DANS), a method that relies on comparison of high-resolution dqfCOSY spectra, proved particularly useful.³⁸ DANS revealed a series of *daf-22*-dependent NMR signals that enabled the identification of several additional ascarosides, including *ascr#8* (Fig. 2) which incorporates a *p*-aminobenzoic acid (PABA) moiety likely

derived from folate metabolism.³⁸ *Ascr#8* was then shown to be highly active in promoting male attraction in synergism with *ascr#2* and *ascr#3* (Fig. 1b), in addition to strong dauer-inducing activity. Synthetic blends of *ascr#2*, *ascr#3*, and *ascr#8* at near-physiological concentrations were found to fully reconstitute the behavioural activity of unfractionated *exo*-metabolome, demonstrating the utility of DANS in connecting biological activity and molecular structures.³⁸

In addition, DANS revealed a group of *daf-22* dependent indole carboxy ascarosides (e.g. *icas#3* and *icas#9*), which carry an indole-3-carboxylate moiety at the 4-position of the ascaroylose and were subsequently shown to act as highly potent hermaphrodite aggregation pheromones (Fig. 2).⁴⁴ The first indole ascaroside, *icas#9*, had previously been identified *via* activity-guided fractionation as a minor component of the dauer pheromone;⁴⁵ however, extensive bioassays revealed that *icas#9* and *icas#3* act as potent attractants and aggregation pheromones at concentrations significantly lower than those required for dauer formation.⁴⁴ Prior to the identification of the indole ascarosides, the existence of an aggregation pheromone in *C. elegans* had not been suspected, demonstrating how comparative metabolomic analyses and subsequent identification of novel biogenic small molecules can lead to the discovery of new and unexpected phenotypes.

2.2 Targeted metabolomics reveals structural and functional diversity

The finding that important behavioural phenotypes in *C. elegans* were elicited by picomolar concentrations of indole ascarosides, demonstrated the requirement to develop more sensitive methods for ascaroside screening. Considering the inherently low sensitivity of NMR spectroscopy in comparison to mass spectrometry (MS) a highly sensitive MS/MS technique for selective detection of known and yet unidentified ascarosides in highly complex metabolome extracts was developed, which depends on the production of a characteristic C₃H₅O₂ fragment ion at *m/z* 73 upon collision induced dissociation (CID) of negatively charged ascaroside-based ions.⁴⁶ Comparative MS/MS-based precursor ion screening of *C. elegans* wild type and mutant metabolomes enabled a functional characterization of several genes implicated in ascaroside biosynthesis (see following sections) and revealed a larger family of NDMMs incorporating building blocks derived from additional primary metabolic pathways, in addition to previously overlooked homologues and isomers.⁴⁶ For example, targeted MS/MS analyses of the *C. elegans* metabolome revealed 4-(2-methylbutenoyl)- and 4-hydroxybenzoyl-substituted ascarosides, *mbas#3* and *hbas#3*, respectively (Fig. 3). Whereas the 4-hydroxybenzoyl moiety is likely derived from tyrosine catabolism, the origin of the 2-methylbutenoic (tiglic) acid moiety in *mbas#3* is less clear, as it could be derived from several different known pathways, for example isoleucine catabolism or mitochondrial short-chain fatty acid metabolism. *hbas#3*, which is produced at much lower concentrations (100–1000 fold less) than simple ascarosides such as *ascr#3*, was shown to be an extremely potent hermaphrodite attractant that is active even at



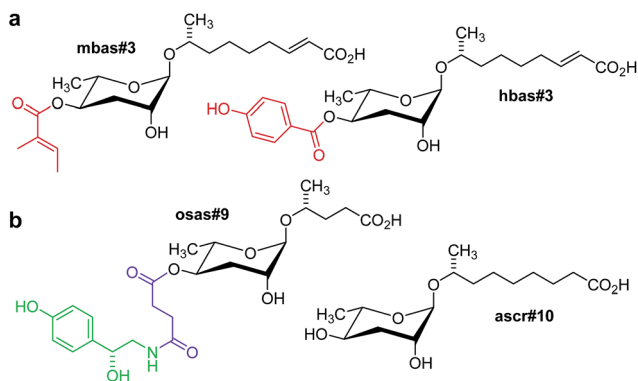


Fig. 3 (a) *mbas#3* and *hbas#3*, two modular ascarosides including tigloyl and *p*-hydroxybenzoyl modules putatively derived from amino acid or short-chain fatty acid metabolism (red) that were identified *via* targeted metabolomics in mixed stage *C. elegans* cultures.⁴⁶ (b) *ascr#10*, a male-produced hermaphrodite attractant, and *osas#9*, a compound produced abundantly by starved L1 larvae,⁴⁷ incorporating succinyl (magenta) and octopamine (green) moieties.⁴⁹

low femtomolar concentrations.⁴⁶ The biological functions of *mbas#3* have not yet been investigated in detail.

Furthermore, targeted ascaroside profiling using precursor ion screening enabled the identification of life-stage and sex-specific ascarosides. It should be noted that most of the original work on small-molecule signalling in *C. elegans* was based on the analysis and biological testing of extracts derived from large liquid cultures containing worms of all ages and life stages, including eggs and larvae. However, given that ascarosides serve a great variety of biological functions, it seemed likely that different life stages would produce different ascaroside blends. Comparative MS/MS analysis of metabolome extracts of samples of a few hundred hand-picked *C. elegans* males and hermaphrodites facilitated the identification of *ascr#10* (Fig. 3) as a highly potent, specifically male-produced hermaphrodite attractant.⁴⁷ In addition, HPLC-MS analysis of different larval stages showed that production of one of the main dauer-inducing ascarosides, *ascr#2*, greatly increases before entering the dauer stage, whereas dauer larvae themselves produce little or no ascarosides.⁴⁸ MS/MS analysis of different developmental stages also resulted in the identification of new types of modular ascaroside derivatives. For example, *osas#9* and *osas#10*, two ascarosides incorporating octopamine-*N*-succinyl moieties, were found to be produced specifically by L1 larvae and function as a dispersal signal in the absence of food (Fig. 3).⁴⁹ In nematodes, octopamine serves as a neurotransmitter in a manner similar to serotonin in higher animals, and thus *osas#9* and *osas#10* appear to connect neurotransmitter signalling with ascaroside-based interorganismal communication.

2.3 Modular assembly and structure–activity relationships

Extensive bioactivity screens using synthetic samples of the identified nematode signalling molecules have shown that even minor structural differences can result in starkly different activity profiles.^{50,51} For example, *ascr#10*, featuring a saturated 9-carbon side chain, strongly attracts hermaphrodites (Fig. 3),

