

Ancestral and monophyletic presence of diplostigmaty in *Sebaea* (Gentianaceae) and its potential role as a morphological mixed mating strategy

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Summary

- Diplostigmaty, the presence of a primary (apical) stigma and secondary (mid-stylar) stigmas along the style, is only known from the genus *Sebaea* (Gentianaceae). Early work indicated that the secondary stigmas provide a mechanism of autogamy, suggesting that it might ensure reproductive assurance.
- Here, we test the monophyly of this unique morphological trait. Using Bayesian methods, we infer a nuclear DNA phylogeny for 96 accessions, including *c.* 50% of the species from the genus *Sebaea*. With this phylogeny, we infer the distribution of ancestral states on critical nodes using parsimony and likelihood methods.
- The inferred nrDNA phylogeny shows that the genus *Sebaea* is divided in two statistically well-supported clades, A and B, consistent with recent estimates. The most recent ancestor (MRCA) of clade A, except the most basal species (*Sebaea pusilla*), is resolved as diplostigmatic. No reversal to a single stigma is observed within this clade.
- We suggest that diplostigmaty is evolutionarily stable through time. We also discuss why this reproductive system is not found elsewhere than in Gentianaceae and the potential advantage of diplostigmaty as a stable mixed mating strategy.

Introduction

The majority of flowering plant species are hermaphroditic (i.e. flowers contain both male (stamens) and female (carpels) organs). Although most species fertilize their ovules with outcrossed pollen, self-pollination is widespread and the shift to self-pollination is a common transition in the evolutionary history of the angiosperms (Stebbins, 1974). Selfing relieves individuals from the requirement of having mating partners and allows rapid colonization, but it

is associated with costs, including inbreeding depression, pollen discounting and in the long term a loss of genetic variability (Barrett, 2002). Mixed mating, where an individual reproduces both by self-fertilization (selfing) and mating with genetically different individuals (outcrossing) occurs in at least one-third of all angiosperm species, as shown by inferences of selfing rates based on genetic markers (Barrett, 2002).

Early theoretical models suggested that inbreeding depression, the main selective factor opposing the evolution

of selfing, can be purged with self-fertilization, a process that favours pure strategies of either outcrossing or selfing (Lande & Schemske, 1985; Takebayashi & Morell, 2001; Goodwillie *et al.*, 2005; Johnston *et al.*, 2009). While further empirical and theoretical studies suggest that mixed mating strategies have evolved and can be evolutionarily stable (Charlesworth & Charlesworth, 1978; Gregorius, 1982; Holsinger, 1991; Haccou & Iwasa, 1995; Johnston *et al.*, 2009), the latter remains controversial (Lloyd, 1979; Lande & Schemske, 1985; Flaxman, 2000; Takebayashi & Morell, 2001; Herlihy & Eckert, 2002; Porcher & Lande, 2005; Schoen & Busch, 2008). Currently, the debate focuses primarily on factors that facilitate the maintenance of mixed mating systems, rather than on assessing whether, or which of the existing mixed mating systems are evolutionarily stable (Goodwillie *et al.*, 2005). It is therefore important to identify suitable study cases for such investigations that may provide examples of stable mixed mating systems.

Here, we investigate the ancestral and monophyletic presence of diplostigmy, a unique morphological trait. We discuss why it is found in Gentianaceae and not in other angiosperm families, and suggest experimental tests of the potential role of diplostigmy as a morphological mixed mating strategy.

