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**Short Sequence-Paper**

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**Cloning and sequencing of a soybean nuclear gene coding  
for a chloroplast translation elongation factor EF-G**

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## Cloning and sequencing of a soybean nuclear gene coding for a chloroplast translation elongation factor EF-G

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Key words: Nuclear *fus* gene; Chloroplast; Translocation factor; EF-G; (Soybean)

A plant nuclear gene coding for a chloroplast specific translation elongation factor EF-G (cEF-G) was cloned and sequenced for the first time. We screened two partial soybean genomic libraries with a short PCR amplified pea DNA probe constructed according to the N-terminal peptide sequence of pea chloroplast EF-G. The gene is three times split, codes for a chloroplast type transit peptide and a protein very similar to bacterial translation elongation factor EF-G. The gene is expressed as evidenced by Northern hybridisations.

It was shown that chloroplast translation elongation factor EF-G activity of *Euglena gracilis* [1] and of pea [2] is light regulated. The expression of the cEF-G gene may be a key element in controlling light induced chloroplast protein synthesis and thereby chloroplast development. The corresponding gene (genes) is (are) located in the nucleus since no sequence homologous to the bacterial EF-G gene (*fus*) was yet detected in any of the completely sequenced chloroplast genomes [3–6]. However, no chloroplast specific nuclear EF-G gene was identified so far. We report here the cloning and sequencing of a soybean nuclear gene with a significant similarity to bacterial *fus* genes.

The N-terminal amino acid sequence of purified pea chloroplast EF-G was determined (Akkaya, M.S. et al., unpublished data) and accordingly short DNA probes were constructed using the PCR technique. Subsequently, we used one such DNA probe (156 bp) in Southern experiments with restricted soybean DNA to identify fragments potentially carrying *fus*-type sequences. In parallel experiments we also used DNA probes cut from the cloned *fus* gene of *Anacystis nidulans* (pAB46, kindly provided by Dr. Sugiura, Nagoya). *Eco*RI fragments in the range of 4 kb and *Bgl*II fragments in the range of 9 kb interacted with

both DNA probes, however, the signal was particularly strong with the pea probe. We then constructed two separate partial genomic libraries with purified *Eco*RI fragments (about 4 kb) and *Bgl*II fragments (about 9 kb) using as vector lambda GT10 and EMBL3, respectively.

In Fig. 1 we show the sequence of a complete nuclear gene coding for a chloroplast transit peptide and a protein having a high sequence identity with EF-G of *Escherichia coli* (59%) [7] and *A. nidulans* (55%) [8], but a definitely lower sequence identity with a yeast mitochondrial EF-G (41%) [9] and a *Chlorella* translocation factor EF-2 (27%) [10]. The gene is split three times by introns of 330, 508 and 288 bp, respectively, which all have the canonical 5'- and 3'-ends of plant nuclear introns (5'-AG.GU-) and an AU rich domain near the 3'-end [11].

In Fig. 2 the putative soybean cEF-G protein is aligned with EF-G proteins of *E. coli* and *A. nidulans* (see also next paragraph). The soybean protein shares extensive domains with the bacterial counterparts notably three GTP-binding sequences. From these and other (not shown) alignments we conclude that the given sequence represents a chloroplast specific EF-G protein.

The N-terminal peptide (85 amino acids, Fig. 1) has typical features of a chloroplast transit peptide [12,13]: (a) zero net charge for the N-terminal 10 amino acids, (b) alanine in second position, (c) a serine-rich central domain, (d) arginine at positions -7 and -8, (e) a





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Short communication

## Isolation and characterization of *gmsti*, a stress-inducible gene from soybean (*Glycine max*) coding for a protein belonging to the TPR (tetratricopeptide repeats) family

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**Key words:** *gmsti* gene, heat stress, tetratricopeptide, soybean, TRP motif

### Abstract

In close vicinity of two *fus* nuclear genes (chloroplast-specific translation elongation factor cEF-G) of soybean (*Glycine max*) we localized a split nuclear gene coding for a protein with tetratricopeptide repeats (TPR). A full-length cDNA was sequenced (1871 nucleotides). It encodes a protein (569 amino acids) with high sequence identity to the yeast STI1 stress-inducible and the human transformation-sensitive IEF SSP 3521 protein which both carry TPR elements. The soybean gene is heat-inducible. This is the first evidence for the existence of plant genes coding for proteins which belong to the TPR family. We call the gene *gmsti* and the protein GMSTI in analogy to the yeast counterpart.