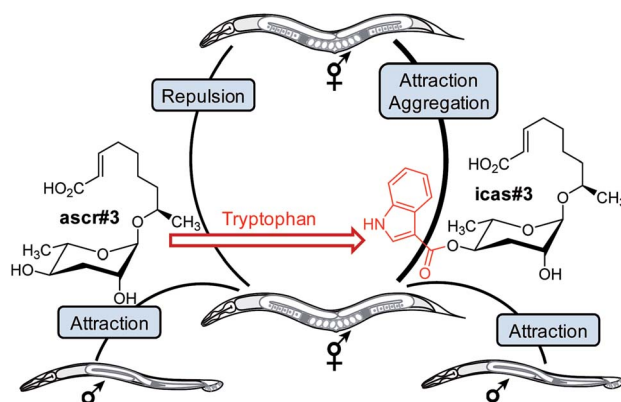


Fig. 4 Attachment of a tryptophan-derived indole carboxy moiety to the deterrent *ascr#3* results in the hermaphrodite attractant *icas#3*. In addition, *ascr#3* is part of the male-attracting pheromone (see text).⁴⁴

whereas *ascr#3*, which differs from *ascr#10* only in that its side chain includes a double bond, acts as a deterrent (Fig. 1b).⁴⁷ Similarly, modifications at the 4-position of the ascarylose scaffold often have a dramatic effect on biological activity. Whereas *ascr#3* acts as a hermaphrodite repellent in higher concentrations, attachment of indole carboxyl or 4-hydroxybenzoyl moieties to its 4-position affords the potent aggregation pheromones *icas#3* and *hbas#3* (Fig. 3 and 4).^{44,46} The indole ascaroside *icas#9*, featuring a saturated 5-carbon side chain, additionally exhibits dauer inducing activity,⁴⁵ whereas *ascr#9*, lacking the indole carboxy moiety, is not active in this assay.⁵¹ Extensive HPLC-MS analyses of worm *exo*-metabolome and worm body extract samples have demonstrated that attachment of the tryptophan-derived indole carboxy moiety is tightly controlled. For example, the potent attraction pheromones *icas#3* and *icas#9* are present in almost identical amounts (Fig. 2), whereas the unmodified *ascr#3* is orders of magnitudes more abundant than the shorter chained *ascr#9*.⁴⁶

Attachment of additional building blocks at the carboxy terminus of the side chains can also modify biological function. As described above, *ascr#8*, which includes a 4-aminobenzoic acid moiety that most likely originates from bacterial folate metabolism³⁸ is a potent male attractant (Fig. 2), whereas the corresponding unmodified ascaroside, *ascr#7* (not shown), is inactive in this assay.³⁸ Modification of the side chain carboxylic acid terminus may also be involved in the regulation of ascaroside secretion and storage. For example, a variety of β -glucosyl ester ascarosides (*e.g.* *glas#3*, Fig. 11) have been identified in *C. elegans* worm body extracts but could not be detected in *exo*-metabolomes samples.⁴⁶ Overall, the observation that even seemingly minor structural changes in NDMMs can greatly affect biological activity strongly suggests that their biosynthesis must be precisely regulated.

3 Ascaroside signalling is highly conserved in nematodes

The production of ascarosides as well as their use as signalling molecules is highly conserved among nematodes, based on the



recent analysis of a number of species representative of several groups of free living, entomopathogenic, and parasitic nematodes.⁵² Ascarosides featuring short to medium length side chains (C₃–C₁₁) are abundantly produced by the free-living *C. elegans*,⁴⁶ *Panagrellus redivivus*,⁵³ and many other nematode species,⁵² whereas ascarosides incorporating longer side chains (C₁₁–C₂₀) have been identified in the entomopathogenic *Heterorhabditis bacteriophora*,^{52,54} the animal parasite *Pelodera strongyloides*,⁵² and several plant-parasitic nematode species.⁵⁵ Ascarosides with very long-chain fatty acid residues (C₂₁–C₃₃) have been identified in the metabolomes of *C. elegans* peroxisomal β -oxidation mutants⁵⁶ as well as in mammalian parasites of the genus *Ascaris*.^{40,57} Considering that many nematode species share identical habitats, overlap in the production of putative ascaroside-based signalling molecules raises interesting questions regarding the maintenance of species-specific communication. Preliminary comparative analysis of biological responses to ascarosides demonstrated that ascaroside functions are highly species-specific.⁵² In addition, more detailed analysis suggests that “simple ascarosides”, consisting of the sugar unit and the lipid side chain only, serve as templates for species-specific modifications and attachment of additional building blocks that are required for specific biological activities. For example, in the entomopathogenic nematode *Heterorhabditis bacteriophora*, the ethanolamide ascaroside easc#18 has been shown to serve as a potent dauer inducing signal, whereas the parent ascaroside, ascr#18 is much less active (Fig. 5).⁵⁴ In another example, sex-specific side chain hydroxylation in the sour paste nematode *Panagrellus redivivus* results in formation of the male-specific dihydroxy ascaroside dhas#18, which acts as a female attractant (Fig. 5), whereas male attraction is modulated by the widely distributed ascr#1 (Fig. 1b).⁵³

Given that the use of ascarosides as signalling molecules is highly conserved among nematodes, it seems possible that other organisms have evolved the capability to detect and respond to these nematode-specific molecular markers. Recent work showed that, in fact, ascarosides mediate cross-kingdom ecological interactions: in fungi that feed on nematodes, sensing of nematode-produced ascarosides was found to induce formation of specific mycelial structures that serve as “nematode traps”.⁵⁸

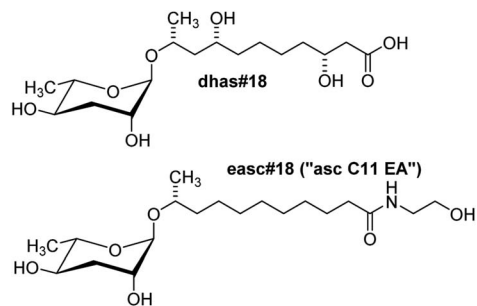


Fig. 5 Structures of the side chain-hydroxylated dhas#18, a pheromone identified from the sour paste nematode *P. redivivus*,⁵⁴ and easc#18, a component of the dauer pheromone of the entomopathogenic nematode *H. bacteriophora*.⁵⁴

3.1 Complex NDMM architectures from *Pristionchus pacificus*

The nematode *Pristionchus pacificus* occurs in association with scarab beetles and has been developed, alongside *C. elegans*, as a satellite model organism for the study of developmental and evolutionary biology.⁵⁹ *P. pacificus* was selected as a satellite model system to *C. elegans* because it is related, but also sufficiently distinct evolutionarily, to enable the comparative study of homologous biosynthetic, cellular, and developmental processes. In contrast to the free-living *C. elegans*, *P. pacificus* forms a close association with scarab beetles, in which dauer larvae persist on live beetles, without negative effect. Following death of the beetle, the larvae resume development and reproduce, feeding on bacteria that consume the beetle carcass. This “necromenic” association with beetles may represent a pre-adaptation to the evolution of parasitism.

Previous work had suggested that, like in *C. elegans*, dauer formation in *P. pacificus* depends on small molecules. Furthermore, it had been shown that in *P. pacificus* a unique dimorphism of mouth development is likely controlled by inter-organismal small-molecule signalling. Under conditions of high population density, worms develop a wider mouth (“eury-stomatous”) equipped with a tooth-like structure that enables a predatory lifestyle consuming other nematodes, whereas under low population density conditions worms develop a narrow mouth (“stenostomatous”) optimized for a bacterial diet.^{60,61} Since none of the ascarosides previously identified from *C. elegans* elicited dauer formation or mouth dimorphism in *P. pacificus*, it appeared that other, species-specific components must be responsible. Extensive analysis of active *P. pacificus* *exo*-metabolome fractions *via* NMR spectroscopy and MS/MS revealed a fascinatingly diverse array of multimodular NDMMs, incorporating building blocks from all major primary metabolic pathways including carbohydrate, lipid, amino acid, nucleoside, and neurotransmitter metabolism (Fig. 6).