Diplostigmy, the presence of a primary (apical) stigma and secondary (mid-stylar) stigmas on the style (Marloth, 1909; Fig. 1) is only known from the genus *Sebaea* (Gentianaceae). *Sebaea* is composed of *c.* 100 species (Schinz, 1906; Kissling *et al.*, 2008) mainly from Africa, of which *c.* 65 are diplostigmatic. The flowers are radially symmetrical (actinomorphic), with five free sepals and five petals that are united basally into a long tube with five free lobes. As a rule, the anthers are exerted and attached by a long filament to the sinus of the corolla lobes (Fig. 1a). Before anther dehiscence, the style head of the gynoeceum (with the apical stigma) is situated slightly below the level of the exerted stamens, while the secondary stigmas (when present) appear lower down on the style. When the anthers begin to open, the apical stigma is situated more or less at the same level as the anthers. The style continues its elongation and is mature when situated slightly above the anthers (avoiding self-pollen deposition on the apical stigma). The secondary stigmas are fully developed only much after the apical stigma reached its maturity, sometimes 2 or 3 d later, consistent with a potential role in reproductive assurance.

In 1913, Hill published a series of glasshouse experiments on several *Sebaea* species (i.e. *S. aurea*, *S. ambigua*, *S. confertiflora* and *S. imbricata*): he removed the apical stigma, the anthers or both. Hill also hand-pollinated the apical stigma, the secondary stigmas or both, and recorded seed set after these treatments. Removing the apical stigma led to significant enlargement of the secondary stigmas, and the plant produced viable seeds, while treatments involving removal of the anthers did not lead to any seed set, suggesting that

cross-fertilization was excluded under the experimental conditions. Together with hand-pollination results, Hill's work demonstrated that (1) these species are self-compatible, (2) the secondary stigmas are functional as an organ allowing self-pollen receipt and germination, and (3) seed set after removal or not of the apical stigma must have resulted from self-pollen deposition on the secondary stigmas. Both Hill's (1913) and Marloth's (1909) studies on *Sebaea* suggest that secondary stigmas allow the production of selfed progeny when cross-pollination (mediated by the primary, apical stigma) fails to fertilize some or all of the available ovules. This idea is also consistent with the relatively basal position of the secondary stigmas on the style, where self-pollen naturally falls (Fig. 1).

The presence of diplostigmy offers a unique opportunity to further test whether such a morphological feature allowing both self- and cross-fertilization as separate functions is stable through evolutionary time. If this mating system is not stable through time (i.e. over a very large number of generations) we should expect to observe reversals to a single stigma, assuming that secondary stigmas also entail some costs, and that owing to the unique stylar architecture (see the Discussion section) a reversal appears to be easy. Moreover, if, as suggested by theory, mixed mating could be advantageous, we should expect an individual selection advantage to diplostigmy (this being reflected at a large time-scale by stability and by diversification of the group with this innovation).

Here, we discuss this prediction by inferring the nrDNA phylogeny for an extended taxonomic sampling of *Sebaea* species and assessing the monophyly of diplostigmy within *Sebaea* by ancestral state inference.

Materials and Methods

Sampling, data collection and gene sequencing

Sebaea species were collected in Africa (from 2004 to 2007) and the sampling was maximized to cover the geographical, ecological and morphological variation of the genus. In addition, two representatives of the neighbouring tribe Chironieae were used as outgroups. The presence of secondary stigmas was recorded for all species sampled based on direct observation of living and herbarium materials.

When available, we used the previously published DNA sequence data of Exaceae (Yuan *et al.*, 2003, 2005). To these pre-existing data, we added 52 new sequences (GenBank Accession numbers FJ665987 to FJ666038) of *Sebaea* and *Exacum* species (Table S1). To obtain our new DNA sequences of nuclear ribosomal internal transcribed spacer (ITS1 and 2) and the 5.8S gene, we followed Yuan *et al.* (2003). To detect mistakes and correct uncertainties in the computer-generated sequence, we compared aligned tracefiles in CHROMASPRO version 1.33 (Technelysium Pty