While studying the structure and function of two soybean nuclear genes coding each for the chloroplast-specific translation elongation factor cEF-G (*fus*) [9, 10] we noticed in close vicinity downstream of both *fus* genes a reading frame with typical 34 amino acid repeats (tetratricopeptide repeat, TPR motif). The TPR motif is defined by a degenerate consensus sequence and is found as tandem repeats in diverse groups of proteins that function, for example, in mitosis, neurogenesis, transcription regulation and protein import [20].

Representatives of the TPR gene family are

found in yeast and other fungi, in cyanobacteria, in *Drosophila* (review in [20]) and in man [12] but not in plants as verified by computer-assisted search of EMBL data files. We screened a soybean cDNA library using a soybean genomic DNA fragment carrying a TPR sequence. We eventually retrieved and sequenced a cDNA clone predicted to encode a protein (GMSTI) with high sequence identity to the transformation-sensitive human protein IEF SSP 3521 [12] and to the stress-inducible yeast protein STI1 [13]. Both, the human and yeast protein contain several TPR motifs but the exact function of the two proteins

is unknown. This report presents the characterization of a soybean nuclear gene sequence which encodes a protein structurally related to the human and yeast stress-inducible proteins. The steady-state concentration of the *gmsti* mRNA sharply increases in soybean plants exposed to elevated temperature. We propose to name this novel soybean gene *gmsti* (*Glycine max sti*) in analogy to the yeast counterpart.

*Mapping and partial sequencing of two genome regions containing the gmsti gene*

A partial genomic *Bgl* II library (ca. 9 kb fragments) was specifically constructed in view of retrieving *fus* genes [9, 10]. Soybean (*Glycine max* cv. Ceresia [18]) was grown in a phytotron (Sanyo) at 25 °C, with a 12 h dark/12 h light photoperiod and at 50% relative humidity. Leaves were harvested after about two weeks of growth, immediately frozen in liquid N<sub>2</sub> and stored at -70 °C until use. DNA and RNA were isolated from 5 to 10 g of leaves according to standard procedures and as published [4]. Total soybean DNA was digested with *Bgl* II. DNA fragments were electrophoresed in a 0.6% agarose gel and fragments of about 9 kb were isolated by electroelution. About 500 ng of 9 kb DNA fragments were ligated at 4 °C for 48 h in a ligation mix containing 1 µg of *Bam* HI λEMBL3 arms (Stratagene), 50 mM Tris-HCl pH 7.5, 10 mM MgCl<sub>2</sub>, 10 mM DTT, 1.5 mM ATP and 1 U of T4 DNA ligase (BRL) in a total volume of 10 µl. *In vitro* packaging reaction was done using Promega's Packagene Lambda DNA Packaging System. Library screening was carried out following standard protocols [16] using as host *Escherichia coli* strain ER 1648 [8] (gift from Biolabs). Sequencing was done as described [17] or according to the protocol for sequencing double-stranded DNA preparations using Sequenase (U.S. Biochemical) or *Taq* polymerase (*fmol* DNA sequencing kit, Promega). DNA sequence data were analysed using the Sequence Analysis Software Package from the Genetics Computer Group (GCG) [5].

Sequencing of regions downstream of *fus* revealed reading frames with TPR motifs (FASTA search, EMBL data files). Figure 1 shows a short genomic sequence containing, respectively, the 3'-terminal part of *gmsti1* with at least one TPR element, the 3'-terminal part of *fus1* and the non-coding part between the two genes of opposite polarity. A similar study with a second genomic clone (*fus2*) gave a similar result (not shown), however, the distance between the 3' ends of the two genes of opposite polarity (*gmsti2*--*fus2*) is about 1000 nucleotides longer as determined by PCR analysis and partial sequencing. Indeed, gene 1 has a deletion in the non-coding 3' part of *fus1* eliminating the polyadenylation site (J. Hernández Torres and E. Stutz, to be published).