Many of the building blocks in the *P. pacificus* NDMMs featured unexpected structural modifications. For example, whereas all known *C. elegans* NDMMs include L-ascarylose as the central scaffold, the *P. pacificus* metabolites are based on two different di-deoxysugars, L-ascarylose and the related L-paratose, a new sugar, the D-enantiomer of which having previously been reported from bacteria.⁶³ Additional structural and functional diversity of the *P. pacificus* NDMMs derives from dimerization of ascarosides. For example, dasc#1, formally derived from dimerization of ascr#1, strongly induces the eury-stomatous mouth form, whereas the monomeric ascr#1 is inactive (Fig. 7).^{62,64} Another dimeric ascaroside, ubas#1, which is formally derived from dimerization of two different ascarosides, the (ω -1)-side chain-functionalized ascr#9 and (ω)-functionalized oscr#9, contributes to dauer formation in some *P. pacificus* strains (Fig. 6). The structures of ubas#1 further includes an unusual ureido isobutyrate moiety that likely originates from metabolism of the nucleobase thymine. Nucleoside metabolism further contributes to dauer signalling in *P. pacificus* *via* a family of paratose-based NDMMs, npar#1–3, which incorporate a L-threonylcarbamoyl adenosine moiety that most



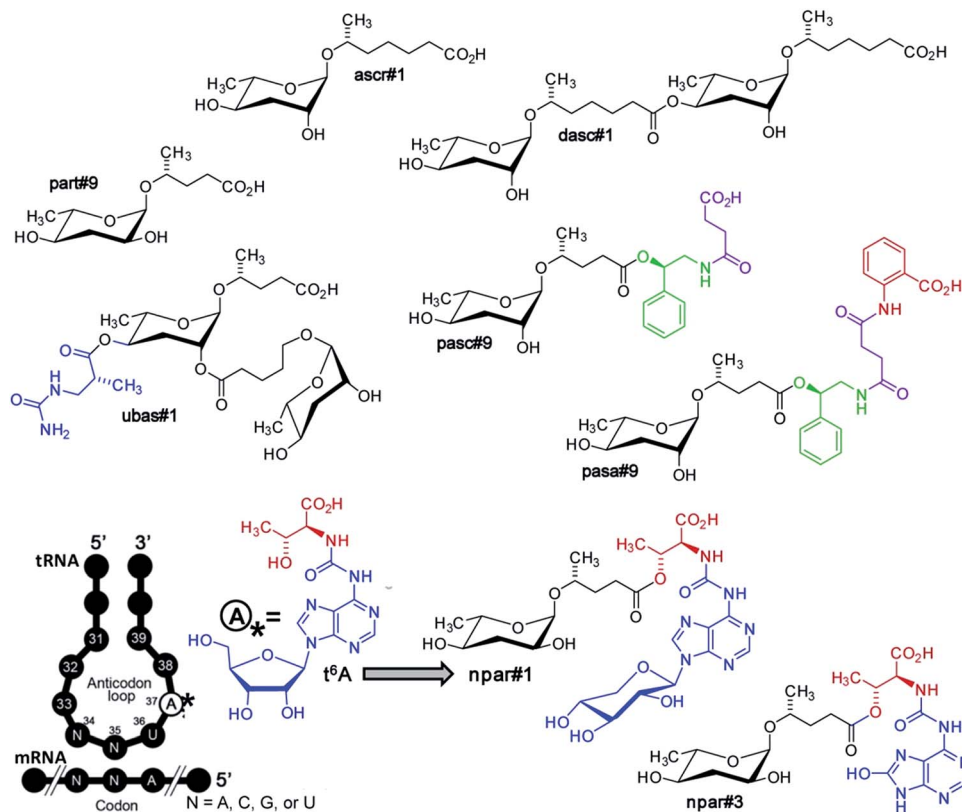


Fig. 6 Ascaroside and paratoside-derived metabolites in *P. pacificus*. Major components of the *P. pacificus* *exo*-metabolome derived from assembly of building blocks from carbohydrate, lipid, amino acid (red), and nucleoside (blue) metabolism, as well as TCA cycle-derived succinate (magenta). Also shown is the highly conserved tRNA nucleoside, N^6 -threonylcarbamoyladenine (t^6A), a putative precursor of paratosides npar#1–3.⁶²

likely originates from canonical threonylcarbamoyl adenosine (t^6A), a highly conserved nucleoside found adjacent to the anticodon triplet of a subset of tRNAs (Fig. 6).⁶² Most surprisingly, however, a xylopyranoside residue was observed in npar#1 instead of the canonical ribofuranoside variant of t^6A found in tRNA.⁶² Physiological concentrations of npar#1 strongly induced dauer formation in *P. pacificus*, whereas the corresponding unmodified paratoside part#9 was much less active.⁶² Two additional derivatives have been described. npar#2 represents the *N*-deglycosylated analogue of npar#1, whereas in npar#3 the adenosine moiety of *N*-deglycosylated npar#1 is replaced by the corresponding 8-hydroxy derivative.⁶⁵ Hydroxylated adenosine derivatives have been shown to be produced as a result of oxidative damage to nucleic acids,⁶⁶ suggesting that npar#3 functions as an indicator of oxidative stress.

P. pacificus also produces a family of NDMMs that, similar to osas#9 in *C. elegans*, incorporate putatively neurotransmitter-derived moieties, e.g. pasc#9 (Fig. 6). Analogous to the incorporation of *N*-succinyl-octopamine in osas#9 (Fig. 3), pasc#9, the most abundant NDMM in all *P. pacificus* strains analyzed so far, incorporates a *N*-succinyl-phenylethanolamine moiety, suggesting that *N*-succinylation plays a role in nematode neurotransmitter metabolism.^{49,62} In contrast to osas#9 in *C. elegans* (Fig. 3), the succinamide in pasc#9 is linked to the carboxy terminus of the side chain instead of the ascarylose.

Recently, a derivative of pasc#9 has been described in which an additional anthranilic acid likely derived from tryptophan is attached to the succinate unit. This compound, named pasa#9, represents the first NDMM composed of 5 different modules

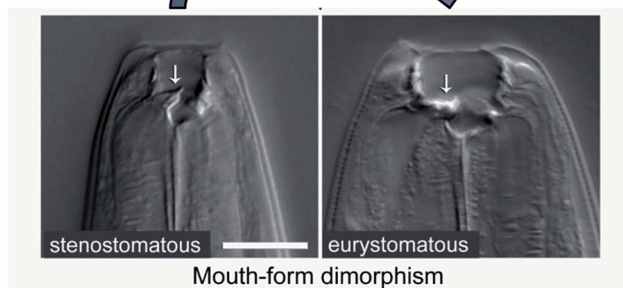
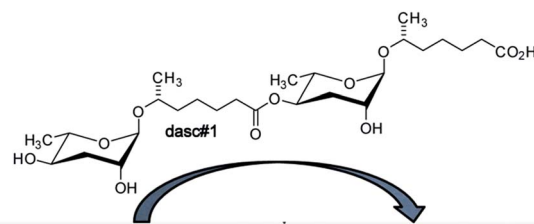


Fig. 7 The dimeric ascaroside dasc#1 promotes development of the wide (eurystomatous) mouth form that enables a predatory lifestyle consuming other nematodes.