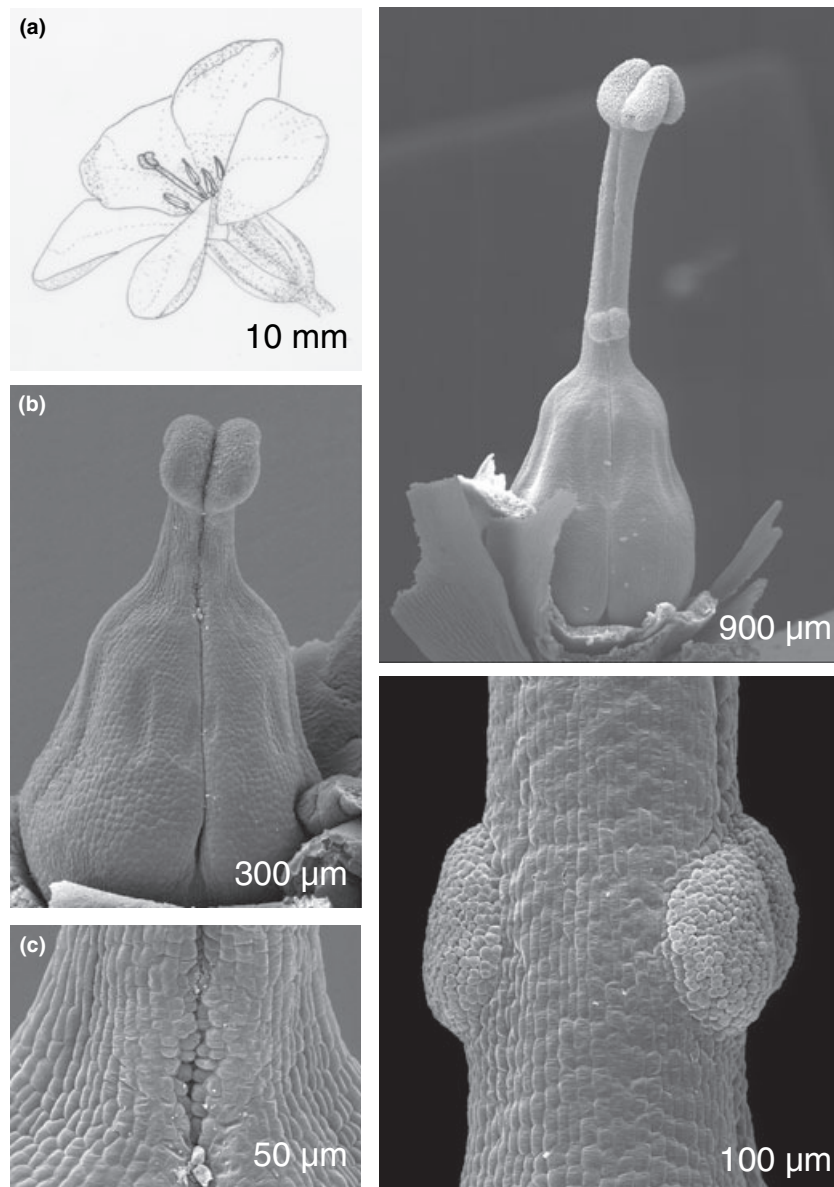


Fig. 1 *Sebaea macrophylla*. (a) Flower; (b) gynoecium at early stage, with carpels already postgenitally fused; (c) close up of part of (b) where the secondary stigmas will develop: the suture of postgenital fusion appears slightly expanded and epidermal papillae of the secondary stigmas are beginning to develop; (d) older gynoecium with secondary stigmas at the base of the style; (e) close up of part of (d), but in dorsal view of a carpel) with secondary stigmas on both sides.

Ltd). Alignment was performed using CLUSTAL W (Thompson *et al.*, 1994) as implemented in BIOEDIT 7.0.1 with subsequent manual improvement. The data matrix containing the aligned sequences is available on request from J.K or directly on TreeBase (preliminary number SN4561).