*A gmsti cDNA sequence coding for a protein with TPR motifs*

A genomic *gmsti1* DNA probe radiolabelled by random priming [6] was used to screen a lambda Uni-ZAP XR soybean cDNA library constructed with mRNA isolated from 12-day epicotyls of soybean cv. Williams. We followed the instructions of the manufacturer (Stratagene) using the host strain *E. coli* XL1-Blue [2]. Positive pBlue-script phagemids were excised *in vivo* from the lambda Uni-ZAP XR vector using the host strain SOLR (Stratagene). Several positive clones were retrieved and a full-length ORF was eventually sequenced coding for a protein structurally related to the human IEF SSP 3521 and yeast STI1 stress-inducible protein. In Fig. 2 the soybean GMSTI protein sequence derived from the cDNA sequence is aligned with the human and yeast counterparts. The soybean protein which is composed of 569 amino acids with a predicted molecular size of about 63 550 Da shares several conserved domains with the human and yeast counterparts, especially in the N- and C-terminal region. One notices, for example, a perfectly conserved β-sheet-forming [3] decamer, NHVLYS-NRSA, which is situated at the end of the first TRP motif (positions 35 to 44). Overall sequence identities between the soybean GMSTI protein

```

1 gatatcccttttcttttagGCTTATACTAACAGAGCTGCATGCTATACAAAACCTGGGGCAATGCCCGAAGGCTTAAAAGATGCAGAGAAATGCATTGAG 100
  A Y S N R A A C Y T K L G A M P E G L K D A E K C I E
101 CTTGATCCAACCTTTTCAAGGCTTATACTAGCAAAGCTGCAGTGCAGTTTCCATGAAAGAATATGACAAAGCTTTGGAAACATACAGGAGGCTTTGA 200
  L D P T F S K G Y T R K G A V Q P S H K E Y D K A L E T Y R E G L K
201 AACATGATCCTAACAAATCAGAAATGCTTATGATGCATAGCAAGGtatggatcgtgctacattcctgctatTTTTGGTTgaattatgccactttccata 300
  H D P N N Q E L L D G I R R
301 tgcacatgacaatcaagtgcctaatacaccatagatatacattcagATGCTGTAGAACAAATTAATAAGGCTAGCCCTGGAGATTTTACACCTGAGGAG 400
  C V E Q I N K A S R C D F T P E E
401 TTAAGGAGAGACAGtggtgacttgttctgataatcttgaactctattgctaatacctgaaatggcaattcattgtgcaaatattaatTTATg 500
  L K E R Q
501 ttttgcagGCCAAGCAATGCAAGACCCAGAGATACAAAGCAATTCGCAAGACCCCTGTTATGACACAAgtagctgctccttggattgaaagtactgt 600
  A K A M Q D P E I Q S I L Q D P V M T Q
601 gttgtgtcaatgtaatagaggaactagctgtgctaacagagagttggcatgtatgcagGTATTGACTGATTTCCAGGAAAATCCCAAGCTGCAGAGG 700
  V L T D F Q E N P R A A E E
701 AACATGTGAAGAATCCAATGCTGATGACAAAGACTCAAAACCTGACAGTCCCGGATGCCAGATGAGATGAATTTGATGGAAGTGGCAATCAAGCAGAG 800
  H V K N P M V N N K T Q N V T V P G C Q M R *
801 AAACAATGACAATGCTATTTTCTGCTGTAACATATATAATTTAAATAGATTGGATTGAAAAAAGAGAGGAGAAATTTGCTATTTGGAGATTTTCTCTGG 900
  ↓
901 CTCCTGCAATCGTTTCTTCTGGAATTCATATATTTGAGCATGTCAAATGCAGTCCATTTATACCTTTTACTGGTCTTCTGCAATGCTGCTCATTCG 1000
  ↑↑
1001 TGTGACTTTTCTGTTGAGTTAAATTCATATAGTACAAGGAAAATTAACAATAAAAAAATACCAAAAAACAACCAACTATTTGACAAATGGACATAATACAA 1100
  ↑↑
1101 CTATCAAAGGTAAGAACTTCTGTTAGGACCAAGCAAGTACATTAAGCGCAACTTCTGCTCTTTGGTGGCAAGTGGTTCTGAATGCTGCTGAGGC 1200
  * A A V E Q E K T A L Q N Q I H Q P
  +---+

```

Fig. 1. Genomic sequence showing the 3'-terminal region of soybean (*Glycine max* cv. Ceresia) *gmst1* and *fus1*, respectively. The amino acid sequence of four exons of GMST1 protein and of the C-terminal region of the chloroplast specific translation elongation factor cEF-G [9] are shown. A TPR motif within the topmost exon is underlined. Introns are in small letters; ↓ position of polyadenylation site in *gmst1*; ↑↑ site of deletion in *fus1*; ←←← transcription polarity of *fus1*.

and the human and yeast counterparts are 44% and 38% respectively (GCG program GAP).