(Fig. 6).⁶⁴ The structure of *pasa#9* demonstrates that additional modules must not necessarily be incorporated *via* direct attachment to the ascaroside scaffold, but may also get linked to other modules, suggesting that NDMMs may be even more structurally diverse than previously suspected. Penta-modular *pasa#9* and *pasc#9* are accompanied by smaller amounts of the cyclic succinimide *pasy#9* (not shown), which may represent a by-product of *pasc#9* biosynthesis or could be derived from decomposition of *pasa#9*.^{62,64}

Many of the new chemical structures proposed on the basis of spectroscopic analysis of the *P. pacificus* metabolome were structurally much more complex than the NDMMs previously identified from *C. elegans* and other nematodes (Fig. 6). As a result, confirmation of the proposed structures *via* total synthesis emerged as a major bottle neck in the characterization of *P. pacificus* NDMMs. For example, the synthesis of *npar#1* was based on a sequence with 13 linear steps. Therefore, proposed structures were prioritized for synthesis based on their relative abundances and results from activity-guided fractionation; however, structures of a large number of additional NDMMs remain to be confirmed *via* synthesis which will then also facilitate their biological evaluation. As in the case of *C. elegans*, the identification of additional NDMMs in *P. pacificus* may lead to the discovery of new small-molecule mediated phenotypes.

3.2 Structure–activity relationships and natural variation in *P. pacificus*

NDMM biosynthesis in *P. pacificus* appears to be tightly regulated. *ascr#1*, including a seven-carbon side chain, forms the 4'-homodimer *dasc#1*, whereas an ascaroside featuring an (ω -1)-functionalized five-carbon side chain forms the scaffold for a heterodimer with an ascaroside featuring an (ω)-functionalized five-carbon side chain in *ubas#1* (Fig. 6).⁶² Notably, homodimers of ascarosides with (ω)- or (ω -1)-functionalized five-carbon side chains are absent. Similarly, there are no ascaroside-based analogues of the paratose-based nucleobase-derived *npar#1–3* (Fig. 6), and no paratose-based analogues of the ascarylose-based *N*-succinyl-phenylethanolamine-containing *pasc#9*, *pasy#9*, and *pasa#9* have been found.⁶²

Correspondingly, the biological activity of the identified NDMMs is strongly structure-dependent.⁶² A screen of six different natural *P. pacificus* strains has further shown that bioactivity of different NDMMs varies greatly between different natural isolates.⁶⁴ These observations indicate that NDMM receptors evolve rapidly, as also suggested by the finding that laboratory strains of *C. elegans* are less sensitive to dauer pheromone than the parent wildtype strain due to rapid accumulation of mutations in receptor-coding genes.⁶⁷ Comparison of the metabolomes of the six different natural *P. pacificus* strains further revealed marked differences in the relative abundances of NDMM in different strains.⁶⁴ For example, the ascaroside *pasc#9* varied more than six-fold between strains. Several NDMMs, for example *ubas#1* and *ubas#2*, were completely absent in two of the six analysed strains. Furthermore, it was found that some *P. pacificus* strains respond

strongly to NDMMs they do not produce themselves, which suggested that NDMMs may play a role in communication and/or competition between different strains.^{64,68} Similarly, analysis of several *C. elegans* wild type isolates indicated that the production⁵² and dauer inducing activity⁶⁹ of *ascr#2* and *ascr#3* is strongly strain-specific, suggesting the possibility of “dishonest” signalling between competing *C. elegans* strains.⁶⁹

4 Ascaroside biosynthesis and primary metabolism

The NDMMs identified from *C. elegans*, *P. pacificus*, and other nematode species incorporate building blocks originating from diverse primary metabolic pathways and hence may encode information about the metabolic state of the producing organism. In addition to carbohydrate and fatty acid metabolism, which together give rise to the ascaroside core structures, integration of amino acid, neurotransmitter, and nucleoside metabolism as well as the citric acid cycle affords a diversity of complex architectures. Their *de novo* biosynthesis appears to proceed without participation of microbial metabolism, since all NDMMs identified from *C. elegans* and *P. pacificus* are also produced in axenic (bacteria-free) cultures,^{46,62} as long as all necessary building blocks – including those the nematodes do not produce themselves, such as bacteria-derived PABA – are provided in the media. Moreover, the highly specific assembly of primary metabolic building blocks, for example the combination of octopamine-derived moieties only with ascarosides of specific side chain lengths, suggests that NDMM biosynthesis is carefully controlled.

1-Ascarylose (or 1-paratose) are glycosidically connected to fatty acid-derived side chains of varying length and functionality that have been shown to originate from peroxisomal β -oxidation,^{45,46,56} producing homologous series of the glycolipid core structures. In some nematode species, the glycolipid profiles are dominated by one or two chain lengths, whereas other species have broader distributions.⁵² Although the origin of these fatty acid chains is not well understood, the general predominance of odd-numbered side chains indicates that these are derived from long-chain odd-numbered fatty acids *via* multiple steps of β -oxidation.

In most ascarosides/paratosides, the fatty acid residue is attached to the dideoxysugar at the (ω -1)-position and the corresponding chiral centre always carries the (*R*)-configuration. Less common are (ω)-linked side chains, as in *ascr#5*, a major component of the *C. elegans* dauer pheromone, which carries an (ω)-linked 3-hydroxypropanoic acid residue (Fig. 1b),³⁷ and several other compounds based on longer chained (ω)-linked side chains identified from both *C. elegans* and *P. pacificus*.^{46,62} As described in the following section, targeted metabolomic analysis of peroxisomal β -oxidation mutants revealed complete series of longer chain homologues with (ω -1)- and (ω)-hydroxylated side chains ranging from 3 to 21 carbons which suggested that (ω -1)- and (ω)-hydroxylation occurs upstream of peroxisomal β -oxidation.⁴⁶



4.1 The role of peroxisomal β -oxidation

The participation of primary metabolic enzymes has been demonstrated only for a few of the building blocks in NDMMS: the origin of the octopamine moiety in *osas#9* and *osas#10* from canonical tyrosine metabolism (Fig. 10)⁴⁹ and, importantly, side chain biosynthesis *via* peroxisomal β -oxidation (Fig. 8). Shortly after describing the dauer pheromone, Golden and Riddle in 1985 reported a gene named *daf-22* that was required for dauer pheromone production.³⁴ Following the identification of ascarosides as major dauer pheromone components, it was observed that *daf-22* mutant worms do not produce short-chain ascarosides,⁴¹ and subsequent work revealed that *daf-22* encodes a homologue of peroxisomal 3-ketoacyl-CoA thiolases that is expressed in the hypodermis and intestine.⁴⁵ Mammalian homologues of *daf-22* have been shown to catalyze the last step of peroxisomal fatty acid β -oxidation, a four-step iterative process that is evolutionarily highly conserved (Fig. 8).

Subsequent studies showed that *C. elegans* genes coding for homologues of enzymes catalyzing the remaining three enzymatic steps in peroxisomal β -oxidation also participate in ascaroside biosynthesis. This includes three acyl-CoA oxidases (*acox-1*,^{46,70} *acox-2* and *acox-3*⁷¹), the enoyl-CoA hydratase *maoc-1*,⁴⁶ and the 3-hydroxyacyl-CoA dehydrogenase *dhs-28*.^{45,46} The role of *acox-1*, *maoc-1*, *dhs-28*, and *daf-22* in chain shortening of ascarosides *via* peroxisomal β -oxidation was investigated using