Phylogenetic analysis

Bayesian inference was conducted using MRBAYES version 3.1.2 (Huelsenbeck *et al.*, 2001; Ronquist & Huelsenbeck, 2003) and the data were partitioned by genes. The most

appropriate model of sequence evolution for each partition was determined using MRMODELTEST version 2.2. (Posada & Crandall, 1998; Nylander, 2004; Posada & Buckley, 2004) to be the general time reversible model (GTR + G) for the ITS1, and the GTR + I + G model for the ITS2, and the K80 model for the 5.8S gene. Default priors were used for the base frequency parameters. Two independent analyses each with four Markov chains, three heated and one cold, starting from a random tree were run simultaneously for 5 million generations with trees sampled every 1000 generations. We used the online program AWTY (Wilgenbusch *et al.*, 2004) to check for stationarity.

Trees generated before the four Markov chains reaching stationarity (the burn-in) were discarded. The remaining trees were used to construct a 50% majority rule consensus tree. High posterior probabilities were recovered for all major lineages.

Morphological character state optimization

The character state evolution (using data on absence/presence of the secondary stigmas) was reconstructed with MESQUITE 1.12 (Maddison & Maddison, 2008) on to the obtained Bayesian trees by using two methods: maximum-likelihood-based discrete Markov k-state 1 parameter models (Lewis, 2001; Maddison & Maddison, 2008); and a maximum parsimony approach (Maddison & Maddison, 2000). The likelihood-based Markov k-state 1 model does not consider any particular plesiomorphic state at the root of the tree, and a character state can change to any other state on any branch of the tree with equal probability. We report proportional likelihood (P.L.) values of states scaled so that the sum of all states is 1. We used a decision threshold of 2.0 in MESQUITE (Maddison & Maddison, 2008) for statistical considerations, while parsimony analyses considered character state transformations unordered.

Results

With both parsimony and likelihood methods, diplostigmaty is unequivocally inferred to have arisen once in *Sebaea* (Fig. 2), in clade A. Moreover, no reversal to a single stigma has occurred within this clade.

The Bayesian topology with posterior probabilities is given in Fig. 2 (arithmetic mean of the $-\log$ likelihood (L) of trees sampled after the burn-in = -7603.02 ; harmonic mean = -7683.94). The topology is in broad agreement with the relationships previously inferred in Exaceae using smaller taxonomic samplings (Yuan *et al.*, 2003, 2005; J. Kissling *et al.*, unpublished).

However, while the present phylogeny does not resolve the basal polytomy of the tribe, it confirms the recently described genus *Klackenbergia* (Kissling *et al.*, 2009) as distinct from *Sebaea*, and statistically supports the division of *Sebaea* into two main clades, hereafter called clade A and clade B, congruent with other estimates (J. Kissling *et al.*, unpublished) (Fig. 2).

Discussion

We used DNA sequence data from the nuclear genome to reconstruct the phylogeny of *Sebaea* and mapped the distribution of diplostigmaty character states, i.e. presence/absence of secondary stigmas, on the obtained phylogenetic tree. The data show that *Sebaea* is divided into two main, statistically well-supported, separate clades

(clades A and B). All members of clade A are diplostigmatic (i.e. have secondary stigmas), except for the basalmost species (*Sebaea pusilla*), indicating that no reversals (loss of the secondary stigmas) have occurred. By contrast, none of the species in clade B is diplostigmatic. The age of *Sebaea* has been previously estimated by penalized likelihood method to be *c.* 20.3–54.3 Myr (Yuan *et al.*, 2003), and this age has been recently confirmed for the diplostigmatic *Sebaea* clade (Kissling, 2007). As *Sebaea* species are small short-lived annuals with probably more than one generation per flowering season, we could reasonably assume *c.* 2×10^6 generations since the origin of diplostigmaty. This provides support for the idea that diplostigmaty has been stable through evolutionary time. However, it should also be taken into account that a character state with a lower net diversification rate could persist for a long time, even if it is not a key innovation.