Structure and function of proteins belonging to the TRP family have been discussed in detail [1, 7, 11, 19, 20]. Sikorski *et al.* [20] compared TRP elements from different proteins and defined three classes of repeats with distinct consensus sequences. According to their data the yeast STI1 TPR elements belong together with those of the mitochondrial MAS70 protein to class III with a consensus pattern given on the bottom line of Fig. 3. In Fig. 3 soybean TPR elements are aligned with the corresponding elements of yeast STI1 and human IEF SSP 3521 protein. We find that the soybean repeats follow a consensus pat-

tern with highly conserved positions: 1 (A), 8 (G), 20 (A), 24 (Y), 32 (P). The soybean TPR elements of block I and III show a good matching score with class III (13 out of 26 consensus positions) but the other soybean repeats in blocks II, IV and V show no preference for any of the three classes. Sikorski *et al.* [20] discuss the possibility that different classes of TPR repeats may coexist within a single protein.

#### Gmst1 gene is heat-inducible

The human IEF SSP 3521 and the yeast STI1 protein accumulate under stress [12, 13]. We



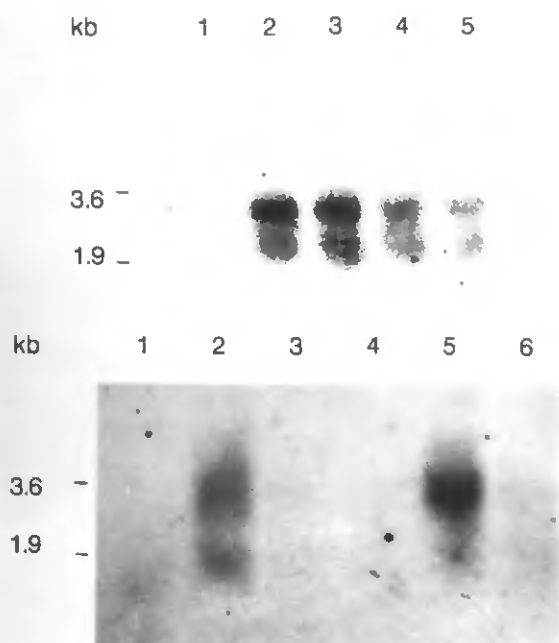


Fig. 4. Northern blot analysis of total soybean RNA. 20  $\mu$ g of total RNA isolated from leaves of plants grown under different temperature conditions were loaded on each lane. Blots were probed with a 1.1 kb single-strand  $^{32}$ P-labelled *gmst1* cDNA probe [14]. A (top) 1, control, 25 °C; 2 to 5, 30 min, 1, 2, 3 h, respectively, at 42 °C. B. 1, control; 25 °C; 2, 30 min, 42 °C; 3, re-exposed for 30 min, 25 °C; 4, re-exposed for 1 h, 25 °C; 5, re-exposed a second time for 30 min, 42 °C; 6, re-exposed a second time for 30 min, 25 °C.

for 30 min (lane 3) and 1 h (lane 4) at 25 °C. The two major broad bands interacting with *gmst1* DNA disappear almost completely. Bringing plants back to 42 °C for 30 min induces again the synthesis of *gmst1* mRNA (lane 5) and after 30 min at 25 °C the strong signals become very faint again. Obviously the steady-state concentration of *gmst1* mRNA is very low in plants grown at 25 °C (essentially below detectability under the given experimental conditions) but its concentration sharply increases under heat stress. At least one of the two *gmst1* genes is transcribed under non-stress conditions as documented by the presence of a full-length cDNA clone.

We have not yet isolated and characterized a putative GMST1 protein. It should have a MW in the range of 63 350 and a pI of 6.07. A soybean heat shock nuclear gene of about equal size (*hsp70*) was characterized [15] but it does not

have any structural relationship to the GMST1 protein. Currently we do not know the function of GMST1 protein but it may have functions similar to the yeast ST11 protein which seems to play a role in mediating the heat shock response to some HSP70 proteins. According to Nicolet and Craig [13] yeast ST11 is essential for cells growing under non-optimal conditions. Plants are exposed to several kinds of stress. The GMST1 protein may be an essential component required for activating the complex stress response as described [21].

Multiple TPR elements arranged in tandem have the capacity to form helix-turn domains and could be assembled into filamentous structures [11]. The function of TPR repeats, however, is not understood. It is tempting to speculate that the multiple repeats in GMST1 and in its yeast and human counterparts are functionally important. Otherwise it would be hard to understand why these repeats are so well conserved in yeast, plants and man.

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Analysis of the nuclear genes (*fus*) coding for the  
chloroplast specific translation elongation  
factor cEF-G in soybean (*Glycine max*)

de M. Jorge Hernandez Torres

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UNIVERSITÉ DE NEUCHÂTEL  
FACULTÉ DES SCIENCES

La Faculté des sciences de l'Université de  
Neuchâtel sur le rapport des membres du jury,

MM. E. Stutz (directeur de thèse),  
P. Schürmann et A. Boschetti (Berne)

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