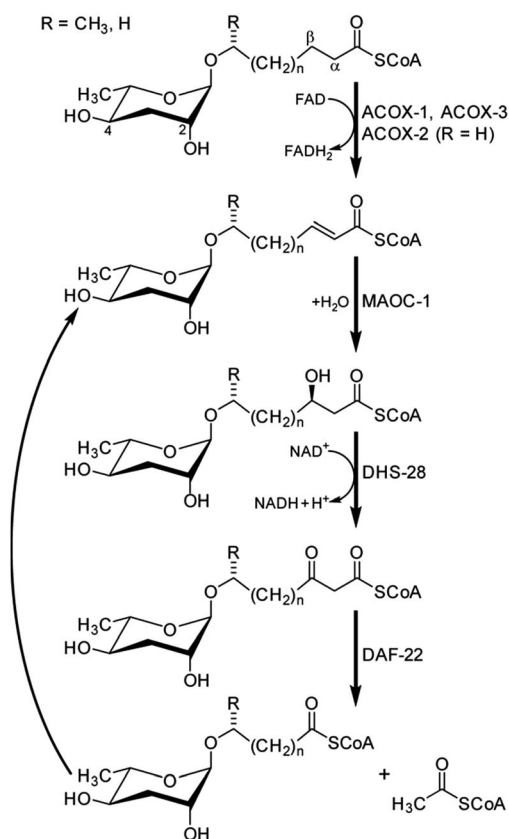


Fig. 8 Side chain shortening of long-chain ascarosides *via* iterative four-step peroxisomal β -oxidation in *C. elegans*.^{46,71}

the ascaroside-targeted MS/MS screen described above.⁴⁶ Mutation of *acox-1*, *maoc-1*, or *dhs-28* was shown to result in accumulation of shunt metabolites with structural features supportive of the enzymatic functions of these genes, which originally had been predicted solely based on homology. For example, production of short-chain ascarosides (<9 carbon side chain) and ascarosides with α,β -unsaturated side chains, which are highly abundant in *C. elegans* wild type, is attenuated in *acox-1* mutant metabolomes. Instead, *acox-1* mutant worms accumulate large amounts of saturated longer-chain ascarosides (especially C_9 , C_{11} , and C_{13}), indicating that *acox-1* serves as acyl-CoA oxidase introducing a double bond in the first step of peroxisomal β -oxidation.⁴⁶ Additional work revealed that homodimers of the acyl-CoA oxidase *acox-1* specifically act on the CoA ester of ascarosides with nine carbon side chains, whereas heterodimers of *acox-3* and *acox-1* act on ascaroside CoA esters of shorter carbon chains. Furthermore, homodimers of *acox-2* appear to act specifically on ascaroside CoA esters of (ω)-oxygenated side chains and thus may control the biosynthesis of the dauer pheromone component *ascr#5*.⁷¹

In contrast to *acox-1*, *maoc-1*, and *dhs-28*, targeted metabolomic analyses *via* HPLC-MS/MS did not reveal any shunt metabolites whose structures could serve to corroborate the predicted function of *daf-22* as a 3-ketoacyl-CoA thiolase.⁴⁶ This was not unexpected, given that β -ketoacids such as they would be expected to form from accumulating substrates of DAF-22 (Fig. 8) easily decarboxylate or may undergo other chemical transformations. To identify such *daf-22*-dependent shunt metabolites, the *daf-22* metabolome was re-analyzed using 2D NMR-based comparative metabolomics with a focus on spectroscopic signatures present in the mutant metabolome, but absent in wildtype. For this purpose, multivariate differential analysis by 2D NMR spectroscopy (mvaDANS) was developed, which integrates automatic crosspeak identification and binning with statistical analysis *via* principal component analysis. mvaDANS then revealed a series of very long-chain ascaroside ethanolamides and other long-chain ascarosides as shunt metabolites in *daf-22* mutants (Fig. 9).⁵⁶ Subsequent re-analysis of the metabolomes of other peroxisomal β -oxidation mutants revealed similar shunt metabolites. Formation of these long-chain ethanolamides in response to defects in *daf-22* and other components of peroxisomal β -oxidation was associated with a severe depletion of endocannabinoid pools, demonstrating an unexpected interaction between peroxisomal lipid β -oxidation and the biosynthesis of endocannabinoids (Fig. 9), which are major regulators of lifespan in *C. elegans*.⁵⁶

4.2 Biogenesis of multimodular ascarosides

Additional modifications of the ascaroside core structure with other primary metabolism-derived moieties have a dramatic effect on the biological activities and appear to be tightly controlled, since, as discussed above, attachment of additional moieties is highly side chain length-specific and most multimodular ascarosides are produced primarily by specific life stages. Their structures suggest that the indolyl-3-carboxylate, 4-hydroxybenzoate, and tiglitate moieties observed in *C. elegans*



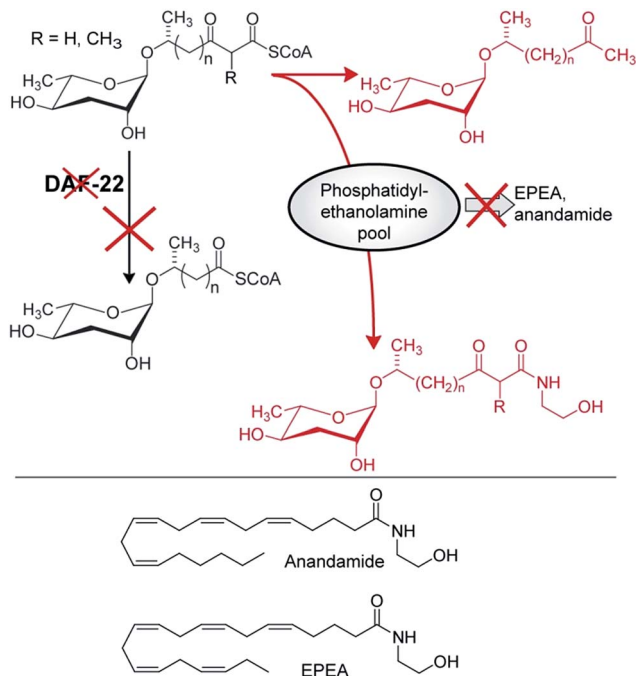


Fig. 9 Interaction of endocannabinoid biosynthesis with peroxisomal β -oxidation. *daf-22* mutation abolishes processing of long-chain acyl-CoA esters, whose conversion into acyl-ethanolamides is associated with reduced EPEA and anandamide production, likely due to depletion of phosphatidylethanolamine pools. Shown in red are shunt metabolites identified in *daf-22* mutant *endo*-metabolomes via *mvaDANS*.⁵⁶

(Fig. 2 & 3) are of amino acid origin, which, however, has only been demonstrated for indole ascarosides, which incorporate isotope labelled tryptophan.⁴⁴ No biosynthetic intermediates have been identified and the enzyme(s) responsible for linking tryptophan metabolism and indole ascaroside biosynthesis have not been identified. Addition of synthetic short-chain ascarosides to *daf-22* mutant cultures (which do not produce short-chain ascarosides) revealed side chain length-specific conversion to the corresponding indole carboxy derivatives, suggesting that attachment of amino acid-derived indol-3-carboxylate moieties occurs downstream of peroxisomal β -oxidation.⁴⁶ Additional evidence for the incorporation of amino acid-derived building blocks originates from the analysis of tyrosine decarboxylase (*tdc-1*) and tyramine β -hydroxylase (*tbh-1*) mutant metabolomes, which revealed that the production of octopamine-*N*-succinamide ascarosides (*osas*) by starved *C. elegans* L1 larvae depends on the production of the neurotransmitter octopamine from *L*-tyrosine via *tdc-1* and *tbh-1* (Fig. 10).⁴⁹

4.3 Ascaroside biosynthesis is strongly regulated

Ascaroside biosynthesis in *C. elegans* strongly depends on environmental factors such as temperature,⁷⁰ population density,⁴⁷ and food availability,^{46,49} as well as developmental stage^{48,49} and gender,⁴⁷ but the underlying mechanisms are poorly understood. Increased production of ascarosides at