Clade A represents a large group of *c.* 65 species, which occur mainly in the Western Cape (South Africa). This species diversity, compared with the other genera of Exaceae (including clade B), suggests a greater diversification for the diplostigmatic *Sebaea*. Furthermore the basalmost species of clade A, *S. pusilla*, has both a similar distribution area and a similar dispersal strategy compared with the rest of the diplostigmatic *Sebaea* (Kissling, 2007). Therefore, climatic or ecological factors alone are unlikely to explain the diversification burst of the rest of that clade. *Sebaea pusilla* also shares with the rest of the clade several reproductive characters such as bilobed stigma, mode of anther dehiscence, and flower colour and size. Although more work is needed to substantiate the correlates of this diversification this is consistent with the idea that the secondary stigmas, which are absent only from this basal species, might constitute an innovation that has contributed the diversification of clade A.

Persistence through time of diplostigmaty and diversification of clade A suggest that diplostigmaty may convey an advantage under certain ecological conditions. A plausible hypothesis for this, which is consistent with Hill's (1913) results but still requires experimental verification in natural populations, is that the presence of a secondary stigmas with delayed maturation might serve to ensure seed set when outcross pollen or pollinators are scarce (reproductive assurance; Jain, 1976), while at the same time the apical stigma can serve to promote opportunities for outcrossing during early anthesis. Thus the persistence through time of diplostigmaty is consistent with the idea that the ability to self if outcrossing fails is always a selective advantage unless failure to outcross is so rare that the cost of the structures to enable selfing exceeds their benefit. An important advantage of diplostigmaty with delayed maturation of the basal stigmas, which may enable its evolutionary stability, is that with this morphological trait the structures that facilitate selfing do not interfere with and do not diminish the opportunity for

outcrossing. Temporal and physical separation of primary and secondary stigmas may thus lower the risk of seed discounting (i.e. that production of selfed seed might use ovules that would have otherwise been outcrossed and more viable) (Barrett, 2002).

Because Hill's experiments indicate that removal of the apical stigma results in an enlargement of the secondary stigmas, and studies on other plants show that floral herbivory can influence the evolution of floral traits (e.g. Steets & Ashman, 2004; Ivey & Carr, 2005; Strauss & Whittall, 2006; Penet *et al.*, 2009), an alternative explanation for the putative ecological advantage of diplostigmaty in *Sebaea* could be that the secondary stigmas evolved because of high florivore pressure, rather than absence of pollinators. The 'surplus' stigma may function to replace the primary stigma lost to herbivores. In this scenario, mixed mating would be a byproduct of selection on diplostigmaty as an herbivore tolerance trait and not reproductive assurance *per se*. However, as we never found flowers with the primary stigmas destroyed neither in the field (> 200 populations) or in herbarium material (> 3000 collections), this scenario appears unlikely.

Whereas this may explain the persistence of this unique trait, it does not explain why secondary stigmas are not present in other groups. The answer may lie in unusual structural features in the development of the gynoecium (J. Kissling & P. K. Endress, unpublished). Although the gynoecium at anthesis appears as seemingly syncarpous (i.e. congenitally united) as in most angiosperms with a unified style, this is not really the case. In early development the two carpels are free at the level of the style and then fuse postgenitally (for the role of postgenital and congenital fusion, see Endress, 2006). Postgenital fusion of carpels by itself is unusual but not rare in angiosperms (Igersheim *et al.*, 2001; Matthews & Endress, 2005; Remizowa *et al.*, 2008). However, what makes it truly unique is that the free upper carpel parts that fuse postgenitally are not plicate (folded) as is usually the case, but are flat (Fig. 3) so that

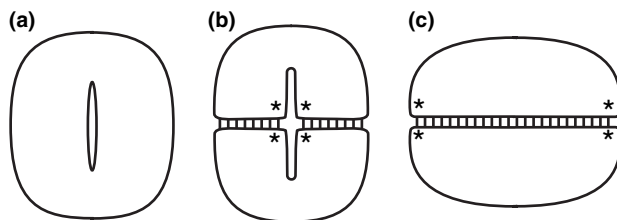


Fig. 3 Three different types of style of angiosperm gynoecia composed of two carpels (schematic transverse sections). (a) Type with the carpels congenitally united (the most common type in angiosperms); (b) type with plicate (folded) carpels postgenitally united (occurring in several families of eudicots and monocots); (c) type with flat carpels postgenitally united (occurring in *Sebaea*, some other Gentianaceae and a few Apocynaceae). Vertical lines between the two carpels, area of postgenital fusion; asterisks indicate carpel margins.