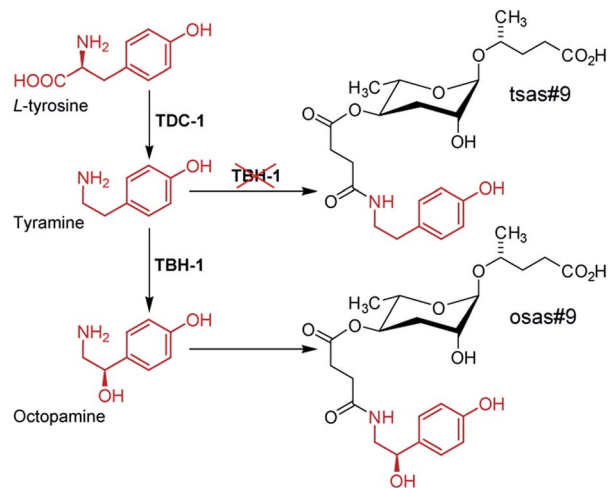


Fig. 10 Model for the biosynthesis of the avoidance signal *osas#9* in *C. elegans*. *tbh-1* mutant worms produce large quantities of the shunt metabolite *tsas#9* due to accumulation of the octopamine precursor tyramine.⁴⁹

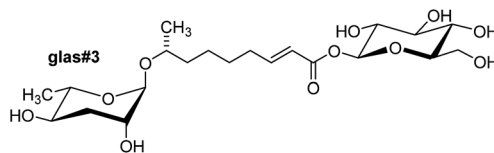


Fig. 11 *glas#3*, an ascaroside glucosyl ester identified in the *C. elegans* *endo*-metabolome.⁴⁶

elevated temperatures corresponds to increased expression of acyl-CoA oxidases *acox-1*,⁷⁰ *acox-2*, and *acox-3*.⁷¹ Furthermore, food availability has been shown to modulate the composition of ascaroside bouquets via differential expression of *acox* genes. Under non dauer inducing conditions (low population density and high food availability) increased expression of *acox-2* results in increased production of (ω)-oxygenated short chained *ascr#5*.⁷¹ Males of *C. elegans* show reduced expression of *acox-1* and consequently release larger amounts of the corresponding substrate, the saturated *ascr#10* that acts as a highly potent hermaphrodite attractant.⁴⁷ Similarly, in the sour paste nematode *Panagrellus redivivus* the female-attracting pheromone *dhas#18* is exclusively produced by males (Fig. 5), whereas the shorter chain male-attracting *ascr#1* is only produced by females (Fig. 1b), suggesting that gender-specific production of sex pheromones originates from gender-specific fine tuning of primary metabolic pathways such as peroxisomal β -oxidation.⁵²

Finally, comparative analysis of *exo*- and *endo*-metabolome (worm body extracts) samples revealed that *C. elegans* and *P. pacificus* exhibit significant control over ascaroside release.^{46,62,64} For example, ascaroside glucosyl esters (e.g. *glas#3*, Fig. 11) have been found only in the *C. elegans* *endo*-metabolome (worm body extract), but not the *exo*-metabolome, suggesting that such glucosides may be involved in ascaroside storage or transport.⁴⁶



5 Ascaroside perception and downstream signalling

The *C. elegans* genome encodes for a large diversity of GTP-binding protein (G protein)-coupled receptors (GPCRs), most of which are expressed in chemosensory neurons.^{72,73} Ascaroside induced developmental phenotypes such as dauer formation depend on at least five different types of chemosensory neurons (ADF, ASG, ASI, ASJ, ASK, and the male-specific CEM neurons),⁷² whereas behavioural phenotypes depend on ASK, ASI, and ADL neurons.^{41,44,74–76} Mutants for G protein alpha subunits *gpa-2* and *gpa-3* are dauer defective; further suggesting that ascaroside perception is mediated by GPCRs.⁷⁷

However, so far only a relatively small number of G-protein coupled receptors involved in ascaroside perception have been identified,^{67,78,79} whereas the diversity of identified ascaroside functions suggests that many more are yet to be discovered. SRBC-64 and SRBC-66 are two heterotrimeric GPCRs expressed in the ASK chemosensory neurons that mediate dauer formation in response to *ascr#1*, *ascr#2*, and *ascr#3* (Fig. 1b).⁷⁹ Two members of the serpentine chemoreceptor class g family, *srg-36* and *srg-37*, discovered through genetic analysis of pheromone resistant *C. elegans* strains obtained from high density cultivation for several years, encode redundant GPCRs for the dauer inducing activity of *ascr#5*, demonstrating that adaptation to specific environments can entail rapid remodelling of the chemoreceptor repertoire. Deletion of a *srg* gene paralogous to *srg-36* and *srg-37* has also been linked to resistance to dauer formation in a laboratory *Caenorhabditis briggsae* strain, indicating that parallel genetic changes can affect life-history traits across species.⁶⁷ Finally, heterodimers of DAF-37 and DAF-38, two GPCRs identified using DAF-8 immunoprecipitation, cooperatively mediate ascaroside perception. *daf-37* mutants are defective in *ascr#2*-mediated dauer-induction but respond normally to other ascarosides, whereas *daf-38* mutants are partially defective to several different ascarosides. Furthermore, cell-specific overexpression revealed that *ascr#2*-mediated dauer formation depends on DAF-37 expression in ASI neurons, whereas DAF-37 expression in ASK neurons regulates behaviour.⁷⁸ Using a photoaffinity-labeled *ascr#2* probe and amplified luminescence assays (AlphaScreen), it was further demonstrated that *ascr#2* binds directly to DAF-37.⁷⁸

Perception of natural dauer pheromone samples, *i.e.* ascaroside-containing *exo*-metabolome, has been shown to act upstream of several conserved signalling pathways, including the insulin, TGF- β , serotonin, and guanylyl cyclase pathways.^{27,31} However, it is not known if and to what extent downstream pathways of individual dauer-inducing ascarosides differ. Nonetheless, it has been shown that different phenotypes induced by the same ascaroside can depend on different downstream signalling components. For example, ascarosides *ascr#2* and *ascr#3* extend lifespan in adult worms *via sir-2.1*, a homologue of the mammalian histone-deacetylase SIRT-1,⁴² and the same two compounds also induce dauer *via* the insulin signalling pathway.³⁶ However, *sir-2.1* is not required for dauer induction by *ascr#2* and *ascr#3*,²⁷ and neither is insulin signalling required for *ascr#2* and *ascr#3*-mediated longevity.⁴²

6 Conclusion

Nematode-derived ascarosides and paratosides represent a “new” class of natural products that cannot easily be classified as “primary” or “secondary” metabolites. Much of their chemical appeal and originality derives from the fact that they incorporate well-known primary metabolites into new structures, sometimes with unexpected chemical modifications. Since the building blocks of NDMMs are directly from conserved primary metabolism, it seems possible that similar types of modular small molecule signals are produced by other metazoans. These possibilities provide strong incentives for a comprehensive re-analysis of vertebrate metabolomes, taking advantage of the approaches developed and validated using nematode model systems.

Further elucidation of the biosynthesis of ascaroside-based NDMMs will reveal how input from conserved primary metabolism is transduced to create signals that regulate development, behaviour, and many other aspects of the life history of nematode model organisms. Moreover, since ascaroside signalling is highly conserved among nematodes, a detailed understanding of their biosynthesis and regulation may enable new approaches for the treatment of human nematode infections or the control of parasitic nematodes in agriculture. Lastly, it should be noted that ascaroside-based NDMMs are likely just one of several families of small molecule signals. Mass spectrometric analyses of the *C. elegans* metabolome have revealed evidence for several 1000 yet unidentified compounds,⁴⁶ and nematode genomes feature many genes that, based on homology, may encode small-molecule biosynthetic pathways, including one gene with clear homology to microbial hybrid PKS/NRPPs (polyketide synthase/non-ribosomal peptide synthetases),⁸⁰ and whose functions remain to be elucidated.

7 Notes and references

- 1 M. Blaxter and G. Koutsovoulos, *Parasitology*, 2014, 1.
- 2 N. A. Mitkowski and G. S. Abawi, *Nematology*, 2003, 5, 77.
- 3 P. J. Hotez, P. J. Brindley, J. M. Bethony, C. H. King, E. J. Pearce and J. Jacobson, *J. Clin. Invest.*, 2008, 118, 1311.
- 4 D. W. Crompton and M. C. Nesheim, *Annu. Rev. Nutr.*, 2002, 22, 35.
- 5 The *C. elegans* Sequencing Consortium, *Science*, 1998, 282, 2012.
- 6 D. N. Greet, *Nature*, 1964, 204, 96.
- 7 L. W. Bone and H. H. Shorey, *J. Chem. Ecol.*, 1978, 4, 595.
- 8 C. D. Green, *Helminthological Abstracts Series B: Plant nematology*, 1980, 49, 81.
- 9 R. N. Huettel, *J. Nematol.*, 1986, 18, 3.
- 10 B. M. MacKinnon, *Parasitol. Today*, 1987, 3, 156.
- 11 H. Jaffe, R. N. Huettel, A. B. Demilo, D. K. Hayes and R. V. Rebois, *J. Chem. Ecol.*, 1989, 15, 2031.
- 12 J. Aumann, E. Dietsche, S. Rutencrantz and H. Ladehoff, *Int. J. Parasitol.*, 1998, 28, 1691.
- 13 J. Aumann and M. Hashem, *Fundam. Appl. Nematol.*, 1993, 16, 43.



- 14 J. Aumann, H. Ladehoff and S. Rutencrantz, *Fundam. Appl. Nematol.*, 1998, **21**, 119.
- 15 J. M. Neves, N. Simoes and M. Mota, *Nematologica*, 1998, **44**, 95.
- 16 M. G. Jonz, E. Riga, A. J. Mercier and J. W. Potter, *Nematology*, 2001, **3**, 55.
- 17 E. E. Lewis, B. Barbarosa and R. Gaugler, *J. Nematol.*, 2002, **34**, 328.
- 18 J. M. Simon and P. W. Sternberg, *Proc. Natl. Acad. Sci. U. S. A.*, 2002, **99**, 1598.
- 19 A. H. Ludewig and F. C. Schroeder, Ascarioside signaling in *C. elegans*, in *WormBook*, ed. The *C. elegans* Research Community, 2013, DOI: 10.1895/wormbook.1.155.1.
- 20 F. C. Schroeder, *ACS Chem. Biol.*, 2006, **1**, 198.
- 21 É. Maupas, *Arch. Zool. Exp. Gen.*, 1900, **8**, 463.
- 22 S. Brenner, *Genetics*, 1974, **77**, 71.
- 23 R. E. Ellis, J. Y. Yuan and H. R. Horvitz, *Annu. Rev. Cell Biol.*, 1991, **7**, 663.
- 24 A. Coulson, J. Sulston, S. Brenner and J. Karn, *Proc. Natl. Acad. Sci. U. S. A.*, 1986, **83**, 7821.
- 25 A. Fire, S. Q. Xu, M. K. Montgomery, S. A. Kostas, S. E. Driver and C. C. Mello, *Nature*, 1998, **391**, 806.
- 26 M. Chalfie, Y. Tu, G. Euskirchen, W. W. Ward and D. C. Prasher, *Science*, 1994, **263**, 802.
- 27 P. J. Hu, Dauer, in *WormBook*, ed. The *C. elegans* Research Community, 2007, DOI: 10.1895/wormbook.1.144.1.
- 28 N. Fielenbach and A. Antebi, *Genes Dev.*, 2008, **22**, 2149.
- 29 D. L. Motola, C. L. Cummins, V. Rottiers, K. K. Sharma, T. Li, Y. Li, K. Suino-Powell, H. E. Xu, R. J. Auchus, A. Antebi and D. J. Mangelsdorf, *Cell*, 2006, **124**, 1209.
- 30 P. Mahanti, N. Bose, A. Bethke, J. C. Judkins, J. Wollam, K. J. Dumas, A. M. Zimmerman, S. L. Campbell, P. J. Hu, A. Antebi and F. C. Schroeder, *Cell Metab.*, 2014, **19**, 73.
- 31 S. S. Lee and F. C. Schroeder, *PLoS Biol.*, 2012, **10**, e1001307.
- 32 J. W. Golden and D. L. Riddle, *Science*, 1982, **218**, 578.
- 33 J. W. Golden and D. L. Riddle, *J. Nematol.*, 1982, **14**, 443.
- 34 J. W. Golden and D. L. Riddle, *Mol. Gen. Genet.*, 1985, **198**, 534.
- 35 P. Y. Jeong, M. Jung, Y. H. Yim, H. Kim, M. Park, E. Hong, W. Lee, Y. H. Kim, K. Kim and Y. K. Paik, *Nature*, 2005, **433**, 541.
- 36 R. A. Butcher, M. Fujita, F. C. Schroeder and J. Clardy, *Nat. Chem. Biol.*, 2007, **3**, 420.
- 37 R. A. Butcher, J. R. Ragains, E. Kim and J. Clardy, *Proc. Natl. Acad. Sci. U. S. A.*, 2008, **105**, 14288.
- 38 C. Pungaliya, J. Srinivasan, B. W. Fox, R. U. Malik, A. H. Ludewig, P. W. Sternberg and F. C. Schroeder, *Proc. Natl. Acad. Sci. U. S. A.*, 2009, **106**, 7708.
- 39 F. Flury, *Arch. Exp. Pathol. Pharmacol.*, 1912, **67**, 275.
- 40 C. Fouquey, J. Polonsky and E. Lederer, *Bull. Soc. Chim. Biol.*, 1957, **39**, 101.
- 41 J. Srinivasan, F. Kaplan, R. Ajredini, C. Zachariah, H. T. Alborn, P. E. A. Teal, R. U. Malik, A. S. Edison, P. W. Sternberg and F. C. Schroeder, *Nature*, 2008, **454**, 1115.
- 42 A. H. Ludewig, Y. Izrayelit, D. Park, R. U. Malik, A. Zimmermann, P. Mahanti, B. W. Fox, A. Bethke, F. Doering, D. L. Riddle and F. C. Schroeder, *Proc. Natl. Acad. Sci. U. S. A.*, 2013, **110**, 5522.
- 43 T. J. Maures, L. N. Booth, B. A. Benayoun, Y. Izrayelit, F. C. Schroeder and A. Brunet, *Science*, 2014, **343**, 541.
- 44 J. Srinivasan, S. H. von Reuss, N. Bose, A. Zaslaver, P. Mahanti, M. C. Ho, O. G. O'Doherty, A. S. Edison, P. W. Sternberg and F. C. Schroeder, *PLoS Biol.*, 2012, **10**, e1001237.
- 45 R. A. Butcher, J. R. Ragains, W. Li, G. Ruvkun, J. Clardy and H. Y. Mak, *Proc. Natl. Acad. Sci. U. S. A.*, 2009, **106**, 1875.
- 46 S. H. von Reuss, N. Bose, J. Srinivasan, J. J. Yim, J. C. Judkins, P. W. Sternberg and F. C. Schroeder, *J. Am. Chem. Soc.*, 2012, **134**, 1817.
- 47 Y. Izrayelit, J. Srinivasan, S. L. Campbell, Y. Jo, S. H. von Reuss, M. C. Genoff, P. W. Sternberg and F. C. Schroeder, *ACS Chem. Biol.*, 2012, **7**, 1321.
- 48 F. Kaplan, J. Srinivasan, P. Mahanti, R. Ajredini, O. Durak, R. Nimalendran, P. W. Sternberg, P. E. Teal, F. C. Schroeder, A. S. Edison and H. T. Alborn, *PLoS One*, 2011, **6**, e17804.
- 49 A. B. Artyukhin, J. J. Yim, J. Srinivasan, Y. Izrayelit, N. Bose, S. H. von Reuss, Y. Jo, J. M. Jordan, L. R. Baugh, M. Cheong, P. W. Sternberg, L. Avery and F. C. Schroeder, *J. Biol. Chem.*, 2013, **288**, 18778.
- 50 F. C. Schroeder, *Chem. Biol.*, 2015, **22**, 7.
- 51 K. A. Hollister, E. S. Conner, X. Zhang, M. Spell, G. M. Bernard, P. Patel, A. C. de Carvalho, R. A. Butcher and J. R. Ragains, *Bioorg. Med. Chem.*, 2013, **21**, 5754.
- 52 A. Choe, S. H. von Reuss, D. Kogan, R. B. Gasser, E. G. Platzer, F. C. Schroeder and P. W. Sternberg, *Curr. Biol.*, 2012, **22**, 772.
- 53 A. Choe, T. Chuman, S. H. von Reuss, A. T. Dossey, J. J. Yim, R. Ajredini, A. A. Kolawa, F. Kaplan, H. T. Alborn, P. E. Teal, F. C. Schroeder, P. W. Sternberg and A. S. Edison, *Proc. Natl. Acad. Sci. U. S. A.*, 2012, **109**, 20949.
- 54 J. H. Noguez, E. S. Conner, Y. Zhou, T. A. Ciche, J. R. Ragains and R. A. Butcher, *ACS Chem. Biol.*, 2012, **7**, 961.
- 55 P. Manosalva, M. Manohar, S. H. von Reuss, S. Chen, A. Koch, F. Kaplan, A. Choe, R. J. Micikas, X. Wang, K.-H. Kogel, P. W. Sternberg, V. M. Williamson, F. C. Schroeder and D. F. Klessig, *Nature Commun.*, 2015, accepted for publication.
- 56 Y. Izrayelit, S. L. Robinette, N. Bose, S. H. von Reuss and F. C. Schroeder, *ACS Chem. Biol.*, 2013, **8**, 314.
- 57 J. P. Bartley, E. A. Bennett and P. A. Darben, *J. Nat. Prod.*, 1996, **59**, 921.
- 58 Y. P. Hsueh, P. Mahanti, F. C. Schroeder and P. W. Sternberg, *Curr. Biol.*, 2013, **23**, 83.
- 59 C. Dieterich, S. W. Clifton, L. N. Schuster, A. Chinwalla, K. Delehaunty, I. Dinkelacker, L. Fulton, R. Fulton, J. Godfrey, P. Minx, M. Mitreva, W. Roeseler, H. Tian, H. Witte, S. P. Yang, R. K. Wilson and R. J. Sommer, *Nat. Genet.*, 2008, **40**, 1193.
- 60 A. Ogawa, G. Bento, G. Bartelmes, C. Dieterich and R. J. Sommer, *Development*, 2011, **138**, 1281.
- 61 J. Sommer Ralf and A. Ogawa, *Curr. Biol.*, 2011, **21**, R758.