their margins are not hidden inside the style but come to lie at the surface of the style. Because, as a rule, the ventral side including the margins of a carpel develops stigmatic tissue, only a stylar architecture as in *Sebaea* has the potential to become stigmatic along the area of postgenital fusion (Fig. 1). Consistent with this, in *Sebaea* the secondary stigmas always develop along the postgenital fusion suture line between the carpels (Fig. 1c). The combination of these two features, postgenital fusion and flat carpels, is what makes this architecture unique. It is only known in Gentianaceae (McCoy, 1940; Baum, 1948; Leinfellner, 1951; Vijayaraghavan & Padmanaban, 1968) and the closely related Apocynaceae (Walker, 1975; Fallen, 1985). Among Gentianaceae, in *Bartonia* and *Lomatogonium* stigmatic tissue occurs all along the line of postgenital fusion of the two carpels. However, there is no separation into an upper and a lower stigma. In the majority of Apocynaceae the stigma is not apical but situated at the base of a 'style head'. However, their pollination apparatus is more complicated, and it is unlikely that the stigma is directly comparable with the secondary stigmas in *Sebaea*. In any case, the presence of two stigmas is only known in *Sebaea*.

In conclusion, diplostigmaty has arisen once in *Sebaea* clade A, and no reversal to a single stigma has occurred, consistent with the idea that the presence of a secondary stigmas provides an ecological advantage. To test this idea, further work should explore the evolutionary stability of diplostigmaty and whether, and to what extent, the secondary stigmas serve for reproductive assurance in natural populations, whereby the presence of this trait across a set of related species allows for replication of experimental tests and comparative analyses.