- 62 N. Bose, A. Ogawa, S. H. von Reuss, J. J. Yim, E. J. Ragsdale, R. J. Sommer and F. C. Schroeder, *Angew. Chem., Int. Ed.*, 2012, **51**, 12438.
- 63 D. A. L. Davies, A. M. Staub, I. Fromme, O. Luderitz and O. Westphal, *Nature*, 1958, **181**, 822.
- 64 N. Bose, J. M. Meyer, J. J. Yim, M. G. Mayer, G. V. Markov, A. Ogawa, F. C. Schroeder and R. J. Sommer, *Curr. Biol.*, 2014, **24**, 1536.
- 65 J. J. Yim, N. Bose, J. M. Meyer, R. J. Sommer and F. C. Schroeder, *Org. Lett.*, 2015, **17**, 1648.
- 66 S. Kanvah, J. Joseph, G. B. Schuster, R. N. Barnett, C. L. Cleveland and U. Landman, *Acc. Chem. Res.*, 2009, **43**, 280.
- 67 P. T. McGrath, Y. Xu, M. Ailion, J. L. Garrison, R. A. Butcher and C. I. Bargmann, *Nature*, 2011, **477**, 321.
- 68 M. G. Mayer and R. J. Sommer, *Proc. R. Soc. B*, 2011, **278**, 2784.
- 69 S. A. Diaz and M. Viney, *Ecol. Evol.*, 2014, **4**, 2058.
- 70 H. J. Joo, K. Y. Kim, Y. H. Yim, Y. X. Jin, H. Kim, M. Y. Kim and Y. K. Paik, *J. Biol. Chem.*, 2010, **285**, 29319.
- 71 X. Zhang, L. Feng, S. Chinta, P. Singh, Y. Wang, J. K. Nunnery and R. A. Butcher, *Proc. Natl. Acad. Sci. U. S. A.*, 2015, **112**, 3955.
- 72 C. I. Bargmann, Chemosensation in *C. elegans*, in *Wormbook*, ed. The *C. elegans* Research Community, 2006, DOI: 10.1895/wormbook.1.123.1.
- 73 C. I. Bargmann, *Nature*, 2006, **444**, 295.
- 74 H. Jang, K. Kim, S. J. Neal, E. Macosko, D. Kim, R. A. Butcher, D. M. Zeiger, C. I. Bargmann and P. Sengupta, *Neuron*, 2012, **75**, 585.
- 75 E. Z. Macosko, N. Pokala, E. H. Feinberg, S. H. Chalasani, R. A. Butcher, J. Clardy and C. I. Bargmann, *Nature*, 2009, **458**, 1171.
- 76 J. Q. White and E. M. Jorgensen, *Neuron*, 2012, **75**, 593.
- 77 R. R. Zwaal, J. E. Mendel, P. W. Sternberg and R. H. Plasterk, *Genetics*, 1997, **145**, 715.
- 78 D. Park, I. O'Doherty, R. K. Somvanshi, A. Bethke, F. C. Schroeder, U. Kumar and D. L. Riddle, *Proc. Natl. Acad. Sci. U. S. A.*, 2012, **109**, 9917.
- 79 K. Kim, K. Sato, M. Shibuya, D. Zeiger, R. Butcher, J. Ragains, J. Clardy, K. Touhara and P. Sengupta, *Science*, 2009, **326**, 994.
- 80 R. V. O'Brien, R. W. Davis, C. Khosla and M. E. Hillenmeyer, *J. Antibiot.*, 2014, **67**, 89.