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References

- Barrett SCH. 2002. The evolution of plant sexual diversity. *Nature Reviews Genetics* 3: 274–284.
- Baum H. 1948. Die Stellung der Samenanlagen am Karpell bei *Asclepias syriaca*, *Cynanchum vincetoxicum* und *Erythraea centaurium*. *Österreichische Botanische Zeitschrift* 95: 251–256.
- Charlesworth D, Charlesworth B. 1978. Population-genetics of partial male-sterility and evolution of monoecy and dioecy. *Heredity* 41: 137–153.
- Endress PK. 2006. Angiosperm floral evolution: morphological developmental framework. *Advances in Botanical Research* 44: 1–61.
- Fallen ME. 1985. The gynoecial development and systematic position of *Allamanda* (Apocynaceae). *American Journal of Botany* 72: 572–579.
- Flaxman SM. 2000. The evolutionary stability of mixed strategies. *Trends in Ecology and Evolution* 15: 482–484.
- Goodwillie C, Kalisz S, Eckert CG. 2005. The evolutionary enigma of mixed mating systems in plants: occurrence, theoretical explanations, and empirical evidence. *Annual Review of Ecology, Evolution and Systematics* 36: 47–79.
- Gregorius HR. 1982. Selection in plant-populations of effectively infinite size. 2. Protectedness of a diallelic polymorphism. *Journal of Theoretical Biology* 96: 689–705.
- Haccou P, Iwasa Y. 1995. Optimal mixed strategies in stochastic environments. *Theoretical Population Biology* 47: 212–243.
- Herlihy CR, Eckert CG. 2002. Genetic cost of reproductive assurance in a self-fertilizing plant. *Nature* 416: 320–323.
- Hill AW. 1913. The floral morphology of the genus *Sebaea*. *Annals of Botany* 27: 479–489.
- Holsinger KE. 1991. Mass-action models of plant mating systems: the evolutionary stability of mixed mating systems. *The American Naturalist* 138: 606–622.
- Huelsenbeck JP, Ronquist F, Nielsen R, Bollback JP. 2001. Bayesian inference of phylogeny and its impact on evolutionary biology. *Science* 294: 2310–2314.
- Igersheim A, Buzgo M, Endress PK. 2001. Gynoecium diversity and systematics in basal monocots. *Botanical Journal of the Linnean Society* 136: 1–65.
- Ivey CT, Carr DE. 2005. Effects of herbivory and inbreeding on the pollinators and mating system of *Mimulus guttatus* (Phrymaceae). *American Journal of Botany* 92: 1641–1649.
- Jain SK. 1976. Evolution of inbreeding in plants. *Annual Review of Ecology and Systematics* 7: 469–495.
- Johnston MO, Porcher E, Cheptou P-O, Eckert CG, Elle E, Geber MA, Kalisz S, Kelly JK, Moeller DA, Vallejo-Marín ??? et al. 2009. Correlations among fertility components can maintain mixed mating in plants. *American Naturalist* 173: 1–11.
- Kissling J. 2007. Phylogenetics of tribe *Exaceae* (Gentianaceae) based on molecular, morphological and karyological data, with special emphasis on the genus *Sebaea* – taxonomic treatment of *Exochaenium*, *Lagenias* and the new genus *Klackenbergia*. PhD thesis, University of Neuchâtel, Neuchâtel, Switzerland.
- Kissling J, Zeltner L, Küpfer P, Mansion G. 2008. Cytogeography of Gentianaceae–Exaceae in Africa, with a special focus on *Sebaea*: the possible role of dysploidy and polyploidy in the evolution of the tribe. *Botanical Journal of the Linnean Society* 158: 556–566.
- Kissling J, Buerki S, Mansion G. 2009. *Klackenbergia* (Gentianaceae – Exaceae): a new endemic genus from Madagascar. *Taxon*, in press.
- Lande R, Schemske DW. 1985. The evolution of self-fertilization and inbreeding depression in plants. 1. Genetic models. *Evolution* 39: 24–40.
- Leinfellner W. 1951. Die Nachahmung der durch kongenitale Verwachsung entstandenen Formen des Gynözeums durch postgenitale Verschmelzungsvorgänge. *Österreichische Botanische Zeitschrift* 98: 403–411.
- Lewis PO. 2001. A likelihood approach to estimating phylogeny from discrete morphological character data. *Systematic Biology* 50: 913–925.
- Lloyd DG. 1979. Some reproductive factors affecting the selection of self-fertilization in plants. *American Naturalist* 113: 67–79.
- Maddison DR, Maddison WP. 2000. *MacClade 4: analysis of phylogeny and character evolution*. Sunderland, MA.
- Maddison DR, Maddison WP. 2008. *Mesquite: a modular system for evolutionary analysis, version 1.12*. <http://mesquiteproject.org/mesquite/mesquite.html>.
- Marloth R. 1909. A diplostigmatic plant, *Sebaea exacoides* (L.) Schinz (*Belmontia cordata* L.). *Transactions of the Royal Society of South Africa* 1: 311–314.
- Matthews ML, Endress PK. 2005. Comparative floral structure and systematics in Crossosomatales (Crossosomataceae, Stachyuraceae, Staphyleaceae, Aphloiaceae, Geissolomataceae, Ixerbaceae, Strasburgeriaceae). *Botanical Journal of the Linnean Society* 147: 1–46.
- McCoy W. 1940. Floral organogenesis in *Frasera carolinensis*. *American Journal of Botany* 27: 600–609.
- Nylander JAA. 2004. *MrModeltest v2*. Program distributed by the author. Evolutionary Biology Centre, Uppsala University, Sweden.
- Penet L, Collin CL, Ashman T-L. 2009. Florivory increases selfing: an experimental study in the wild strawberry, *Fragaria virginiana*. *Plant Biology* 11: 38–45.
- Porcher E, Lande R. 2005. The evolution of self-fertilization and inbreeding depression under pollen discounting and pollen limitation. *Journal of Evolutionary Biology* 18: 497–508.
- Posada D, Buckley TR. 2004. Model selection and model averaging in phylogenetics: advantages of the Akaike information criterion and Bayesian approaches over likelihood ratio tests. *Systematic Biology* 53: 793–808.
- Posada D, Crandall KA. 1998. Modeltest: testing the model of DNA substitution. *Bioinformatics* 14: 817–818.
- Remizowa M, Sokoloff D, Kondo K. 2008. Floral evolution in the monocot family Nartheciaceae (Dioscoreales): evidence from anatomy and development in *Metanartheceum luteo-viride* Maxim. *Botanical Journal of the Linnean Society* 158: 1–18.
- Ronquist F, Huelsenbeck JP. 2003. MrBayes 3: Bayesian phylogenetic inference under mixed models. *Bioinformatics* 19: 1572–1574.
- Schinz H. 1906. Beiträge zur Kenntniss der Afrikanischen Flora. Gentianaceae. *Bulletin de l'Herbier Boissier* 6: 714–746.
- Schoen DJ, Busch JW. 2008. On the evolution of self-fertilization in a metapopulation. *International Journal of Plant Sciences* 169: 119–127.
- Stebbins GL. 1974. *Flowering plants: evolution above the species level*. Cambridge, MA: Harvard University Press.
- Steets JA, Ashman T-L. 2004. Herbivory alters the expression of a mixed-mating system. *American Journal of Botany* 91: 1046–1051.
- Strauss SY, Whittall JB. 2006. Non-pollinator agents of selection on floral traits. In: Harder LD, Barrett SCH, eds. *The ecology and evolution of flowers*. Oxford, UK: Oxford University Press, 120–138.
- Takebayashi N, Morell PL. 2001. Is self-fertilization an evolutionary dead end? Revisiting an old hypothesis with genetic theories and a macroevolutionary approach. *American Journal of Botany* 88: 1143–1150.
- Thompson JD, Higgins DG, Gibson TJ. 1994. Clustal w: Improving the sensitivity of progressive multiple sequence alignment through sequence weighting, position-specific gap penalties and weight matrix choice. *Nucleic Acids Research* 22: 4673–4680.
- Vijayaraghavan MR, Padmanaban U. 1968. Morphology and embryology of *Centaurium ramosissimum* Druce and affinities of the family Gentianaceae. *Beiträge zur Biologie der Pflanzen* 46: 15–37.
- Walker DB. 1975. Postgenital carpel fusion in *Catharanthus roseus* (Apocynaceae). 1. Light and scanning electron-microscopic study of gynoecial ontogeny. *American Journal of Botany* 62: 457–467.
- Wilgenbusch JC, Warren DL, Swofford DL. 2004. AWTY: a system for graphical exploration of MCMC convergence in Bayesian phylogenetic inference. <http://ceb.csit.fsu.edu/awty>.

Yuan YM, Wohlhauser S, Moller M, Chassot P, Mansion G, Grant J, Küpfer P, Klackenberg J. 2003. Monophyly and relationships of the tribe Exaceae (Gentianaceae) inferred from nuclear ribosomal and chloroplast DNA sequences. *Molecular Phylogenetics and Evolution* 28: 500–517.

Yuan YM, Wohlhauser S, Moller M, Klackenberg J, Callmander M, Küpfer P. 2005. Phylogeny and biogeography of *Exacum* (Gentianaceae): a disjunctive distribution in the Indian Ocean basin resulted from long distance dispersal and extensive radiation. *Systematic Biology* 54: 21–34.

Table S1 Species of Exaceae and outgroups sampled for the present study (data are presented in the following order: Taxon, Voucher information, Origin, GenBank accession numbers)